

**Turkish Journal of Botany** 

http://journals.tubitak.gov.tr/botany/

Turk J Bot (2021) 45: 601-612 © TÜBİTAK doi:10.3906/bot-2109-38

### **Invited Review Article**

### The trade-off between UVB sensitivity and tolerance against other stresses in African rice species

Gideon Sadikiel MMBANDO, Jun HIDEMA\*

Graduate School of Life Sciences, Tohoku University, Sendai 980-8577, Japan

<b>Received:</b> 23.09.2021 • Accept	ted/Published Online: 29.11.2021 •	Final Version: 28.12.2021
--------------------------------------	------------------------------------	---------------------------

Abstract: Climate change and ozone depletion have caused increased ultraviolet B (UVB; 280-315 nm) radiation levels on earth, which has been predicted to cause decreased growth and yield of various crops, including rice, the most important staple food worldwide. UVB radiation sensitivity varies extensively in Asian rice (Oryza sativa L.) and African rice (Oryza glaberrima Steud. and Oryza barthii A. Chev.) cultivars, and the activity of cyclobutane pyrimidine dimer (CPD) photolyase that repairs UVB-induced CPD is an essential factor underlying UVB sensitivity in rice. Unlike Asian varieties, African rice possesses different origins, and it is cultivated in the African continent under environmental stresses and unstable climatic conditions and is well-adapted to various biotic and abiotic stresses. However, information regarding UVB sensitivity in African rice remains lacking. In this review, we describe recent research examining the sensitivity of rice species to UVB radiation, and we focus on UVB sensitivity and CPD photolyase genotypes with emphasis on African rice species. Consequently, the novel CPD photolyase genotype found only in African rice species results in a more severe phenotype termed "Super-hypersensitive" in cultivars grown only in West Africa, particularly at O. glaberrima domestication centres. We also describe possible reasons for the high UVB sensitivity of African rice cultivars in relation to plant morphology and other environmental stresses such as floods and pathogens. Finally, using the available knowledge, we suggest possible ways to develop multiple stress-resistant plants that can cope well in tropical environments under numerous environmental stresses. This review provides more tools for increasing food production for feeding the outgrowing population, particularly in tropical areas such as Africa.

Key words: African Rice, CPD photolyase, environmental multiple stresses, trade-off, UVB radiation, resistance

### 1. Introduction

As sessile organisms, plants are exposed to various biotic and abiotic stress conditions that affect their growth, development, and reproduction. Abiotic stresses include drought, heat, cold, salinity, ultraviolet (UV) radiation, and others, while biotic stresses are typically caused by phytophagous insects, bacterial pathogens, nematodes, and fungi. To counter the effects of these stresses, plants have evolved multiple mechanisms that help them to sense and acclimate to these environmental stresses (Gupta et al., 2015), and these mechanisms include shedding of affected tissue, stress neutralisation, renewal of tissue growth, and damage repair.

Sunlight is crucial for supporting life on our planet, but it consists of UV radiation. Photosynthetic organisms require sunlight and are, therefore, exposed to UV radiation. The UV region of the spectrum is divided into three portions based on wavelength, and these include UVA (315-400 nm), UVB (280-315 nm), and UVC (less than 280 nm). The shortest UV waves are the most detrimental to living organisms. Although the ozone layer

completely absorbs UVC radiation, this type of radiation is the most hazardous. In contrast, UVA cannot be absorbed by the ozone layer and, thus, is transmitted to the surface of the earth; however, it is less harmful than the other wavelengths of UV radiation (Hollosy, 2002). Although it contributes only a minor proportion (1.5%) (Cockell and Horneck, 2001; Horneck et al., 2010) of the total radiation among the three types of UV radiation, UVB is of high importance due to its ability to severely damage DNA and, thus, affect plant growth and development. In the past, there has been worldwide concern regarding the impact of increased UVB radiation on the surface of the earth and the organisms living there due to the destruction of the stratospheric ozone layer. Although the Montreal Protocol has prevented a significant increase in UVB radiation, as of 2020, we are now observing an unprecedented increase in UVB levels (Neale et al., 2020) in the Arctic due to stratospheric ozone depletion.

As they depend on sunlight for photosynthesis throughout their entire lives, plants are constantly exposed to harmful sunlight UVB radiation that suppresses

<sup>\*</sup> Correspondence: jun.hidema.e8@tohoku.ac.jp



photosynthesis and protein synthesis, ultimately leading to a decrease in growth and productivity (Teramura et al., 1990, 1991; Kumagai et al., 2001; Hidema et al., 2005). Rice is one of the most important staple grains worldwide and is cultivated globally, including Africa where the amount of UVB radiation from sunlight is high (Jablonski and Chaplin, 2010). Cultivated rice belongs to the genus Oryza (AA diploid genome group). In this genus, there are only two cultivated rice species with two taxonomically distinct species, including Oryza glaberrima (O. glaberrima) and Oryza sativa (O. sativa), that were separated independently approximately 640,000 years ago. O. sativa was originated in Asia, while O. glaberrima was in West Africa (Wang et al., 2014; Meyer et al., 2016). Although O. glaberrima is cultivated in wetland conditions in Africa in an unstable environment and under poor traditional farming system, it has developed a number of useful traits to resist various biotic and abiotic stresses (Second, 1985; Brar and Khush, 1997; Sanchez et al., 2013). Rice is an important food in Africa and is not only a staple food but also highly appreciated for its taste, culinary qualities, and ritual values. However, it is cultivated under multiple environmental stresses such as flooding, drought, harsh temperatures, pests, pathogens, and UVB radiation, all of which may affect its productivity. Thus, understanding the possible interaction mechanisms between one stress such as UVB radiation and other stresses such as floods and pathogens will be of great importance in providing methods to develop multiple resistance crop plants possessing high yields and productivity.

UVB radiation exerts deleterious effects on plants primarily by inhibiting photosynthesis through the reduction of the expression of key photosynthesis genes. In response to UVB irradiation, the D1 and D2 (waterplastoquinone oxidoreductase complex) proteins of photosystem II in the chloroplast thylakoid are quickly degraded, and chlorophyll and the activity and amount of Rubisco are decreased (Mackerness, 1997; Jansen et al., 1998; Mackerness et al., 1999). UVB irradiation of plants increases hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) levels, damages the thylakoid membranes, and destroys chlorophyll and carotenoids (Karpinski et al., 1997; Takeuchi et al., 2003). Additionally, DNA is sensitive to UVB radiation, and, thus, absorption of UVB causes severe DNA lesions. The two UVB-induced DNA lesions are cyclobutane pyrimidine dimers (CPDs) and pyrimidine (6-4) pyrimidone photoproducts ([6-4] photoproducts) (Britt, 1996) that interfere with both DNA replication and transcription. UVB can also affect plants through photooxidation or the formation of reactive oxygen species (ROS) and free radicals that are produced during photosensitization (Caldwell, 1993; Foyer et al., 1994). Among all of these mechanisms, CPDs result in the majority of the DNA damage (approximately 75%),

while the (6-4) photoproducts account for the remainder (Mitchell and Nairn, 1989). Therefore, CPDs are the main cause of growth inhibition in rice plants (Hidema et al., 2007), and such damage may be lethal to various organisms (Brash et al., 1987) and may even cause cancer in humans (Brash et al., 1991). These damages can interfere with transcription and replication, and this is believed to be the cause of the adverse effects observed in plants. To cope with such damage, plants use two main mechanisms for DNA repair of CPDs and (6-4) photoproducts that are termed "photorepair (photoreactivation)" and "nucleotide excision repair (dark repair)". In photorepair, the enzyme photolyase binds to the CPDs or to 6-4 photoproducts, and using photon energy from UVA to blue light (350-550 nm), it monomerizes or reverses the dimers in a lesion-specific manner, ultimately releasing DNA to its native state (Sancar, 1994). Conversely, for dark repair the photoproducts are removed from DNA through nucleotide excision repair (NER) processes that involve multiple complex steps to facilitate damage recognition, chromatin remodelling, excision of the damaged oligonucleotide, gap-filling DNA synthesis, and strand ligation within the correct order. Thus, in higher plants, photorepair is believed to be the major and most effective pathway for repairing UV-induced DNA lesions (Pang and Hays, 1991; Quaite et al., 1994; Hidema et al., 1997), and this is likely due to the slower nature of the dark repair process compared to that of photorepair (Britt, 1999) and the knowledge that this process is highly energy-consuming due to the involvement of numerous enzymes. Despite this, both photorepair and dark repair of DNA have been reported in various plant species (Hidema et al., 2000).

UVB sensitivity varies widely among Asian rice species, and this difference is primarily caused by the difference in CPD photolyase activity (Teranishi et al., 2004; Hidema et al., 2005, 2007; Iwamatsu et al., 2008). Therefore, CPD photolyase may act as an essential factor for plants to withstand UVB-induced DNA damage. O. sativa transgenic rice plants possessing CPD photolyase overexpression exhibited a significant increase in UVB resistance compared to that of wild-type plants, thus, suggesting the importance of this enzyme in UVB tolerance (Hidema et al., 2007; Teranishi et al., 2012). However, it remains unknown how African rice species survive the harmful effects of tropical UVB radiation. There is little knowledge regarding the UVB sensitivity and CPD photolyase genotypes of African cultivated rice species in relation to those of Asian rice. It is of interest to determine if there any possible relationship between African rice morphology and UVB sensitivity. Additionally, it is of interest to determine if UVB sensitivity result can form a trade-off with other environmental stresses in African rice species and the UVB-sensitive Asian rice Surjamkhi.

In this review, first, we describe the UVB sensitivity and CPD photolyase genotypes of African rice compared to those of Asian rice. We also explain the morphological and geographical adaptation differences between the two species that led to various UVB-sensitive African rice cultivars and possibly the crosstalk between UVB sensitivity and other environmental stresses. Next, we describe the potential applications of the available resources of Asian rice cultivars to develop UV-resistant African rice cultivars that may exhibit a higher yield under tropical environmental conditions with high levels of UVB radiation. Finally, using the available knowledge, we suggest possible ways to develop multiple stress resistant plants that can cope well in tropical environments under numerous environmental stresses. This review provides more tools for increasing food production for feeding the outgrowing population, particularly in tropical areas such as Africa.

### 2. Historical background

2.1 Variation of UVB sensitivity in African cultivated rice The activity of CPD photolyase is necessary for plants to survive harmful UVB irradiation stress (Britt et al., 1993). Our previous data demonstrated that UVB sensitivity varies widely among Asian rice cultivars. The evaluation of UVB sensitivity was divided into three groups according to the degree of UVB sensitivity: UVB-resistant, UVBsensitive, and UVB-hypersensitive. The differences in UVB sensitivity among Asian rice cultivars were primarily caused by different CPD photorepair activities due to different CPD photolyase genotypes that were caused by spontaneous polymorphisms within the CPD photolyase gene that altered both the structure and activity of the enzyme (Teranishi et al., 2004; Hidema et al., 2005; Ueda et al., 2005; Iwamatsu et al., 2008): the CPD photolyase activity is higher in the UVB-resistant rice than in the UVBsensitive rice, and the activity in UVB-hypersensitive rice is lower than that in the UV-sensitive rice. Interestingly, most of indica rice cultivated in Southeast Asia, which has a relatively high amount of UVB radiation, belongs to UVB-hypersensitive group (Sato and Kumagia, 1993; Hidema and Kumagai, 2006). This led us to investigate the UVB sensitivity of African rice cultivars.

Recently, Mmbando et al. (2020) examined the UVB sensitivity of 15 African rice cultivars belonging to *O. sativa*, *O. barthii*, and *O. glaberrima* that are cultivated in different geographical locations on the African continent in comparison to Asian rice UVB-resistant (*O. sativa ssp. japonica*, Sasanishiki), sensitive (*O. sativa ssp. japonica*, Norin 1), and hypersensitive (*O. sativa ssp. indica*, Surjamkhi) rice cultivars. Surprisingly, they observed that most of the African rice species examined in that study were highly sensitive to UVB radiation compared

to the sensitivity in Asian rice species, and certain cultivars exhibited even more severe damage phenotypes (UVB-super-hypersensitive) than did the hypersensitive Asian rice cultivar Surjamkhi (Figure 1A) (Hidema et al., 2005). Based on the UVB sensitivity index, they classified the UVB sensitivity of African rice into three groups that included UVB-sensitive that were similar to Asian rice (Norin 1), hypersensitive that were similar to the Asian rice Surjamkhi, and super-hypersensitive that was newly discovered in their study and was identified only in African rice species. There were no African rice cultivars with a higher UVB resistance index than that of Sasanishiki. Moreover, it was evident that the UVB-superhypersensitivity of African rice cultivars was caused by low total CPD photolyase activity due to both the low amount and activity of CPD photolyase. The reduction in the activity of CPD photolyase was due to new specific amino acid changes in the CPD photolyase gene of these cultivars that affected the structure and activity of the enzyme (Mmbando et al., 2020). This study was surprising, as we know that UVB radiation is higher in tropical areas such as Africa (Jablonski and Chaplin, 2010); however, most of the African rice species exhibited increased sensitivity to UVB radiation stress, and there were no UVB-resistant African rice species. This study suggests that although the newly identified polymorphisms reduce the total CPD photolyase activity of African rice, they may also play a significant role in surviving and adapting African rice in West Africa.

## 2.2 Crucial factors for determining UVB sensitivity in African rice cultivars.

It is of interest to determine the crucial factors controlling UVB sensitivity in African rice cultivars. The UVB-superhypersensitive African rice cultivars exhibited both a low amount and activity of CPD photolyase. The low CPD photolyase activity was due to the new polymorphisms that were conserved mainly in African rice species (Figure 1B). Additionally, the CPD photolyase content differed even among cultivars with the same genotype in both Asian and African rice (Mmbando et al., 2020). African rice is grown in a tropical environment with a higher amount of UVB radiation stress, and therefore, one should expect to possess a higher total CPD photolyase activity. Interestingly, the low total CPD photolyase activity was not only observed in UVB-super-hypersensitive African rice cultivars but also observed in the Asian rice Surjamkhi; thus, suggesting that the low total CPD photolyase activity among these cultivars contributed to a higher degree of sensitivity to UVB radiation (Mmbando et al., 2020).

There are a number of other UVB-sensitive Asian rice cultivars that belong to the *aus* ecotype from the Bengal region (Sato and Kumagai, 1993; Ueda et al., 2005; Hidema and Kumagai, 2006), and the major question is how these genotypes can survive the high UVB radiation with such a low total CPD photolyase activity. Mmbando et al. (2020) proposed that the higher sensitivity among these cultivars may be beneficial for surviving other environmental stresses such as those caused by pathogens or herbivores (Kunz et al., 2006; Kunz et al., 2008; Qi et al., 2018). Alternatively, they suggested that, unlike Asian rice, the origin of African rice may be deep-sea rice (paddy rice), and, therefore, it became more resistant to African flood conditions by evolving distinct adaptive morphological structures with high growth rates and long stems (Jackson and Ram, 2003; Sakagami et al., 2009; Sakagami, 2012). However, in contrast, it may not possess an adaptation mechanism for UVB, as ultraviolet radiation does not easily pass through the water. This hypothesis will be explained in detail in the last section. However, it is of interest to determine the CPD photolyase genotypes possessed by African rice species and if they possess a genotype similar to that of Asian rice species.

### 2.3 Novel CPD photolyase polymorphisms cause UVBsuper-hypersensitivity in African rice cultivars

We previously reported three CPD photolyase genotypes among cultivated and wild rice, and these genotypes were highly correlated with UVB sensitivity. One difference in the nucleotide adenine (A) at position 377 (exon 2) in Sasanishiki was changed to guanine (G) in Norin 1, and this causes amino acid changes at position 126 from glutamine (Q) to arginine (R). Moreover, the Sasanishiki and Surjamkhi CPD photolyase sequences differ by several positions (377, 888, 939, and 1248); however, it was the only alteration at positions 377 and 888 that led to a change. Specifically, the CAG codons at both positions 126 and 296 encoding Q in the Sasanishiki were changed to CGG and CAC codons in Surjamkhi that encode R and histidine (H), respectively (Teranishi et al., 2004; Ueda et al., 2005; Hidema et al., 2005). Thus, the UVB-resistant (Sasanishiki) at positions 126 and 296 were Q126-Q296 ("Sasa-type"), while those in UVB-sensitive (Norin 1) were R126-Q296 ("Nori-type") and UVB-hypersensitive (Surjamkhi) were R<sup>126</sup>- H<sup>296</sup> ("Sur-type") (Iwamatsu et al., 2008). Hidema et al. (2000) reported the mutation in Norin 1 compared to Sasanishiki caused by a decrease in the rate of binding to CPD through the use of photoflash analysis. Moreover, the QTL analysis test of the linkage between CPD photolyase and UVB sensitivity (Hidema et al., 2005) revealed that UVB sensitivity is a quantitative inherited trait. Furthermore, Ueda et al. reported that qUVR-10 exhibits the largest genetic differences among QTLs associated with UVB resistance and encodes CPD photolyase (Ueda et al., 2005). These differences in genotypes primarily affected CPD photolyase activity. Therefore, the differences in UVB sensitivity among Asian and wild cultivars are determined by the genotype of the CPD photolyase (Teranishi et al., 2004; Hidema et al., 2005; Iwamatsu et al., 2008). Based on this, it is of interest to further examine the CPD photolyase genotypes of African rice species.

The Sasa-type CPD photolyase genotype was not observed among all African rice cultivars examined in the study by Mmbando et al. (2020) and all cultivars possessed R<sup>126</sup> as in the Nori- and Sur-types. The Q<sup>296</sup> genotype, similar to the Sasa- and Nori-types, was identified in UVB-super-hypersensitive African rice (TOB7307, Jiakawo Wodewo, MB3, C7251, TOG12380, TOG14928, and Maro Goudo), while the H<sup>296</sup> genotype, as in the Surtype, was observed in all African tropical O. sativa (TOS) and only one tropical O. barthii (TOB14466) that were examined in that study. They also observed new amino acid substitutions in African cultivars that possessed a Q<sup>296</sup>. The cytosine at position 232 in exon 1 (Sasanishiki) was changed to thymine in UVB-super-hypersensitive African rice cultivars, thus, leading to an amino acid change at position P78 to S78. Additionally, guanine at position 848 in exon 4 (Sasanishiki) was changed to cytosine in the same cultivars and also in TOG12380, thus, leading to an amino acid change at position G<sup>283</sup> to A<sup>283</sup>. Of note, all African cultivars possessing the "S78-R126-A283-Q296" genotype with the exception of O. glaberrima (MB3), were classified as UVB-super-hypersensitive type. These results suggest that the "S78-R126-A283-Q296, genotype largely affects the UVB sensitivity of African rice species. Furthermore, the "S78-R126-A283-Q296" genotype was primarily found in UVB-super-hypersensitive African rice (O. glaberrima and O. barthii) and not in Ancestors or Asian rice species (Mmbando et al., 2020). These cultivars are also grown in the proposed domestication region of O. glaberrima in West Africa (Semon et al., 2005). Conversely, the countries of origin of all African O. sativa cultivars possessing genotypes identical to that of the Asian cultivar Surjamkhi (O. sativa ssp. indica) (P78-R126-G283-H296) were distributed widely throughout Africa and included countries proposed as the principal entry zone during the introduction of Asian O. sativa into West Africa. Asian O. sativa may have, thus, been introduced to various areas of Africa (including West Africa) by humans, perhaps during the period of the Atlantic slave trade (beginning ca. 1550) or earlier through trans-Saharan trade routes.

However, if these polymorphisms are advantageous for African rice species is yet to be determined, and it must be further explored if it is an adaptive response obtained during the domestication process. Nevertheless, this review raises the possibility of an interaction between UVB sensitivity and other environmental stresses, such as floods, pests and herbivores, among these species. These points require future clarification.



**Figure 1.** Variety of UVB sensitivity and CPD photolyase polymorphisms in African rice cultivars. (A) Asian (*Oryza sativa; japonica* rice Sasanishiki and Noron 1 and *indica* rice Surjamkhi) and African rice cultivars (Tropical *Oryza sativa;* TOS8086 and TOS10589, *O. barthii;* TOB7307, *O. glaberrima;* TOG12380, MB and Jiakawo W.) were grown in a growth cabinet for 21 days with (+UVB) or without (-UVB) UVB radiation. UVB sensitivity varied widely among Asian and African rice cultivars. Bars = 5 cm. (B) Cyclobutane pyrimidine dimers (CPD) photolyase genotype of African rice cultivars. The amino acid residues of CPD photolyase at the place where the mutation was identified, and its base sequence are shown. This figure is partially modified the figure published in Mmbando et al. (2020).

# 2.4 Growing conditions and morphological features that may influence the UVB sensitivity of African rice cultivars.

UVB radiation inhibits the growth and productivity of economically important crops such as rice by reducing cell division and elongation (Fina et al., 2017; Hopkins et al., 2002; Rajendiran and Ramanujam, 2003; Teramura, 1983). African rice cultivars are grown in the wetland of Africa in an unstable environment and under poor or traditional farming techniques with low human interference, and they are, therefore, prone to not only UVB radiation stress but also various stresses caused by climate fluctuations such as flooding (Sakagami and Kawano, 2011; Sakagami, 2012), drought, and temperature. As this rice has survived a harsh environment with low human interference, tolerance mechanisms have been developed to cope with various biotic and abiotic stresses (Second, 1985; Brar and Khush, 1997; Sanchez et al., 2013). Flooding has been demonstrated to be a significant abiotic stress affecting agricultural productivity of rice worldwide in areas with controlled irrigated conditions, deep-water rice, and rice in tidal wetlands, and these types of rice are well known to develop some morphological structures that enable them to survive such deep-water agriculture systems, including shoot and internodal elongation (Kende et al., 2002). African rice is known as a type of deep-water rice, and most of the African rice cultivars have developed submergence tolerance mechanisms (Sakagami and Kawano, 2011) and adapt well during flooding compared to these characteristics in Asian rice. Therefore, unlike Asian rice, the majority of African rice (O. glaberrima) cultivars have developed some structural and morphological adaptations as an escape strategy for surviving flooding conditions in West Africa (Jackson and Ram, 2003; Sakagami et al., 2009; Sakagami, 2012). Due to its taller structure and high potential for photosynthesis during flooding conditions (Sakagami et al., 2009), O. glaberrima is still preferred over O. sativa and is still cultivated today (Ndjiondjop et al., 2010) despite the knowledge that it possesses a lower productivity compared to that of Asian rice O. sativa (Oladele et al., 2016). However, during short-term submergence (flash flooding) caused by unstable climatic conditions, adaptation strategies for surviving flooding, including shoot and internode elongation, will make these species vulnerable to lodging and result in photoperiodic sensitivity (Sakagami and Kawano, 2011; Sakagami, 2012) and UVB sensitivity (Mmbando et al., 2020). Consequently, O. glaberrima differed in ligule shape, panicle branching, and tiller number formation (Portères, 1955; Besancon, 1993; Agnoun et al., 2012). Vigorous tillering, high leaf area index, and high specific leaf area of O. glaberrima contribute to its high competitiveness against weeds (WARDA, 1996; Rodenburg et al., 2009)

and flooding (Sakagami, 2012). However, rice plants possessing more tiller numbers and broader leaves may experience more UVB damage, as more tiller numbers and plant surface area will be exposed to UVB radiation. Therefore, it is reasonable to assume that the high tiller number, broad leaves, taller structure, longer shoot, and internode elongation will make O. glaberrima cultivars resistant to flooding but susceptible to UVB radiation. This could explain why most O. glaberrima cultivars exhibited super-hypersensitivity, while tropical O. sativa, an Asian rice that may have been introduced to Africa with low leaf area index, exhibited various UVB sensitivity phenotypes that were all less severe compared to that of O. glaberrima (Figure 1A) (Mmbando et al., 2020). Thus, it is possible that African rice species have evolved different plant shapes that expose them to different amounts of UVB radiation stress.

As African rice was cultivated under submergence conditions by individuals occupying the flood plains of the Niger River ~3,000 years ago (Portères, 1962; Portères and Harlan, 1976), O. glaberrima may not have perceived UVB radiation as the most serious stress from the environment due to the majority of its parts (shoot and leaves) typically growing underwater and thus experiencing less UVB damage. In this regard, UVB radiation may not penetrate deep water, and even if it penetrates, the intensity may be lower than the desubmergence growing condition. Consequently, O. glaberrima has grown mostly under the flooded environment in West Africa and has evolved predominantly unique plant features such as shoot elongation, taller plants, and large leaf area (Porteres, 1955; Besancon, 1993; Sakagami, 2012) to better survive in weeds and flooded environments. However, these features make it vulnerable to other stresses such as UVB radiation (Mmbando et al., 2020). Furthermore, it remains to be clarified if the submergence growing conditions of O. glaberrima affect its metabolic rate in a manner that results in UVB-super-hypersensitivity. Moreover, it has yet to be determined if some African rice cultivars grown in the upland system under desubmergence conditions have developed a UVB tolerance mechanism similar to that of the Asian rice O. sativa. The relationship between UVB sensitivity and flood mechanisms of African rice cultivars and yield productivity under field conditions will be an exciting topic to explore in the future. Recently, Mmbando et al. (2020) developed UVB-tolerant African rice with transgenic technology that may perform well under the tropical conditions of Africa with high UVB radiation stress, and, thus, this rice holds a promising future for the African continent in regard to increasing rice yield production. However, if transgenic UVB-resistant African rice with high CPD photolyase activity will perform well under field conditions in Africa with multiple biotic and abiotic stresses has yet to be determined.

## 2.5 A possible reason for UVB-super-hypersensitivity in African rice species.

Due to their sessile nature, plants are exposed to various environmental stresses, including not only UVB radiation but also cold, heat, high light, drought, salinity, and others that may affect their growth and productivity. It has been demonstrated that exposure of plants to ambient UVB radiation can enhance the resistance to other stresses such as pathogens, and the capacity of the plant to repair DNA damage may be a crucial factor in determining the types of responses elicited by different fluence rates (Kalbin et al., 2001; Kunz et al., 2006; Kunz et al., 2008; Kobayashi et al., 2014; Piofczyk et al., 2015; Robson et al., 2015; Parada et al., 2015; Qi et al., 2018; Demkura and Ballaré, 2012). Kunz et al. (2008) reported that UV-induced DNA damage could activate the defence mechanism against Hyaloperonospora parasitica. African rice cultivars are grown in a tropical environment under multiple stresses and are well adapted to various biotic and abiotic stresses such as drought, soil acidity, iron, and, aluminium toxicity (Second, 1985; Brar and Khush, 1997; Sanchez et al., 2013). Mmbando et al. (2020) demonstrated that most O. glaberrima were UVB-super-hypersensitive compared to the sensitivity of the Asian rice O. sativa, and cultivars with this genotype were primarily identified in West Africa in locations that were believed to be domestication centres for O. glaberrima. In that study, they speculated that such a high sensitivity to UVB radiation could represent a positive adaptation response obtained by O. glaberrima during the domestication process in order to survive multiple stresses present on the African continent. Indeed, UV radiation has been proposed to act on downstream signalling pathways in elements that closely resemble pathogen defence, including pathways involving reactive oxygen species (Mackerness et al., 2001), calcium (Frohnmeyer et al., 1999), mitogen-activated protein kinase (MAPK) (Ulm et al., 2001), ethylene, jasmonic acid, and salicylic acid (Brosché and Strid, 2003), thus suggesting its involvement in the defence signalling pathway. Therefore, it is possible that the high CPD levels in UVB-super-hypersensitive African rice cultivars with lower CPD photolyase activity induce high defence responses or systemic defence responses that enable them to survive tropical climates that are prone to pathogens. As plants encounter multiple environmental stresses under normal growing conditions, future studies should examine the yield of UVB-superhypersensitive African rice cultivars grown under normal field conditions. Furthermore, it has been reported that even without UV irradiation, a triple Arabidopsis mutant that is defective in NER and also photoreactivation of CPDs with tt5 mutation conferring chalcone isomerase deficiency (Li et al., 1993) exhibited resistance to infection by H. parasitica (Kunz et al., 2008). The tt5 mutant

exhibited a deficiency in flavonoid production and was hypersensitive to UV radiation (Li et al., 1993), and thus, it was sensitive to reactive oxygen species, exhibited a high incidence of DNA strand breaks, and possessed a high increase in recombination frequency compared to that in wild-type plants (Filkowski et al., 2004). Therefore, it is intriguing to suggest that in addition to UV photoproducts, accumulation of other endogenous DNA damage caused by reactive oxygen species could also trigger resistance to pathogens (Kunz et al., 2006). These points will be an important topic for future research.

## 2.6 Development of a multiple resistance African rice plant

UVB radiation and pathogens may compromise the yield and productivity of rice plants. CPD photolyase has been overexpressed in transgenic rice plants, thus enhancing UVB resistance compared to that of wild-type plants (Hidema et al., 2007; Teranishi et al., 2012). This suggests the necessity of this enzyme to enhance productivity. However, under normal field conditions, particularly in tropical areas such as Africa, UVB radiation is not the only stress affecting rice productivity. Other stresses such as temperature, drought, and pathogens also reduce the yield. Such stresses reduce the productivity of various crops, including rice, and the same plant can be affected simultaneously by UVB radiation and other stresses. To date, environmental relevant UVB dose have been shown to enhance plants functions. For example, moderate doses of UVB were found to enhance the heat tolerance of cucumber (Teklemariam and Blake, 2003) and the cold tolerance of rhododendron (Chalker-Scott and Scott, 2004). Furthermore, several studies have shown the possible link between plant response to UVB and drought tolerance (Bandurska and Cieślak, 2013). Such effect of increase drought tolerance by UVB radiation was suggested to be through accumulation of foliar flavonoids and phenols (Ren et al., 2007), or via increasing proline synthesis and decreasing of stomata conductance (Poulson et al., 2006), or through photomorphogenic effects (Gitz and Liu-Gitz, 2003). Moreover, increase production of UVB-protective secondary metabolites has shown to affect plants colonization by herbivorous and arthropods (Mazza et al., 1999; Mewis et al., 2012). In addition, Hideg et al. (2013), demonstrated that a suitable UVB radiation can induce defensive mechanisms and reduce oxidative damage similar to (Fujibe et al., 2004) who indicated the production of active oxygen species (AOS)-scavenging enzymes by UVB and (Wang et al., 2007) on the reduction of efficacy of paraguat due to and important factors of the cross-resistance between oxidative stress and UVB. UVB radiation has been shown to inhibit the growth of plants through the downstream targeting of growthregulating factors by miRNA396 (Casadevall et al., 2013; Fina et al., 2017). In contrast, miRNA396 also modulates the innate immunity of fungi (Soto-Suárez et al., 2017). Therefore, fine-tuning miRNA369 may offer a possibility for generating both UVB- and pathogen-resistant plants. Alternatively, UVB photoproducts in UV photorepair mutants trigger resistance to certain pathogens, and there is a possible connection between CPD accumulation and increased pathogen resistance (Li et al., 1993; Kucera et al., 2003; Kunz et al., 2008). Thus, downstream control of UVB-induced growth inhibition that allows for a high accumulation of CPDs or other UV-induced DNA photoproducts without causing growth inhibition may also allow for priming of the defence process among these plants.

Breeding both UVB- and pathogen-resistant cultivars is challenging, and a high immune resistance level often results in yield penalties (Brown, 2002; Brown and Rant, 2013). To minimise the fitness cost, a UVB-resistant plant that can turn off its immune system in the absence of pathogens and then reactivate it in the presence of pathogens should be developed. Although plants lack a heat-shock factor-like transcription factor that specifically binds to the cis-element translocon 1 (TL1: GAAGAAGAA) needed for the activation of antimicrobial genes (Pajerowska-Mukhtar et al., 2012), engineering plants with TL1-binding factor, TBF1, may offer promising ways to develop multiple resistance plant with low fitness cost. Recently, transcription factor (TF), TBF1 has been demonstrated to reduce the fitness penalties associated with enhanced disease resistance in Arabidopsis and rice (Xu et al., 2017). TBF1 is well established to play an essential role in the growth-to-defence switch upon immune induction (Pajerowska-Mukhtar et al., 2012), and, therefore, TF TBF1 analysis in the context of CPD photolyase overexpressing plants may provide a promising means to develop a UVB- and disease-resistant primed plant. Future studies should explore the possibility of creating multiple resistance cultivars by targeting either miRNA 396, TF TBF1, or other factors and by regulating the amount of CPDs or the rate of oxidative damage. Climate change and ozone depletion will continue to affect most tropical areas, including Africa. The high amount of UVB radiation and virulence of pathogens will also evolve due to unstable weather conditions. Consequently, developing a UVB-tolerant and disease-resistant primed rice plant will be of enormous importance, particularly in Africa, as this is a tropical continent with a high level of UVB radiation in the sunlight and a location subject to pathogen infection and that possesses a rapidly growing population. UVB-resistant transgenic primed plants will not only minimise the use of chemicals for combating various pathogens that are detrimental to our health and environment, but they will also increase the yield of this economically important crop that can eliminate hunger and poverty and feed the rapidly growing population of Africa.

### 3. Conclusion

African rice, O. glaberrima, is well adapted for cultivation in West Africa and is tolerant to biotic and abiotic stresses such as drought, soil acidity, and iron and aluminium toxicity. African rice, thus, has mechanisms to adapt to African climates and to protect itself against biotic and abiotic stresses. On the contrary, comparing the UVB sensitivity of rice cultivars with its cultivated region, very surprisingly, highly UVB sensitive cultivars (UVBhypersensitive or -super-hypersensitive cultivars) with low CPD photolyase activity have been domesticated in tropical areas, such as South Asia or African continent, where the amount of UVB radiation is relatively high although UVBresistant cultivars are expected to be cultivated in areas with high UVB radiation. Considering that highly UVBsensitive rice cultivars with low CPD photolyase activity have been selected and cultivated during the long history of rice cultivation, a possible crosslink might exist between CPD photolyase activity and other tropical environmental stresses, and high UVB-sensitivity may be a beneficial trait for surviving in tropical areas. Indeed, UVB radiation can activate the pathways of defensive mechanisms and increase resistance to fungal disease, through induction of pathogenesis related proteins (PRs) in leaves of several species.

Sunlight is essential for photosynthetic plants, but, because they are constantly exposed to UVB, the plants are damaged by UVB. With such constant UVB damage, a strategy to acquire a mechanism of tolerance to various other unexpected stresses may be rational. Consequently, plants are always faced with multiple stresses at the same time. Therefore, the understanding of the network interactions, including crosstalk with multiple signals caused by stresses, should be helpful for development of improved rice cultivars. The knowledge provided here will help to develop multiple resistant African rice possessing a high yield that can be used for feeding the outgrowing population of the African continent, thereby, ultimately reducing hunger and poverty.

### Acknowledgements

The research described in this review was supported by Grants-in-Aid for Scientific Research (KAKENHI) (no. 17H01872, and 20H04330 to J.H.). We would like to thank Dr. Marie Noelle Ndjiondjop (head of the Rice Biodiversity Center for Africa) for providing the African rice seeds used in this study.

#### References

- Agnoun Y, Biaou SSH, Sié M, Vodouhè RS, Ahanchédé A (2012). The African rice *Oryza glaberrima* Steud: Knowledge distribution and prospects. International Journal of Biology 4: 158-180. doi:10.5539/ijb.v4n3p158
- Bandurska H, Cieślak M (2013). The interactive effect of water deficit and UV-B radiation on salicylic acid accumulation in barley roots and leaves. Environmental and Experimental Botany 94: 9-18. doi: 10.1016/j.envexpbot.2012.03.001
- Brar DS, Khush GS (1997). Alien introgression in rice. Plant Molecular Biology 35: 35-47. doi: 10.1023/A:1005825519998
- Brash DE, Seetharamt S, Kraemertt KH, Seidmant MM, Bredbergt A (1987). Photoproduct frequency is not the major determinant of UV base substitution hot spots or cold spots in human cells (UV carcinogenesis/DNA structure/xeroderma pigmentosum/DNA repair/shuttle vector). Proceedings of the National Academy of Sciences of United States of America 84: 3782-3786. doi: 10.1073/pnas.84.11.3782
- Brash DE, Rudolph JA, Simon JA, Lin A, McKenna GJ et al. (1991). A role for sunlight in skin cancer: UV-induced p53 mutations in squamous cell carcinoma. Proceedings of the National Academy of Sciences of United States of America 88: 10124-10128. doi: 10.1073/pnas.88.22.10124
- Britt AB (1996). DNA damage and repair in plants. Annual Reviews of Plant Biology 47: 75-100. doi: 10.1146/annurev. arplant.47.1.75
- Britt AB (1999). Molecular genetics of DNA repair in higher plants. Trends in Plant Science 4: 20-25. doi: 10.1016/S1360-1385(98)01355-7
- Britt AB, Chen JJ, Wykoff D, Mitchell D (1993). A UV-sensitive mutant of *Arabidopsis* defective in the repair of pyrimidinepyrimidinone(6-4) dimers. Science. 261: 1571-1574. doi: 10.1126/science.8372351
- Brosché M, Strid Å (2003). Molecular events following perception of ultraviolet-B radiation by plants. Physiologia Plantarum. 117: 1-10. doi: 10.1034/j.1399-3054.2003.1170101.x
- Brown JKM (2002). Yield penalties of disease resistance in crops. Current Opinion Plant Biology 5: 339-344. doi: 10.1016/ S1369-5266(02)00270-4
- Brown JKM, Rant JC (2013). Fitness costs and trade-offs of disease resistance and their consequences for breeding arable crops. Plant Pathology 62: 83-95. doi: 10.1111/ppa.12163
- Caldwell CR (1993). Ultraviolet-induced photodegradation of cucumber (*Cucumis sativus* L.) microsomal and soluble protein tryptophanyl residues *in vitro*. Plant Physiology. 101: 947-953. doi: 10.1104/pp.101.3.947
- Casadevall R, Rodriguez RE, Debernardi JM, Palatnik JF, Casati P (2013). Repression of growth regulating factors by the microRNA396 inhibits cell proliferation by UV-B radiation in *Arabidopsis* leaves. Plant Cell 25: 3570-3583. doi: 10.1105/tpc.113.117473

- Chalker-Scott L, Scott JD (2004). Elevated ultraviolet-B radiation induces cross-protection to cold in leaves of Rhododendron under field conditions. Photochemical and Photobiology 79: 199-204. doi: 10.1562/0031-8655(2004)079<0199:eurict>2.0. co;2
- Cockell CS, Horneck G (2001). The history of the UV Radiation climate of the earth—theoretical and space-based observations. Photochemical and Photobiology 73: 447-451. doi: 10.1562/0031-8655(2001)073<0447:thotur>2.0.co;2.
- Demkura PV, Ballaré CL (2012). UVR8 mediates UV-B-induced arabidopsis defense responses against *Botrytis cinerea* by controlling sinapate accumulation. Molecular Plant 5: 642-652. doi: 10.1093/mp/sss025
- Filkowski J, Kovalchuk O, Kovalchuk I (2004). Genome stability of vtc1, *tt4*, and *tt5 Arabidopsis thaliana* mutants impaired in protection against oxidative stress. Plant Journal 38: 60-69. doi: 10.1093/mp/sss025
- Fina J, Casadevall R, AbdElgawad H, Prinsen E, Markakis MN et al. (2017). UV-B inhibits leaf growth through changes in growth regulating factors and gibberellin levels. Plant Physiology 174: 1110-1126. doi: 10.1104/pp.17.00365
- Foyer CH, Lelandais M, Kunert KJ (1994). Photooxidative stress in plants. Physiologia Plantarum 92: 696-717. doi: 10.1111/ j.1399-3054.1994.tb03042.x
- Frohnmeyer H, Loyall L, Blatt MR, Grabov A (1999). Millisecond UV-B irradiation evokes prolonged elevation of cytosolicfree Ca<sup>2+</sup> and stimulates gene expression in transgenic parsley cell cultures. Plant Journal. 20: 109-117. doi: 10.1046/j.1365-313x.1999.00584.x
- Fujibe T, Saji H, Arakawa K, Yabe N, Takeuchi Y et al. (2004). A methyl viologen-resistant mutant of *Arabidopsis*, which is allelic to ozone-sensitive *rcd1*, is tolerant to supplemental ultraviolet-B irradiation. Plant Physiology 134: 275-85. doi: 10.1104/pp.103.033480
- Gitz DC, Liu-Gitz L (2003). How do UV photomorphogenic responses confer water stress tolerance?. Photochemistry and Photobiology 78: 529-534. doi: 10.1562/0031-8655(2003)0780529HDUPRC2.0.CO2
- Gupta DK, Palma JM, Corpas FJ (2015). In Gupta DK, Palma JM, Corpas FJ (editors) Reactive oxygen species and oxidative damage in plants under stress. Springer, Heidelberg, pp. 1-22. doi:10.1007/978-3-319-20421-5
- Hidema J, Kang H, Kumagai T (1996). Differences in the sensitivity to UVB radiation of two cultivars of rice (*Oryza sativa* L .). Plant & Cell Physiology 37: 742-747. doi: 10.1093/oxfordjournals. pcp.a029008
- Hidema J, Kumagai T, Sutherland JC, Sutherland BM (1997). Ultraviolet B-sensitive rice cultivar deficient in cyclobutyl pyrimidine dimer repair. Plant Physiology 113: 39-44. doi: 10.1104/pp.113.1.39

- Hidema J, Kumagai T, Sutherland BM (2000). UV radiation-sensitive norin 1 rice contains defective cyclobutane pyrimidine dimer photolyase. Plant Cell 12: 1569-78. doi: 10.1105/tpc.12.9.1569
- Hidema J, Song IK, Sato T, Kumagai T (2001). Relationship between ultraviolet-B sensitivity and cyclobutane pyrimidine dimer photorepair in rice. Journal of Radiation Research 42: 295-303. doi: 10.1269/jrr.42.295
- Hidema J, Teranishi M, Iwamatsu Y, Hirouchi T, Ueda T et al. (2005). Spontaneously occurring mutations in the cyclobutane pyrimidine dimer photolyase gene cause different sensitivities to ultraviolet-B in rice. Plant Journal 43: 57-67. doi: 10.1111/j.1365-313X.2005.02428.x
- Hidema J, Kumagai T (2006). Sensitivity of rice to ultraviolet-B radiation. Annals of Botany 97: 933-942. doi: 10.1093/aob/ mcl044
- Hidema J, Taguchi T, Ono T, Teranishi M, Yamamoto K et al. (2007). Increase in CPD photolyase activity functions effectively to prevent growth inhibition caused by UVB radiation. Plant Journal 50: 70-79. doi: 10.1111/j.1365-313X.2007.03041.x
- Hollosy F (2002). Evaluation of lipophilicity and antitumour activity of parallel carboxamide libraries. Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences 780: 355-363. doi: 10.1016/s1570-0232(02)00545-7.
- Hopkins L, Bond MA, Tobin AK (2002). Ultraviolet-B radiation reduces the rates of cell division and elongation in the primary leaf of wheat (*Triticum aestivum* L. cv Marls Huntsman). Plant, Cell & Environment 25: 617-624. doi: 10.1046/j.1365-3040.2002.00834.x
- Horneck G, Klaus DM, Mancinelli RL (2010). Space microbiology. Microbiology Molecular Biology Review 74: 121-156. doi: 10.1128/MMBR.00016-09
- Iwamatsu Y, Aoki C, Takahashi M, Teranishi M, Ding Y et al. (2008). UVB sensitivity and cyclobutane pyrimidine dimer (CPD) photolyase genotypes in cultivated and wild rice species. Photochemical & Photobiology Sciences 7: 311-320. doi: 10.1039/B719034D
- Jablonski NG, Chaplin G (2010). Human skin pigmentation as an adaptation to UV radiation. Proceedings of the National Academy of Sciences of United States of America 107: 8962-8968. doi: 10.1073/pnas.0914628107
- Jackson MB, Ram PC (2003). Physiological and molecular basis of susceptibility and tolerance of rice plants to complete submergence. Annals of Botany 91: 227-241. doi: 10.1093/aob/ mcf242
- Jansen MAK, Gaba V, Greenberg BM (1998). Higher plants and UV-B radiation: Balancing damage, repair and acclimation. Trends in Plant Science 3: 131-135. doi: 10.1016/S1360-1385(98)01215-1
- Kalbin G, Hidema J, Brosché M, Kumagai T, Bornman JF et al. (2001). UV-B-induced DNA damage and expression of defence genes under UV-B stress: Tissue-specific molecular marker analysis in leaves. Plant, Cell & Environment 24: 983-990. doi: 10.1046/j.1365-3040.2001.00748.x

- Karpinski S, Escobar C, Karpinska B, Creissen G, Mullineaux PM (1997). Photosynthetic electron transport regulates the expression of cytosolic ascorbate peroxidase genes in Arabidopsis during excess light stress. Plant Cell 9: 627-640. doi: 10.1105/tpc.9.4.627.
- Kende H, van der Knaap E, Cho H-T (2002). Deepwater Rice: A Model Plant to Study Stem Elongation. Plant Physiol. 118: 1105-1110.
- Kobayashi M, Kanto T, Fujikawa T, Yamada M, Ishiwata M et al. (2014). Supplemental UV radiation controls rose powdery mildew disease under the greenhouse conditions. Environmental Control in Biology 51: 157-163. doi: 10.2525/ ecb.51.157
- Kucera B, Leubner-Metzger G, Wellmann E (2003). Distinct ultraviolet-signaling pathways in bean leaves. DNA Damage is associated with  $\beta$ -1,3-glucanase gene induction, but not with flavonoid formation. Plant Physiology 133: 1445-1452. doi: 10.1104/pp.103.029520
- Kumagai T, Hidema J, Kang HS, Sato T (2001). Effects of supplemental UV-B radiation on the growth and yield of two cultivars of Japanese lowland rice (*Oryza sativa* L.) under the field in a cool rice-growing region of Japan. Agriculture Ecosystems & Environment Ecosyst. Environ. 83: 201-208. doi: 10.1016/ S0167-8809(00)00180-8
- Kunz BA, Cahill DM, Mohr PG, Osmond MJ, Vonarx EJ (2006). Plant responses to UV radiation and links to pathogen resistance. International Review of Cytology 255: 1-40. doi: 10.1016/ S0074-7696(06)55001-6
- Kunz BA, Dando PK, Grice DM, Mohr PG, Schenk PM et al. (2008). UV-induced DNA damage romotes resistance to the biotrophic pathogen *Hyaloperonospora parasitica* in Arabidopsis. Plant Physiology 148: 1021-1031. doi: 10.1104/pp.108.125435
- Li J, Ou-Lee TM, Raba R, Amundson RG, Last RL (1993). Arabidopsis flavonoid mutants are hypersensitive to UV-B irradiation. Plant Cell 5: 171-179. doi: 10.1105/tpc.5.2.171
- Mackerness SAH (1997). Ultraviolet-B effects on transcript levels for photosynthetic genes are not mediated through carbohydrate metabolism. Plant, Cell & Environment 20: 1431-1437. doi: 10.1046/j.1365-3040.1997.d01-39.x
- Mackerness SAH, Surplus SL, Blake P, John CF, Buchanan-Wollaston V et al. (1999). Ultraviolet-B-induced stress and changes in gene expression in *Arabidopsis thaliana*: role of signalling pathways controlled by jasmonic acid, ethylene and reactive oxygen species. Plant, Cell & Environment 22: 1413-1423. doi: 10.1046/j.1365-3040.1999.00499.x
- Mackerness SAH, John CF, Jordan B, Thomas B (2001). Early signaling components in ultraviolet-B responses: Distinct roles for different reactive oxygen species and nitric oxide. FEBS Letters 489: 237-242. doi: 10.1016/S0014-5793(01)02103-2
- Mazza CA, Zavala J, Scopel AL, Ballaré CL (1999). Perception of solar UVB radiation by phytophagous insects: behavioral responses and ecosystem implications. Proceedings of the National Academy of Sciences of United States of America 96: 980–985.

- Mewis I, Schreiner M, Nguyen CN, Krumbein A, Ulrichs C et al. (2012). UV-B irradiation changes specifically the secondary metabolite profile in broccoli sprouts: induced signaling overlaps with defense response to biotic stressors. Plant & Cell Physiology 53: 1546-1560. doi: 10.1093/pcp/pcs096
- Meyer RS, Choi YJ, Sanches M, Plessis A, Flowers JM et al. (2016). Domestication history and geographical adaptation inferred from a SNP map of African rice. Nature Genetics 48: 1083-1088. doi:10.1038/ng.3633
- Mitchell DL, Nairn RS (1989). The biology of the (6–4) photoproduct. Photochemistry and Photobiology 49: 805-819. doi: 10.1111/ j.1751-1097.1989.tb05578.x
- Mmbando GS, Teranishi M, Hidema J (2020). Very high sensitivity of African rice to artificial ultraviolet-B radiation caused by genotype and quantity of cyclobutane pyrimidine dimer photolyase. Scientific Reports10: 1-14. doi: https://doi. org/10.1038/s41598-020-59720-x
- Ndjiondjop MN, Manneh B, Cissoko M, Drame NK, Kakai RG et al. (2010). Drought resistance in an interspecific backcross population of rice (*Oryza* spp.) derived from the cross WAB56-104 (*O. sativa*)×CG14 (*O. glaberrima*). Plant Science 179: 364-373. doi: 10.1016/j.plantsci.2010.06.006
- Neale RE, Barnes PW, Robson TM, Neale PJ, Williamson CE et al. (2020) Environmental effects of stratospheric ozone depletion, UV radiation, and interactions with climate change: UNEP Environmental Effects Assessment Panel, Update 2020. Photochemical & Photobiological Sciences: 20: 1–67. https:// doi.org/10.1007/s43630-020-00001-x
- Oladele SO, Adegbaju MS, Awodun MA (2016). Harnessing the potentials of African rice (*Oryza glaberrima* Steud.) in the quest for self-sufficient and increased yield in sub-saharan Africa. Journal of Global and Ecology 4: 157-167. Website: https://www.ikprress.org/index.php/JOGAE/article/view/778
- Pajerowska-Mukhtar KM, Wang W, Tada Y, Oka N, Tucker CL et al. (2012). The HSF-like transcription factor TBF1 is a major molecular switch for plant growth-to-defense transition. Current Biology 22: 103-112. doi: 10.1016/j.cub.2011.12.015.
- Pang Q, Hays JB (1991). UV-B-inducible and temperature-sensitive photoreactivation of cyclobutane pyrimidine dimers in *Arabidopsis thaliana*. Plant Physiology 95: 536-543. doi: 10.1104/pp.95.2.536
- Parada RY, Mon-nai W, Ueno M, Kihara J, Arase S (2015). Red-lightinduced resistance to brown spot disease caused by *Bipolaris* oryzae in rice. Journal of Phytopathology 163: 116-123. doi: 10.1111/jph.12288
- Piofczyk T, Jeena G, Pecinka A (2015). *Arabidopsis thaliana* natural variation reveals connections between UV radiation stress and plant pathogen-like defense responses. Plant Physiology and Biochemistry 93: 34-43. doi: 10.1016/j.plaphy.2015.01.011.
- Porteres R (1955). History of the first samples of *Oryza glaberrima* collected from Africa. Journal of Tropical Agriculture and Appled Biology 2: 535-537.

- Portères R (1955). Historique sur les premiers échantillons d'Oryza glaberrima St. recueillis en Afrique. Journal d'agriculture traditionnelle et de botanique appliquée 2: 535-537. https:// www.persee.fr/doc/jatba\_0021-7662\_1955\_num\_2\_10\_2257
- Portères R (1962). Berceaux agricoles primaires sur le continent africain. Journal of African History 3: 195-210. http://www. jstor.org/stable/179739.
- Portères R, Harlan J (1976). In Harlan JR, de Wet JMJ, Stamlar ABL (editors) The origin of African plant domestication. ASA Review of eBooks 5: 120-123. doi: 10.2307/532430
- Poulson ME, Boeger MRT, Donahue RA (2006). Response of photosynthesis to high light and drought for *Arabidopsis thaliana* grown under a UV-B enhanced light regime. Photosynthetic Research 90: 79-90. doi: 10.1007/s11120-006-9116-2
- Qi J, Zhang M, Lu C, Hettenhausen C, Tan Q, Cao G et al. (2018). Ultraviolet-B enhances the resistance of multiple plant species to lepidopteran insect herbivory through the jasmonic acid pathway. Scientific Reports 8: 1-9. doi: 10.1038/s41598-017-18600-7
- Quaite FE, Sutherland JC, Sutherland BM (1994). Isolation of highmolecular-weight plant DNA for DNA damage quantitation: relative effects of solar 297 nm UVB and 365 nm radiation. Plant Molecular Biology 24: 475-483. doi: 10.1007/BF00024115
- Rajendiran K, Ramanujam MP (2003). Alleviation of ultraviolet-B radiation-induced growth inhibition of green gram by triadimefon. Biologia Plantarum 46: 621-624. doi: 10.1023/a:1024840301092
- Ren J, Dai W, Xuan Z, Yao Y, Korpelainen H et al. (2007). The effect of drought and enhanced UV-B radiation on the growth and physiological traits of two contrasting poplar species. Forest Ecology and Managment 239: 112-119. doi: 10.1016/j. foreco.2006.11.014
- Robson TM, Hartikainen SM, Aphalo PJ (2015). How does solar ultraviolet-B radiation improve drought tolerance of silver birch (*Betula pendula* Roth.) seedlings? Plant, Cell & Environment 38: 953-967. doi: 10.1111/pce.12405
- Rodenburg J, Saito K, Kakaï RG, Touré A, Mariko M et al. (2009). Weed competitiveness of the lowland rice varieties of NERICA in the southern Guinea Savanna. Field Crops Research 114: 411-418. doi:10.1016/j.fcr.2009.09.014
- Sakagami J-I, Joho Y, Ito O (2009). Contrasting physiological responses by cultivars of *Oryza sativa* and *O. glaberrima* to prolonged submergence. Annals. Botany 103: 171-180. doi: 10.1093/aob/ mcn201
- Sakagami J-I, Kawano N (2011). Survival of submerged rice in a floodprone region of West Africa. Tropics 20: 55-66. doi: 10.3759/ tropics.20.55
- Sakagami J-I (2012). Submergence tolerance of rice species, *Oryza glaberrima* Steudel. Applied Photosynthesis 17: 353-364. doi: 10.5772/26023
- Sancar A (1994). Mechanisms of DNA excision repair. Science. 266: 1954-1956. doi: 10.1126/science.7801120

- Sanchez PL, Wing RA, Brar DS (2013). The wild relative of rice: Genomes and Genomics. Genetics and genomics of rice Spronger, New York, pp. 9-25. doi: 10.1007/978-1-4614-7903-1\_2
- Sato T, Kumagai T (1993). Cultivar differences in resistance to the inhibitory effects of near-UV radiation among Asian ecotype and Japanese lowland and upland cultivars of rice (*Oryza* sativa L.). Japanese Journal of Breeding 43: 61-68. doi: 10.1270/ jsbbs1951.43.61
- Second G (1985). In Sharma AK, Sharma A (editors) A new insight into the genome differentiation in *Oryza* L. through isozymic studies. Advancees in Chromosome and Cell Genetics Oxford & IBH, New Delhi, pp. 45-78. https://agris.fao.org/agrissearch/search.do?recordID=US201301441111
- Semon M, Nielsen R, Jones MP, McCouch SR (2005). The population structure of African cultivated rice (*Oryza glaberrima* Steud.): evidence for elevated levels of linkage disequilibrium caused by admixture with *O. sativa* and ecological adaptation. Genetics 169: 1639-1647. doi: 10.1534/genetics.104.033175.
- Soto-Suárez M, Baldrich P, Weigel D, Rubio-Somoza I, San Segundo B (2017). The Arabidopsis miR396 mediates pathogen-associated molecular pattern-triggered immune responses against fungal pathogens. Scientific Reports 7: 1-14. doi:10.1038/srep44898
- Teklemariam T, Blake TJ (2003). Effects of UVB preconditioning on heat tolerance of cucumber (*Cucumis sativus* L.). Environmental and Experimental Botany 50: 169-182. doi: 10.1016/S0098-8472(03)00024-8
- Teramura AH (1983). Effects of ultraviolet-B radiation on the growth and yield of crop plants. Physiologia Plantrum 58: 415-427. doi: 10.1111/j.1399-3054.1983.tb04203.x
- Teramura AH, Sullivan JH, Lydon J (1990). Effects of UV-B radiation on soybean yield and seed quality: a 6-year field study. Physiologia Plantrum 80: 5-11. doi. https://doi. org/10.1111/j.1399-3054.1990.tb04367.x

- Teramura AH, Ziska LH, Sztein AE (1991). Changes in growth and photosynthetic capacity of rice with increased UV-B radiation. Physiologia Plantrum 83: 373–380. doi: https://doi. org/10.1111/j.1399-3054.1991.tb00108.x
- Teranishi M, Iwamatsu Y, Hidema J, Kumagai T (2004). Ultraviolet-B sensitivities in Japanese lowland rice cultivars: cyclobutane pyrimidine dimer photolyase activity and gene mutation. Plant & Cell Physiololgy 45: 1848-1856. doi: 10.1093/pcp/pch215
- Teranishi M, Taguchi T, Ono T, Hidema J (2012). Augmentation of CPD photolyase activity in japonica and indica rice increases their UVB resistance but still leaves the difference in their sensitivities. Photochemistry & Photobiology Sciences 11: 812-820. doi: 10.1039/c2pp05392f
- Ueda T, Sato T, Hidema J, Hirouchi T, Yamamoto K et al. (2005). qUVR-10, a major quantitative trait locus for ultraviolet-B resistance in rice, encodes cyclobutane pyrimidine dimer photolyase. Genetics 171: 1941-1950. doi: 10.1534/ genetics.105.044735
- Ulm R, Ichimura K, Mizoguchi T, Peck SC, Zhu T et al. (2001). Distinct regulation of salinity and genotoxic stress responses by Arabidopsis MAP kinase phosphatase 1. EMBO Journal 21: 6483-6493. doi: 10.1093/emboj/cdf646
- Wang M, Yu Y, Haberer G, Marri PR, Fan C et al. (2014). The genome sequence of African rice (*Oryza glaberrima*) and evidence for independent domestication. Nature Genetices 46: 982-988. doi: 10.1038/ng.3044
- Nwanze KF, Terry ER (1996). West Africa Rice Development Association (WARDA) Annual Report 1996. https://pdf.usaid. gov/pdf\_docs/Pnach999.pdf
- Xu G, Yuan M, Ai C, Liu L, Zhuang E et al. (2017). uORF-mediated translation allows engineered plant disease resistance without fitness costs. Nature 545: 491-494. doi: 10.1038/nature22372