

Chemical and biotransformation of Gelomulide F: a rare diterpene lactone

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Received: 22.01.2012

Chemical and biotransformations of a rare diterpene lactone, Gelomulide F (1), from the cytotoxic extract of leaves of *Suregada multiflora* were studied. Fermentation of compound 1 with *Sachromyces cerevisiae* transformed it to Gelomulide D (2) and E (3), whereas its treatment with 2N KOH yielded Gelomulide D (2). In addition, a novel compound (4) was obtained by its diastereo and chemoselective reduction with NaBH₄. All of these compounds were characterized on the basis of extensive ¹H-NMR, ¹³C-NMR, 2D NMR, and mass spectral analyses.

Key Words: Diterpene lactone, Gelomulide, *Suregada multiflora*, *Sachromyces cerevisiae*, diastereo and chemoselective reduction

Introduction

A series of rare diterpene lactones were isolated from the leaves of *Suregada multiflora*.¹ In continuation of our previous work to accomplish structural modifications in this rare class of diterpene lactones,² the present investigation was carried out on Gelomulide F (1). Both chemical and biotransformation methods were employed and structurally interesting analogs (2-4) were obtained.

Experimental

Column chromatography (CC): Merck silica gel 60 (70-230 mesh) TLC: Merck silica gel GF 254 (0.25 mm); detection by vanillin reagent. All reagents used were of analytical grades. Supermarket baker's yeast NATU

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(China) was used. M. pts.: a micromelting point apparatus (Yanaco MP-S3). $[\alpha]_D$: JASCO DIP-360 digital polarimeter. UV spectra: a Hitachi U 3200 spectrophotometer; λ_{max} (log ε) in nm. IR spectra: JASCO A-302 IR spectrophotometer; in CHCl₃ on KBr disks; in cm⁻¹. ¹H- and ¹³C-NMR spectra: on Bruker Avance: at 400/100 MHz; resp., δ in ppm referenced with respect to the residual solvent signal of CDCl₃, coupling constants J in Hz. Mass spectra: a double-focusing mass spectrometer (Varian MAT 311 A); high-resolution electron impact mass spectra (HR/EI-MS) with a Jeol HX 110 mass spectrometer; in m/z (rel. %).

Microbial transformation of Gelomulide F (1) by Saccharomyces cerevisiae

Saccharomyces cerevisiae (baker's yeast, 40 g) was added in portions to a stirred solution of sugar (100 g) in water (300 mL) in a conical flask at 35 °C. Gelomulide F (200 mg in 2 mL of DMF) was added after 4 h. The reaction was monitored by TLC. After 4 days, the incubation mixture was filtered and extracted with chloroform. The combined extracts were dried over anhydrous Na_2SO_4 , and the solvent was evaporated to give 321 mg of brown residue. The crude residue was chromatographed on a silica gel column and eluted with EtOAc/hexane to obtain the transformed metabolites **2** (38%) and **3** (22%).

β -Elimination of Gelomulide F (1) by KOH

Compound 1 (10 mg, 0.25 mmol) in THF (2 mL) was put in a 25-mL round-bottomed flask and 2 mL of 2 N KOH was added. The solution was stirred at room temperature. After 2 h, the reaction mixture was neutralized with aq. HCl and extracted with $CH_2 Cl_2$. The organic layer was washed with ammonium chloride, dried over magnesium sulfate, and evaporated under vacuum. The residue was recrystallized by acetone-hexane to obtain compound 2 (85%).

2-Ene-8 β ,14 β -epoxy-1-oxo-13,15-abiatene-16,12-olide (2)

Appearance: Colorless prism; R_f : 0.34 (20% acetone-hexane); mp: 229 °C; $[\alpha]_D$: +85° (CHCl₃; c 0.003); UV: (MeOH) λ_{max} (log ε) 223 nm (5.6).); IR: ν_{max} : 2954, 1746, 1665, 1438, 1363, 1249, 1090 cm⁻¹; EI-MS: m/z (rel. int. %.): 328 (M⁺, 52), 310 (16), 295 (12), 267 (32), 232 (27), 214 (27) 186 (33), 160 (20), 150 (89), 137 (70), 96 (49), 53 (100).; HREI-MS: 328.1648 (for C₂₀H₂₄O₄, 328.1674); ¹H- and ¹³C-NMR: Table.

6β -Acetoxy-2-ene-1-oxo- 8β , 14β -epoxy-13, 15-abiatene-16, 12-olide (3)

Appearance: Colorless needles; R_f: 0.27 (20% acetone-hexane); mp: 267 °C; $[\alpha]_D$: -13.3° (CHCl₃; c 0.003); UV: (MeOH) λ_{max} (log ε) 227 nm (6.3); IR: ν_{max} cm⁻¹: 2960, 1758, 1730, 1240, 1668, 1460, 1378 and 1025; EI-MS: m/z (rel. int.%): 386 (M⁺, 17), 326 (54), 308 (43), 293 (18), 283 (9), 265 (37), 230 (8), 212 (10), 163 (17), 137 (100), 105 (37); HREI-MS: m/z = 386.1542 (for C₂₅H₂₂O₄, 386.1518); ¹H- and ¹³C-NMR: Table.

Reduction of Gelomulide F (1) by $NaBH_4$

Compound 1 (20 mg, 0.5 mmol) was treated with $NaBH_4$ (3 mg, 0.8 mmol) in MeOH under stirring at room temperature for 1 h. The reaction was quenched by addition of aq. HCl. The reaction mixture was extracted

with diethylether, and the organic layer was washed with NH_4 Cl solution, dried over MgSO₄, and concentrated over a rotary evaporator. The white solid obtained showed only one product (4) on TLC.

	1		2		3		4	
	$\delta(\mathrm{H})$	$\delta(C)$	$\delta(\mathrm{H})$	$\delta(C)$	$\delta(\mathrm{H})$	$\delta(C)$	$\delta(\mathrm{H})$	$\delta(C)$
1		210.2		204.5		202.6	3.64 <i>t</i> (2.6)	72.0
2	2.38 <i>dd</i> (13.5, 3.4) 3.17 <i>dd</i> (13.5, 3.6)	39.5	5.8 <i>d</i> (10)	124.1	5.82 d (10)	122.6	1.92-2.2 m	30.3
3	5.02 <i>t</i> (3.5)	80.6	6.4 <i>d</i> (10)	155.0	6.31 <i>d</i> (10)	156.5	4.94 <i>t</i> (2.8)	78.5
6		20.1		21.0	5.13 <i>ddd</i> (11,4.9)	70.1		
9	2.68 d (7.3)	40.8	2.9 d (7.2)	39.8	2.66 d (7.1)	38.9	2.60 d (7.6)	41.6
12	4.84 <i>ddd</i> (13, 5.4, 2)	75.7	4.83 <i>ddd</i> (12.1, 5.4, 2.1)	74.8	4.79 m ($w_{1/2} = 19$)	76.1	4.56 m ($w_{1/2} = 16$)	73.2
13	-	154.6	-	156.4	-	153.1	2.90 m	37.2
14	3.8 <i>s</i>	56.0	3.7 s	56.4	3.8 s	55.6	3.00 s	59.0
15	-	128.1	-	128.7	-	129.4	2.90 m	37.2
16	-	173.8	-	171.2	-	173.2	-	
17	1.94 <i>d</i> (1.8)	8.7	1.95 d (2.0)	9.0	1.96 d (1.7)	8.81	1.40 <i>d</i> (6.6)	11.5
18	0.96 s	27.7	1.10 <i>s</i>	31.3	1.16 s	33.7	0.91(<i>s</i>)	28.1
19	1.20 s	22.1	1.11 s	22.7	1.30 s	22.5	0.94 (s)	22.3
20	1.30 s	16.8	1.31 s	18.8	1.37 s	19.2	0.83(<i>s</i>)	20.6
OCO <u>CH</u> 3	2.03 s	20.9			2.05 s	21.4	2.1 s	21.6
O <u>CO</u> CH ₃	_	170.0	-		-	170.1	-	169.9

Table. ¹H- and ¹³C-NMR data of compounds **1-4** in CDCl₃; δ in ppm (J in Hz).

3β -acetoxy- 1β -hydroxy- 8β , 14β -epoxy-abiatane-16, 12-olide (4)

Appearance: White powder; Yield: (73%); mp: 264 °C; $[\alpha]_D$: -18.3° (CHCl₃; c 0.003); UV: (MeOH) λ_{max} (log ε) 220 nm (6.1); IR: ν_{max} cm⁻¹: 3464, 2927, 1752, 1449, and 1028; EI-MS: m/z (rel. int. %): 392 (13), 336 (2), 319 (6), 285 (19), 241 (70), 135 (20), 55 (100); FAB-MS (+ve): 393; ¹H- and ¹³C-NMR: Table.

Results and discussion

Saccharomyces cerevisiae (baker's yeast) was selected due to its well known ability to reduce carbonyl groups. However, interestingly, Gelomulide F (1) was transformed into compound 2 via β -elimination and 3 through subsequent acetylation at C-6 (Figure 1). Comparison of all the physical and spectral data identified the transformed product as Gelomulide D (2) and E (3), which have already been isolated from the same plant.¹ Treatment of substrate 1 with ethanolic KOH also afforded Gelomulide D (2) through β -elimination of the Chemical and biotransformation of Gelomulide F: a rare..., H. Y. GONDAL, M. I. CHOUDHARY

3-OAc group. Hence, based on these transformations, it can be proposed that Gelomulide D (2) and E (3) are biogenetically derived from Gelomulide F (1), which is a major constituent of the leaf extract of S. multiflora.¹



Figure 1. Compounds 1-4.

Moreover, a very interesting compound (4) was obtained by chemo- and diastereoselective reduction of Gelomulide F (1) with NaBH₄ at room temperature (Figure 1). The EIMS spectrum of compound 4 exhibited the M⁺ at m/z 392 and FAB-MS (+ve) at m/z 393. In the IR spectrum, the presence of –OH (3578 cm⁻¹) and the absence of ketonic absorption indicated that the keto group at C-1 of Gelomulide F (1) was reduced be to a hydroxyl group. The ¹H-NMR spectrum of compound 4 (Table) showed an extra methine signal at δ 3.64 attributed to H-1. The ¹³C-NMR spectrum further confirmed this modification as a new methine carbon that appeared at δ 72.0 instead of carbonyl carbon resonating at δ 210.2 in Gelomulide F (1). The stereochemistry of the hydroxyl group at C-1 was deduced to be β -oriented based on the NOE interactions between H-1 (δ_H 3.64) and α -oriented Me-20 (δ_H 0.83) in the NOESY experiment (Figure 2). It is well documented that prediction of the predominant product of a metal hydride reduction depends on the space requirement.³ We envisioned, as the axial approach of the hydride is strongly inhibited by an axial 3-OAc group in Gelomulide F (1), that the hydride approaches from the less hindered equatorial side (Figure 3), and as a result the predominant product is kinetically controlled axial alcohol in compound **4**.

Another interesting selectivity was introduced by NaBH₄ as the result of conjugate reduction of the lactone ring. The singlet of C-14 proton, geminal to the epoxy group, was shifted upfield at δ 3.00, while Me-17 appeared at δ 1.4 (d, J = 6.6 Hz), suggesting that the double bond in the lactone ring was also reduced. NOE interactions between H-12 and H-15 indicated that Me-17 is β -oriented. A 2-proton multiplet at δ 2.90 was attributed to H-13 and H-15, which also showed HMBC interaction with C-14 and C-16 (Figure 2). All this spectral evidence supported the structure of the reduced product (4) as 3β -acetoxy- 1β -hydroxy- 8β , 14β -epoxy-abiatane-16, 12-olide. The product is plausibly formed by initial conjugate addition of hydride anion to β -carbon (C-13), followed by ketonization of the less stable enol product (Figure 3). Selective conjugate reduction of α , β -unsaturated carbonyl compounds is already reported, usually requiring efficient catalysts like rhodium amido complexes, indium(III) chloride, or Rh(PPh)₃Cl.⁴⁻⁶ α , β -Unsaturated esters can reduced to saturated analogs by NaBH₄ only if an additional electron withdrawing group is present at the α -position or under drastic conditions, 7^{-12} whereas reduction of α , β -unsaturated lactones is rarely reported.¹³ The current

report is, therefore, a significant addition for the purpose of achieving conjugate addition of α , β -unsaturated lactone in very mild conditions.





Figure 2. Key NOE and HMBC interactions in compound 4.

Figure 3. Plausible selectivity in reduction of compounds 1 to 4.

Acknowledgment

The authors are grateful to the HEC (Higher Education Commission Pakistan) for providing a research grant under the NURP program.

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