

**Research Article** 

# Characterization of a $Cu^{2+}$ -selective fluorescent probe derived from rhodamine B with 1,2,4-triazole as subunit and its application in cell imaging

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Abstract: A rhodamine B derivative containing 1,2,4-triazole as subunit was characterized as an "off–on" type Cu<sup>2+</sup>-selective fluorescent probe. It exhibited high selectivity and sensitivity for Cu<sup>2+</sup> in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES) and underwent ring opening. A prominent fluorescence enhancement at 570 nm was observed in the presence of Cu<sup>2+</sup> with the change in the absorption spectrum, and a 1:1 metal–ligand complex was formed. With the optimized experimental conditions, the probe exhibited a dynamic response range for Cu<sup>2+</sup> from 8.0 × 10<sup>-7</sup> to 7.5 × 10<sup>-6</sup> M with a detection limit of  $2.3 \times 10^{-7}$  M in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES). Its application in Cu<sup>2+</sup> imaging in living cells was also studied.

Key words: Fluorescent probe, rhodamine B, triazole, Cu<sup>2+</sup>

# 1. Introduction

The detection of heavy transition-metal ions has attracted a lot of interest recently.  $^{1-3}$  Among them, copper is an essential trace element in both plants and animals, including humans. Deficiency and excess of copper could cause serious imbalance of human body functions, which damage the human brain and multiple systems.  $^{4-7}$ Therefore, the development of methods for easy detection of  $Cu^{2+}$  is of great importance for the environment and human health. Compared with the conventional methods for detecting  $Cu^{2+}$ , such as atomic absorption spectrometry (AAS), inductively coupled plasma-atomic emission spectrometry (ICP-AES), and inductively coupled plasma-mass spectroscopy (ICP-MS), fluorescence spectroscopy displayed high selectivity and sensitivity, was easy to operate, and had low detection limits. In addition, the equipment of detection was simple without complex multistage sample preparation.<sup>8-12</sup>

The property of the probes was determined by the fluorophore and recognition site. It is well known that rhodamine B was always chosen as fluorophore because of its unique structural characteristics and photophysical properties, that is, it appeared colorless and nonfluorescent in spirolactam form, but displayed remarkable color change and fluorescence in the ring-opened amide.<sup>13–17</sup> The selectivity and sensitivity of a probe was mainly decided by the recognition sites. 1,2,4-Triazole has lone electron pairs on N, which provide good coordination property to metal ions, and several 1,2,4-triazole containing host compounds have been synthesized for the detection of  $Cu^{2+}$ .<sup>13</sup> According to the soft–hard acid–base theory, S shows good affinity to  $Cu^{2+}$ , and so a –SH group was introduced in the system to improve the coordination ability of probe **P**. Furthermore, the

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semirigid property of 1,2,4-triazole containing complex could effectively chelate  $Cu^{2+}$  according to the ionic radius and also limit the geometric structure of the complex. In the present work, a  $Cu^{2+}$ -selective fluorescent probe derived from rhodamine B containing 1,2,4-triazole as subunit was proposed (Figure 1). Its application for imaging  $Cu^{2+}$  in living cells was also described.



Figure 1. Synthesis route of probe P.

## 2. Results and discussion

# 2.1. Effect of pH on P and P with $Cu^{2+}$

The pH dependence of the fluorescence intensity of  $\mathbf{P}$  and the  $\mathbf{P}$ -Cu<sup>2+</sup> system is shown in Figure 2. The results revealed that the fluorescence of the free  $\mathbf{P}$  could be negligible; however, a significant fluorescence enhancement was observed upon the addition of Cu<sup>2+</sup>, which was attributed to the opening of the spirolactam ring of the rhodamine unit. These data demonstrated that  $\mathbf{P}$  could work within a wide pH range of 5.8–8.4, which made it possible for the detection of Cu<sup>2+</sup> under physiological pH conditions. To exclude the influence of acidity on the test, pH 7.0 was fixed in the further research.



**Figure 2.** pH-dependent fluorescence of **P** (10  $\mu$ M) ( $\bullet$ , in red) and **P** (10  $\mu$ M) plus 100  $\mu$ M Cu<sup>2+</sup> ( $\blacksquare$ ) in HEPES buffers as a function of different pH values.

## 2.2. Uv-vis spectral response of P

In the UV-vis spectrum of  $\mathbf{P}$ , the absorption with various metal ions was recorded in ethanol-water solution (9:1, v:v, pH 7.0, 20 mM HEPES) (Figure 3). The results showed that a peak at 556 nm appeared with the addition of  $Cu^{2+}$ , and the colorless solution of  $\mathbf{P}$  was changed to an intense pink due to the spirolactam ring

opening of the rhodamine unit.  $Hg^{2+}$  and  $Ni^{2+}$  had negligible interference, while other metal ions, such as  $Na^+$ ,  $K^+$ ,  $Ag^+$ ,  $Ca^{2+}$ ,  $Mg^{2+}$ ,  $Zn^{2+}$ ,  $Pb^{2+}$ ,  $Cd^{2+}$ ,  $Co^{2+}$ ,  $Mn^{2+}$ , and  $Cr^{3+}$  did not show any influence on the absorbance of **P** under identical conditions.

#### 2.3. Fluorescence spectral response of P

The fluorescence property of **P** was measured to investigate the probe's selectivity in ethanol-water solution (9:1, v:v, pH 7.0, 20 mM HEPES) with addition of different metal ions (Figure 4). Compared with other tested metal ions, only  $Cu^{2+}$  caused a significant "turn-on" fluorescence response at 575 nm, and Hg<sup>2+</sup> had negligible interference. It indicated that **P** could selectively recognize  $Cu^{2+}$  in ethanol-water solution (9:1, v:v, pH 7.0, 20 mM HEPES) and the interference of other tested metal ions in the detection of  $Cu^{2+}$  could be negligible.

In the emission spectra (Figure 5), the fluorescence peak at 575 nm increased upon the addition of  $Cu^{2+}$ ; the linear portion of the plot of fluorescence intensity vs.  $Cu^{2+}$  could be used to detect the unknown concentration of  $Cu^{2+}$  over the range of  $8.0 \times 10^{-7}$  to  $7.5 \times 10^{-6}$  M with a detection limit of  $2.3 \times 10^{-7}$  M.



Figure 3. UV-vis spectra of **P** (10  $\mu$ M) with different metal ions (100  $\mu$ M) in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES).

Figure 4. Fluorescence spectra of **P** (10  $\mu$ M) with different metal ions (100  $\mu$ M) in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES).

One challenge for the probe is to obtain a specific detection system for  $\operatorname{Cu}^{2+}$  over a wide range of potentially competing ions, since the system might show cross-sensitivity toward other metal ions. Therefore, the competition experiments were conducted in the presence of 1 equiv. of  $\operatorname{Cu}^{2+}$  mixed with 5 equiv. of other metal ions as mentioned above. No significant variation in fluorescence intensity was found by comparison with the same amounts of  $\operatorname{Cu}^{2+}$  solution without other metal ions, and the relative error was less than  $\pm 5\%$  (Figure 6). For probe **P**, cross-sensitivity to the other metal ions was not observed, while an excellent selectivity toward  $\operatorname{Cu}^{2+}$  was exhibited. Thus, it indicated that the probe **P** was a  $\operatorname{Cu}^{2+}$ -specific fluorescent probe.

#### 2.4. The proposed reaction mechanism

The Job's plot was drawn to prove the complex ratio of  $\mathbf{P}$  with  $\mathrm{Cu}^{2+}$  (Figure 7). Total concentration of  $\mathbf{P}$  and  $\mathrm{Cu}^{2+}$  was kept at a fixed 50  $\mu$ M. The results showed that the maximum fluorescent emission intensity of  $\mathbf{P}$ -Cu<sup>2+</sup> complex appeared at 0.5, which indicated that a  $\mathbf{P}$ -Cu<sup>2+</sup> complex was formed in 1:1 mole ratio.





Figure 5. Fluorescence response of **P** (10  $\mu$ M) with various concentrations of Cu<sup>2+</sup> in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES).

**Figure 6.** Fluorescence response of **P** (10  $\mu$  M) to Cu<sup>2+</sup> ions (10  $\mu$  M) or to a mixture of the specified metal ions (50  $\mu$  M) with Cu<sup>2+</sup> ions (10  $\mu$  M) in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES).

To further understand the reaction mechanism of probe  $\mathbf{P}$  to  $\mathrm{Cu}^{2+}$ , EDTA titration experiments were conducted to examine the reversibility of the probe  $\mathbf{P}$  with  $\mathrm{Cu}^{2+}$  (Figure 8). Upon the addition of 50  $\mu$ M EDTA to the mixture of  $\mathbf{P}$  (10  $\mu$ M) and  $\mathrm{Cu}^{2+}$  (10  $\mu$ M) in ethanol–water solution (9:1, v:v, pH 7.0, 20 mM HEPES), the fluorescent emission intensity of  $\mathbf{P}$ – $\mathrm{Cu}^{2+}$  was significantly reduced and the color changed from pink to almost colorless. When  $\mathrm{Cu}^{2+}$  was added to the system again, the signals were almost completely reproduced, and the colorless solution turned pink. The results demonstrated that the binding of  $\mathbf{P}$  and  $\mathrm{Cu}^{2+}$ 



Figure 7. Job's plot of **P** with  $Cu^{2+}$  according to the method of continuous variation. The total concentration of **P** and  $Cu^{2+}$  was 50  $\mu$ M.



**Figure 8.** Reversible titration response of **P** to Cu<sup>2+</sup> in ethanolwater solution (9:1, v:v, pH 7.0, 20 mM HEPES): (a) **P** (10  $\mu$ M); (b) **P** (10  $\mu$ M) + Cu<sup>2+</sup> (10  $\mu$ M); (c) **P** (10  $\mu$ M) + Cu<sup>2+</sup> (10  $\mu$ M) + EDTA (50  $\mu$ M); (d) **P** (10  $\mu$ M) + Cu<sup>2+</sup> (10  $\mu$ M) + EDTA (50  $\mu$ M) + Cu<sup>2+</sup> (0.1 mM).

was a reversible process. According to the experimental results, the reaction mechanism was proposed as shown in Figure 9.



Figure 9. Proposed binding mode of  ${\bf P}$  and  ${\rm Cu}^{2+}$ 



Figure 10. Confocal fluorescence and brightfield images of HepG2 cells. a) Cells stained with 10  $\mu$ M P for 30 min at 37 °C; b) cells supplemented with 1  $\mu$ M CuCl<sub>2</sub> in the growth media for 30 min at 37 °C and then incubated with 10  $\mu$ M P for 30 min at 37 °C; c) bright field image of cells shown in a); d) bright field image of cells shown in b).

# 2.5. Preliminary analytical application

To further demonstrate the practical applicability of the probe  $\mathbf{P}$ , confocal microscopy experiments were further carried out, and the fluorescence images of HepG2 cells were recorded before and after the addition of Cu<sup>2+</sup> (Figure 10). The cells incubated with  $\mathbf{P}$  for 30 min at 37 °C showed very weak fluorescence, as shown in Figure 10a. When cells stained with  $\mathbf{P}$  were incubated with CuCl<sub>2</sub> (1  $\mu$ M), the color of the HepG2 cells showed significant changes (Figure 10b). The bright field images of Figure 10a and Figure 10b were shown as Figure 10c and Figure 10d, and the shapes of cells indicated that  $\mathbf{P}$  has low toxicity. These results suggested that probe  $\mathbf{P}$  can penetrate the cell membrane and might be used for detecting Cu<sup>2+</sup> in living cells.

In conclusion, a novel Cu<sup>2+</sup>-selective rhodamine B fluorescent probe containing 1,2,4-triazole as subunit was constructed. Cu<sup>2+</sup> could induce spirolactam ring opening of the rhodamine unit and achieved an "off–on" effect. The probe **P** can detect as low as  $2.3 \times 10^{-7}$  M Cu<sup>2+</sup>. In addition, the probe **P** was successfully used to detect Cu<sup>2+</sup> in living cells.

### 3. Experimental

#### 3.1. Reagents and instruments

All reagents and solvents are of analytical grade and used without further purification. The metal ions and anions salts employed were NaCl, KCl,  $CaCl_2 \cdot 2H_2O$ ,  $MgCl_2 \cdot 6H_2O$ ,  $Zn(NO_3)_2 \cdot 6H_2O$ ,  $PbCl_2$ ,  $CdCl_2$ ,  $CrCl_3 \cdot 6H_2O$ ,  $CoCl_2 \cdot 6H_2O$ ,  $NiCl_2 \cdot 6H_2O$ ,  $HgCl_2$ ,  $CuCl_2 \cdot 2H_2O$ ,  $FeCl_3 \cdot 6H_2O$ , and  $AgNO_3$ .

Fluorescence emission spectra were conducted on a Hitachi 4600 spectrofluorometer. UV-Vis spectra were obtained on a Hitachi U-2910 spectrophotometer. Nuclear magnetic resonance (NMR) spectra were measured with a Bruker AV 400 instrument and chemical shifts are given in ppm from tetramethylsilane (TMS). Mass spectra (MS) were recorded on a Thermo TSQ Quantum Access Agilent 1100.

## 3.2. Synthesis of compound P

# Compounds 1 and 2 were synthesized as reported.<sup>18,19</sup>

Compounds 1 (0.13 g, 1.0 mM) and 2 (0.496 g, 1.0 mM) were mixed in ethanol (40 mL). The reaction mixture was stirred at 80 °C for 4 h. After the reaction was finished, the solution was removed under reduced pressure. The precipitate so obtained was filtered and purified with silica gel column chromatography (petroleum ether/acetic ether = 5:1, v:v) to afford **P** as yellow solid. Yields: 83.4%. MS (ES+) m/z: 609.27 [M + H]<sup>+</sup>. <sup>1</sup>H NMR ( $\delta$  ppm,  $d_6$ -DMSO): <sup>1</sup>H NMR: 13.74 (s, 1H), 9.82 (d, 1H, J = 8.2), 8.34 (d, 1H, J = 8.2), 7.96 (d, 1H, J = 7.4), 7.65 (t, 1H, J = 7.4), 7.58 (t, 1H, J = 7.4), 6.45 (t, 4H, J = 8.3), 6.63 (t, 2H, J = 10.8), 7.08 (d, 1H, J = 7.6), 3.32 (m, 8H, J = 8.4), 2.21 (s, 3H), 1.08 (t, 12H, J = 7.8). <sup>13</sup>C NMR ( $\delta$  ppm,  $d_6$ -DMSO): 165.57, 161.88, 159.59, 153.19, 152.85, 149.62, 149.52, 143.24, 135.85, 132.41, 129.93, 129.55, 128.50, 127.88, 124.83, 124.42, 109.23, 105.17, 98.32, 66.49, 65.92, 44.57, 30.91, 19.55, 14.44, 13.29, 11.52, 11.29.

#### 3.3. General spectroscopic methods

Metal ions and chemosensor  $\mathbf{P}$  were dissolved in deionized water and DMSO to obtain 1.0 mM stock solutions, respectively. Before spectroscopic measurements, the solution was freshly prepared by diluting the high concentration stock solution with the corresponding solution. For all measurements, excitation/emission slit widths were 5/10 nm and excitation wavelength was 550 nm.

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#### References

- 1. Nolan, E. M.; Lippard, S. J. Chem. Rev. 2008, 108, 3443-3480.
- 2. Jeong, Y.; Yoon, J. Inorg. Chim. Acta 2012, 381, 2-14.

- 3. Xu, Z. C.; Yoon, J.; Spring, D. R. Chem. Soc. Rev. 2010, 39, 1996–2006.
- 4. Radisky, D.; Kaplan, J. J. Biol. Chem. 1999, 274, 4481-4484.
- 5. Rode, B. M.; Suwannachot, Y. Coordin. Chem. Rev. 1999, 190-192, 1085-1099.
- 6. Tapiero, H.; Townsend, D. M.; Tew, K. D. Biomed. Pharmacother. 2003, 57, 386–398.
- 7. Kwon, H.; Lee, K.; Kim, H. J. Chem. Comm. 2011, 47, 1773–1775.
- Peng, X. J.; Du, J. J.; Fan, J. L.; Wang, J. Y.; Wu, Y. K.; Zhao, J. Z.; Sun, S. G.; Xu, T. J. Am. Chem. Soc. 2007, 129, 1500–1501.
- 9. Huang, J. H.; Xu, Y. F.; Qian, X. H. Dalton Trans. 2009, 10, 1761–1766.
- 10. Lee, M. H.; Kim, H. J.; Yoon, S. W.; Park, N. J.; Kim, J. S. Org. Lett. 2008, 10, 213-216.
- 11. Wu, D. Y.; Huang, W.; Duan, C. Y.; Lin, Z. H.; Meng, Q. J. Inorg. Chem. 2007, 46, 1538–1540.
- 12. Yang, X. F.; Guo, X. Q.; Zhao, Y. B. Talanta 2002, 57, 883-890.
- 13. Zhang, J.; Yu, C. W.; Qian, S. Y.; Lu, G.; Chen, J. L. Dyes Pigm. 2012, 92, 1370–1375.
- 14. Du, J. J.; Fan, J. L.; Peng, X. J.; Sun, P. P. Org. Lett. 2010, 12, 476–479.
- 15. Weerasinghe, A. J.; Abebe, F. A.; Sinn, E. Tetrahedron Lett. 2011, 52, 5648–5651.
- 16. Kim, H.; Lee, M.; Kim, H.; Kim, J.; Yoon, J. Chem. Soc. Rev. 2008, 37, 1465–1472.
- 17. Yu, C. W.; Zhang, J.; Wang, R.; Chen, L. X. Org. Biomol. Chem. 2010, 8, 5277–5279.
- 18. Yu, C. W.; Zhang, J.; Li, J. H.; Liu, P.; Wei, P. H.; Chen, L. X. Microchim. Acta 2011, 174, 247-255.
- 19. Liu, C. Y.; Zhao, Q. Q.; Li, J. Chem. Reagents 2001, 23, 344-345 (in Chinese).