

A Novel Glycosidicly Linked Piperidine Alkaloid From *Cyclamen Coum*

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The structure of a novel piperidine type alkaloid from *Cyclamen coum* was established as 2- β -D-glycopyranosyl-2-undecil-3,5-dihydroxy-6-carboxypiperidine, **1**, whose structure has been deduced from spectral data.

Introduction

The occurrence of piperidine alkaloid in marine organisms is mentioned in the literature¹⁻⁶. In the course of our studies on bioactive substances from cyclamen organisms, we isolated glucose and undecil substituted piperidine-type alkaloid derivative from the *Cyclamen coum* and its structure was deduced as **1** from its NMR and FAB-MS spectral data.

Experimental

Instrumentation: NMR spectra were recorded on a Bruker AC 200L NMR at 200 MHz instrument in C₅H₅N using TMS as internal standard. IR spectra were taken on a Perkin Elmer 1600 spectrophotometer. (+) FAB-MS spectra were recorded on a Zabspec MS instrument, Flash column chromatography was performed on a silica gel 60 (230-400 mesh) and preparative TLC was performed with precoated silica gel F₂₅₄(20 × 20 cm 0.2 mm) plates. A voucher specimen has been deposited in deepfreeze at the Department of Chemistry, Karadeniz Technical University.

Isolation of compound 1: Specimens of the *Cyclamen coum* were collected in the Giresun Yağlıdere region, in the north of Turkey, in March, 1995. The chopped wet plants (~ 1500 g) were extracted with cold CH₃OH (1.5 lt, 3 times, 24 hours each). The total aqueous CH₃OH extract was filtered, and the filtrate was concentrated on a rotary evaporator at 30 °C. The aqueous extract thus obtained (0.4 liter) was extracted with CHCl₃ (150 ml, 3 times). After collecting CHCl₃ extract (450 ml), it was evaporated *in vacuo* at 30-35 °C. The crude mixture obtained (0.9 g) was chromatographed on a Kieselgel 60 (40 g, 230-400 mesh) flash column chromatograph. Elution with n-hexane, followed by discontinuous gradient elution with n-hexane-CHCl₃ (3:1-1:4) and CHCl₃ and then discontinuous gradient elution with CHCl₃-CH₃OH (9:1-2:3) and finally with CHCl₃-CH₃OH-H₂O (2:2.6:0.4) gave 43 fractions (*ca.* 15-20 ml each). Fractions

39-40 were combined after the analyses of TLC to give the ninth fraction (24.2 mg). The ninth fraction was chromatographed on a Kieselgel 60 (6 g, 230-400 mesh) flash column chromatograph. Elution with, respectively, n-hexane (30 ml), CHCl₃ (30 ml), and then discontinuous gradient elution with CHCl₃-CH₃OH (50 ml) (10:1-10:2) gave 34 fractions (*ca.* 3-4 ml each). Fractions 25-28 were combined after TLC analysis to give compound **1** (17.2 mg) (CHCl₃-CH₃OH, 0:0.5, R_f = 0.35) IR (KBr) ν max 3500-2500, 3500-3200, 3320, 2927, 1630, 1378-1360, 1075, 1048, 1025 cm⁻¹; ¹H NMR (C₅D₅N, 200 MHz) and ¹³C NMR (C₅D₅N, 50 MHz) (see Table 1); positive FAB-MS (MNBA) m/z 493(5) [M]⁺, 409(43) [M-85+H]⁺, 295(100) [M-175-H₂O+H]⁺, 235(55) [M-179-2H₂O-CO₂ + H]⁺, 159(10) [M-179-side-chaine(155)]⁺; and 155, 141, 127, 113, 99, 85, 71, 57.

Table 1. NMR Data for Compound **1** (200 MHz, C₅D₅N).

No	1 ^a		
	¹³ C (δ,ppm) ^b	APT	¹ H (δ,ppm) ^c
1	-	-	8.48 d, J=9 Hz
2	70.46	C	
3	78.48	CH	5.50
4	35.59	CH ₂	2.09-1.09
5	75.92	CH	4.62
6	51.75	CH	5.31
-COOH.....	175.68	C	^d
1'.....	105.62	CH	5.00 d, J=7.4 Hz
2'.....	75.18	CH	4.02
3'.....	78.48	CH	3.90
4'.....	72.46	CH	4.12
5'.....	78.49	CH	4.30
6'.....	62.64	CH ₂	4.50-4.12
1''.....	35.59	CH ₂	1.65
2''.....	34.00	CH ₂	1.24
3''.....	32.13	CH ₂	1.24
4''.....	29.63	CH ₂	1.24
5''.....	30.02	CH ₂	1.24
6''.....	30.02	CH ₂	1.24
7''.....	30.02	CH ₂	1.24
8''.....	27.95	CH ₂	1.24
9''.....	25.90	CH ₂	1.24
10''.....	22.95	CH ₂	1.24
11''.....	14.30	CH ₃	0.92 t, J=6.15 Hz

^aChemical shifts (ppm) are relative to internal TMS in C₅D₅N.

^bAssignments assisted by HETCOR data.

^cAssignments assisted by COSY data.

^dUnobsorved, in exchange with solvent

Hydrolysis of Compound 1: Compound **1** (4 mg) was hydrolyzed with 10% H₂SO₄ for 4 hours. The residue obtained showed the presence of D-glucose when compared with an authentic sample of this sugar on TLC (CH₃OH, R_f = 0.74).

Result and Discussion

The new piperidine type of glycosidically linked alkaloid was isolated from a methanolic extract of *Cyclamen coum*. We assigned the alkaloid structure based on the following evidence. The ^1H NMR (pyridine- d_5 , 200 MHz) spectrum of the compound exhibited a characteristic signal for an anomeric proton at δ 5.00 ppm. The coupling constant ($J = 7.4$ Hz) implied a β -configuration of the sugar residue. The ^{13}C NMR (pyridine- d_5 , 50 MHz) spectra of **1** showed one signal at δ 105.62 ppm for the anomeric carbon signal, also indicate of a β -configuration⁶⁻⁷.

The -NH- protons were observed at δ 8.48 (1H, d, $J=9$ Hz) ppm in the ^1H NMR spectrum. The carboxylic proton was unobserved in the ^1H NMR spectrum because of exchange with the solvent⁸. The ^1H NMR spectrum further showed piperidine ring signals at δ 5.50 (1H), 5.31(1H), 4.62 (1H) and 2.09-1.09 (2H) ppm. The side chain protons were also observed in the ^1H NMR spectrum at δ 1.65 (2H), 1.24 (18H) and 0.92 (3H, t, $J = 6.15$ Hz) ppm.

Standard 1D and 2D NMR procedures were employed to elucidate the structure of compound **1**. Conventional ^1H (200 MHz) and ^{13}C (50 MHz) NMR spectra combined with multiplicity-selected (APT) ^{13}C data yielded the gross structure of the molecule and showed it to consist of a hydroxy and carboxy substituted piperidine ring ($\text{C}_6\text{H}_9\text{O}_4$), a monosaccharide (C_6) sugar and long chain hydrocarbon (C_{11}) moiety. The COSY map afforded a comprehensive description of through-bond proton-proton connectivities. Corroborative evidence for the molecular structure thus derived was gleaned from the ^{13}C - ^1H chemical shift correlation (HETCOR).

The broad-bond ^{13}C NMR spectrum (pyridine- d_5 , 50 MHz) of compound **1** showed a carboxylic carbonyl signal at δ 175.68 ppm. The IR spectrum also showed bands for carboxyl ($\text{C}=\text{O}$; 1630, COO-H ; 2500-3500 cm^{-1}) and 2° amine (-NH-; 3320 cm^{-1}) functionalities.

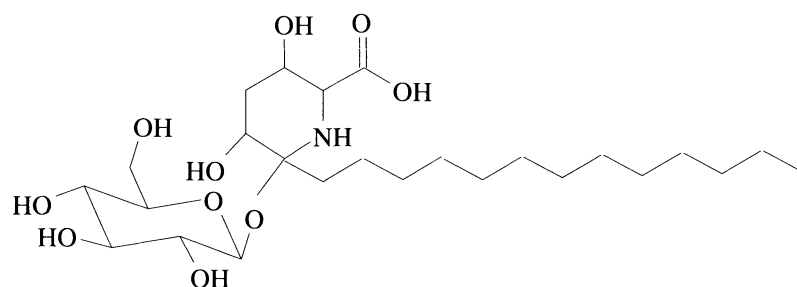
In order to identify the sugar moiety, compound **1** was hydrolyzed with 10% H_2SO_4 . The residue obtained showed the presence of D-glucose when compared with the authentic sample of this sugar on TLC.

The assigned ^1H and ^{13}C resonance for compound **1** is shown in Table 1. Comparisons of the spectral data in Table 1 with the published spectra of related alkaloids^{1-5,9-13} showed glucone to be a glucopyranosyl, and alkaloid a substituted piperidine type ring having hydroxy, carboxy and long chain hydrocarbon. The positive ion FAB mass spectrum (MNBA) of glucosidically linked piperidine type alkaloid exhibited prominent ions at m/z 493(5) $[\text{M}]^+$, 409(43) $[\text{M}-85+\text{H}]^+$, 295(100) $[\text{M}-175-\text{H}_2\text{O}+\text{H}]^+$, 225(55) $[\text{M}-179-2\text{H}_2\text{O}-\text{CO}_2+\text{H}]^+$ and 159(10) $[\text{M}-179\text{-side-chain}(155)]^+$ corresponding to $\text{C}_{23}\text{H}_{43}\text{NO}_{10}$ (Figure 1). The length of the side-chain was determined with the aid of ^{13}C , APT NMR and FAB-MS spectra¹⁴.

Thus, we conclude that compound **1** has the structure 2- β -glucopyranosyl-2-undecil-3,5-dihydroxy-6-carboxy piperidine, which is a novel natural product elucidate from the *Cyclamen coum*. The stereochemistry and synthesis of this compound is currently under investigation.

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2-β-D-glycopyranosyl-2-undecil-3,5-dihydroxy-6-carboxy piperidine, **1**

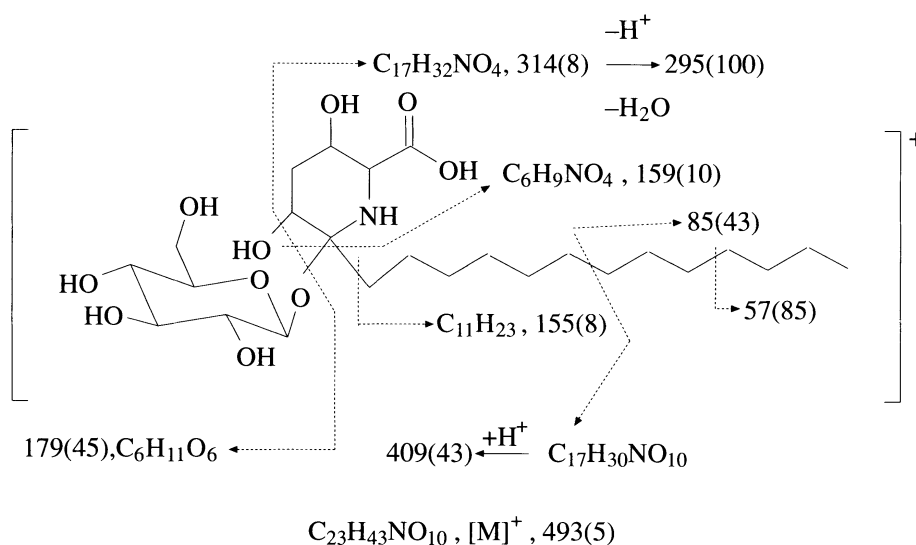


Figure 1. FAB-MS (*m/z*) spectral analysis of compound **1**.

References

1. M. Ishibashi, Y. Ohizumi, T. Sasaki, H. Nakamura, Y. Hirata and J. Kobayashi, *J. Org. Chem.*, **52**, 450 (1987).
2. J. Kobayashi, K. Naitoh, Y. Doi, K. Dekiand, M. Ishibashi, *J. Org. Chem.*, **60**, 6941 (1995).
3. E.A. Jares-Erijman, C.P. Bapat, A. Lithgow-Bertelloni, K.L. Rinehart and R. Sakai, *J. Org. Chem.*, **58**, 5732 (1993).
4. N.K. Gulavita and P.J. Scheuer, *J. Org. Chem.*, **54**, 369 (1989).
5. F. Kong and D.J. Faulkner, *J. Org. Chem.*, **58**, 970 (1993).
6. D.C. Jain, R.S. Thakur, A. Bahpai and A.R. Sood, *Phytochemistry*, **27**, 1216 (1988).
7. S. Seo, Y. Tomita, K. Tori and Y. Yoshimura, *J. Am. Chem. Soc.*, **100**, 3331 (1978).
8. W.W. Simons, "the **Sadtler Handbook of Proton NMR Spectra**", pp 1093 (Heyden & Son Ltd., Spectrum House, Hillview Gardens, London N.W., 1978).
9. T. Kiguchi, Y. Yuumoto, I. Ninomiya and T. Naito, *Tetrahedron Letters*, **33**, 7389 (1992).
10. S. Kanpp and J.J. Hale, *J. Org. Chem.*, **58**, 2650 (1993).
11. F.W. Wehrli, A.P. Marchand and S. Wehrli, pp. 355-425 John Wilwy & Sons, New York, 1988.

12. P. Clerc and S. Simon, "**Spectral Data for Structure Determination of Organic Compounds**", Springer-Verlag Heidelberg, New York, 1989.
13. E. Breitmaier, W. Voelter, "**Carbon-13 NMR Spectroscopy**", Third edition, pp. 277-276 (VCH Publishers, New York, NY), 1990.
14. J.A. Findlay, Z.Q. He and L.A. Calhoun, **J. Nat. Prod.**, **53**, 1015 (1990).