

## Effects of green tea on ACE gene expression in rat liver in CCl<sub>4</sub>-induced cirrhosis

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Received: 28.11.2011 • Accepted: 18.10.2012 • Published Online: 29.07.2013 • Printed: 19.08.2013

**Aim:** This study was designed to investigate the beneficial effects of green tea administration on angiotensin converting enzyme (ACE) gene expression in CCl<sub>4</sub>-induced liver cirrhosis in rats.

**Materials and methods:** A total of 24 male Wistar rats were divided into 3 groups as: Group 1, normal untreated rats; Group 2, CCl<sub>4</sub>-induced cirrhotic rats; Group 3: CCl<sub>4</sub>-induced cirrhotic + green tea treated rats. The beneficial effects of green tea were measured by alanine aminotransferase (ALT), alkaline phosphatase (ALP), total and direct bilirubin level, tissue malondialdehyde (MDA), super oxide dismutase (SOD), and catalase (CAT). The genomic DNA was isolated from excised tissue to determine the ACE genotypes using specific primers. The ACE gene expression in liver tissue was assessed using the quantitative RT-PCR method.

**Results:** Liver cirrhosis was indicated by high plasma ALT, direct bilirubin level, tissue MDA, and low SOD. The antioxidant enzymes SOD and CAT were low ( $P < 0.01$ ) in cirrhotic and green tea-treated rats. High activity ( $P < 0.01$ ) of ALT was observed in green tea-treated cirrhotic rats. The total and direct bilirubin levels were high ( $P < 0.01$ ) in CCl<sub>4</sub>-treated cirrhotic rats while they were low ( $P < 0.05$ ) in green tea-treated cirrhotic rats. The tissue MDA was high ( $P < 0.01$ ) in CCl<sub>4</sub>- and green tea-treated cirrhotic rats. ACE gene expression after 8 weeks of CCl<sub>4</sub> treatment in cirrhotic rats was significantly high ( $P < 0.05$ ), and was reversed in green tea-treated cirrhotic rats in comparison to controls.

**Conclusion:** The administration of green tea tries to correct the deteriorative biochemical and genetic changes during CCl<sub>4</sub>-induced liver cirrhosis in rats. The long-term consumption of green tea has beneficial effects on abnormally increased ACE gene activity during liver cirrhosis caused by CCl<sub>4</sub> administration in rats.

**Key words:** Green tea, liver cirrhosis, ALT, ALP, total and direct bilirubin, ACE gene

### 1. Introduction

Liver problems are a major health issue globally. The management of these problems seems inadequate, looking at their high frequency and their morbidity and mortality rates. It is now well documented that the management and treatment strategies being used for hepatic disorders produce severe adverse effects (1). Liver scars and progressive damage to the liver tissue start with subendothelial or hepatic fibrosis, and develop with nodule formation. This condition may be caused by unhealthy fats, alcohol, toxins, and viruses, as well as some therapeutic agents that can affect liver cells and produce cirrhosis (2).

Angiotensin converting enzyme (ACE) is a zinc metalloproteinase widely distributed on the surface of endothelial and epithelial cells (3). The renin angiotensin system (RAS) is a cascade of the circulatory system

basically involved in the regulation of blood pressure and water-electrolytes balance (4,5). ACE is the key enzyme in this system, which converts angiotensin I to the potent vasoconstrictor angiotensin II (6). The RAS is known to play a role in the pathophysiology of various diseases, including fibrosis in the lung, kidney, and heart, during chronic inflammation through the regulation of cell growth, inflammation, oxidative stress, angiogenesis, fibrosis, and cirrhosis (7). The ACE gene expression and its role as a potential disease marker have been investigated in several diseases (8). However, the abnormalities in expression of the ACE gene during liver injury and cirrhosis, with its contribution to the development of disease, have not yet been defined.

The development and progression of liver cirrhosis in experimental animals are widely studied by CCl<sub>4</sub> administration. The majority of cases, including liver

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cirrhosis and fibrosis, have implemented these models for deposition of extracellular matrix (9). The haloalkane free radicals, which are formed during the biotransformation process of  $\text{CCl}_4$ , can damage the hepatocytes, making the liver an important target for  $\text{CCl}_4$  (10).

The use of plant extracts, in the form of herbal medicine, is being exploited to tackle various medical issues. A large number of beneficial health effects are reported to be produced by these medicinal plants, including antibacterial, antifungal, antitumor, antihypertensive, antidiabetic, and antiinflammatory effects (11–13).

*Camellia sinensis*, commonly called green tea, exhibits a wide range of beneficial effects on human and animal health. It contains antioxidants like polyphenols, which contribute to the prevention of cancers (14). It has beneficial effects in collagen-induced arthritis, inflammatory bowel disease, and paw edema (15). The long-term consumption of green tea has been reported to greatly reduce the risk of liver injury (16). The cellular damage and abnormal levels of antioxidants are caused by oxidative stress and cellular damage leading to DNA damage. The polyphenols contained in the tea are antimutagenic and anticarcinogenic due to their inhibition of cancer cell proliferation and induction of apoptosis (17). By considering the above-mentioned facts, this study was conducted to evaluate the beneficial effects of green tea consumption on ACE gene expression in  $\text{CCl}_4$ -induced liver cirrhosis in rats.

## 2. Materials and methods

### 2.1. Study design and animals

A total of 24 male albino Wistar rats (200–250 g body weight), purchased from the animal house of the International Center for Chemical and Biological Sciences, University of Karachi, were selected for the study. The rats were acclimatized to the laboratory environment for 1 week before the commencement of the experiment. All the rats were caged with a sawdust-covered floor in a quiet and temperature-controlled room ( $23 \pm 4$  °C). Rats were given free access to a standard rat diet and water. All the protocols regarding this study were approved by the institutional ethical committee and conducted according to the ethical guidelines for the use of animals in laboratory experiments. The age- and sex-matched rats were divided into 3 experimental groups (8 rats per group): Group 1, the control, was fed on the standard diet and water. Group 2 was treated with  $\text{CCl}_4$  (0.8 mg/kg body weight (b.w.), intraperitoneally (i.p)). The dose was given intraperitoneally at 1145 hours, once a week for 8 weeks. Group 3 received  $\text{CCl}_4$  (0.8 mg/kg b.w., i.p) weekly for 8 weeks as well as green tea extract (5 %) orally on a daily basis. The volume of green tea consumed by each rat was measured at 1130 hours every morning. The mean intake of green tea extract in these rats was  $45.5 \pm 12.56$  mL on day 1, which was increased to  $115.5 \pm 15.45$  mL on day 45.

### 2.2. Collection of samples

After 8 weeks of treatment, blood samples were collected by decapitating the animals. After the trimming of connective tissues, liver tissues were rinsed with saline to eliminate blood contamination, dried, weighed, and then kept at  $-80$  °C until analyzed.

### 2.3. Preparation of liver homogenate

A small piece of liver was weighed, perfused in saline, and homogenized in ice-cold potassium chloride (1.17 %) with the help of a homogenizer. The resultant was centrifuged at  $800 \times g$  at 4 °C for 5 min in order to separate the nuclear debris. The supernatant was centrifuged at  $10,500 \times g$  at 4 °C for 20 min to get the postmitochondrial supernatant. This was used to estimate catalase (CAT), super oxide dismutase (SOD), and malondialdehyde (MDA) activities.

### 2.4. Estimation of liver enzymes

Plasma alanine aminotransferase (ALT), alkaline phosphatase (ALP), and total and direct bilirubin levels were analyzed using commercially available reagent kits from Randox Laboratories Ltd., UK.

### 2.5. Estimation of CAT activity

The previously described method of Sinha et al. (18) was used to measure the CAT activity.

### 2.6. Estimation of SOD

The method of Kono (19) was used to estimate the activity of SOD in the cell-free supernatant.

### 2.7. Assessment of tissue lipid peroxidation

A total of 10  $\mu\text{L}$  of butylated hydroxytoluene (0.5 M in acetonitrile) was added to prevent oxidation of the homogenate, and the homogenate was stored at  $-70$  °C until analysis for MDA.

### 2.8. Estimation of MDA

The tissue lipid peroxidation was measured by means of the MDA content. The method of Ohkawa et al. (20) was used in the form of thiobarbituric acid reacting substances.

### 2.9. Estimation of total and direct bilirubin

The total and direct bilirubin level was estimated by a previously described method (21).

### 2.10. Genotyping

Genomic DNA was isolated from excised tissue as described previously (22). To determine the ACE genotypes, primers were used, as described by Hilbert (23), to amplify the microsatellite located at the 50 end of the intron between exons 13 and 14.

### 2.11. ACE gene expression in liver tissue

The expression of ACE gene in liver tissue was assessed using the quantitative reverse-transcription polymerase chain reaction (RT-PCR) method. RNA was isolated using Total RNA Prep Plus (A and A Biotechnology, Gdansk, Poland). In brief, an amplification reaction was performed in 12.5  $\mu\text{L}$  of total volume, containing a pair of specific

primers: 5' CAGCTTCATCATCCAGTTCC 3' and 5' CTAGGAAGAGCAGCACCCAC 3'. The PCR program consisted of 30 cycles at an annealing temperature of 52–64 °C. Restriction fragments were subsequently analyzed in 2% agarose gel stained with the help of ethidium bromide (24) (Figure 1).

**2.12. Statistical analyses**

The results are shown as mean ± standard error of them mean (SEM). Statistical significance and differences between control and test groups were examined by Student's t-test. Statistical probabilities of P < 0.01 and P

< 0.05 were taken to be significant. All analyses were done using SPSS 17.0 for Windows.

**3. Results**

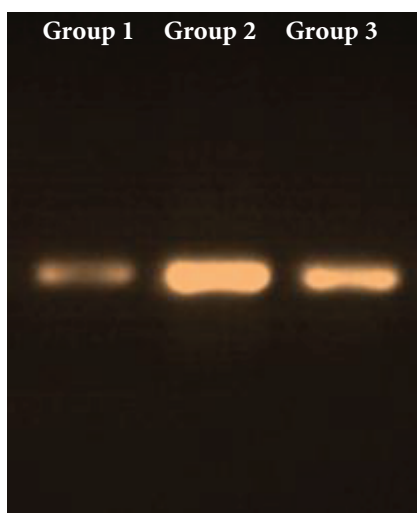
The activity of the antioxidant enzyme SOD was significantly lower (P < 0.01) in CCl<sub>4</sub>-treated cirrhotic and green tea-treated rats, whereas no difference was found in green tea-treated cirrhotic rats when compared to controls. CAT activity was significantly lower (P < 0.05) in CCl<sub>4</sub>-treated cirrhotic rats. No change was observed in the CAT level in green tea-treated cirrhotic rats as compared to the controls (Table 1).

A high ALT activity (P < 0.01) was observed in green tea-treated cirrhotic rats, whereas no significant difference was observed in the activity of ALP in green tea-treated cirrhotic rats compared to the controls (Table 2).

Total bilirubin was significantly higher (P < 0.01) in CCl<sub>4</sub>-treated cirrhotic rats and lower (P < 0.05) in green tea-treated cirrhotic rats when compared to the controls. The direct bilirubin level was also high (P < 0.01) in CCl<sub>4</sub>-treated cirrhotic and green tea-treated cirrhotic rats as compared to controls (Table 3).

The tissue lipid peroxidation, as measured by tissue MDA, was found to be significantly higher (P < 0.01) in CCl<sub>4</sub>-treated cirrhotic and green tea-treated cirrhotic rats compared to controls (Table 4).

Figure 2 shows the expression of the ACE gene in the experimental groups. The expression of the ACE gene after 8 weeks of CCl<sub>4</sub> treatment in cirrhotic rats was significantly increased (P < 0.05) in comparison with tissue samples from the control group. This gene expression was found to be reversed in the green tea-treated cirrhotic rats (Figure 2).



**Figure 1.** Agarose gel electrophoresis showing the amplification for the ACE gene in controls (Group 1), CCl<sub>4</sub>-induced cirrhotic rats (Group 2), and green tea-treated cirrhotic rats (Group 3).

**Table 1.** Effects of green tea on SOD and CAT activity in liver damage in rats.

Parameters	Group 1 (Controls)	Group 2 (CCl <sub>4</sub> -treated rats)	Group 3 (CCl <sub>4</sub> + green tea-treated rats)
SOD (U/g)	26.08 ± 2.84	9.37 ± 0.88*	21.4 ± 3.77
CAT (mmol/g)	3.97 ± 0.34	1.505 ± 0.24**	2.739 ± 1.53

In all tables, values are mean ± SEM. Significant difference between controls and test groups was determined by Student's t-test; \*: P < 0.01, \*\*: P < 0.05 as compared to controls.

**Table 2.** Effects of green tea on liver enzymes in liver damage in rats.

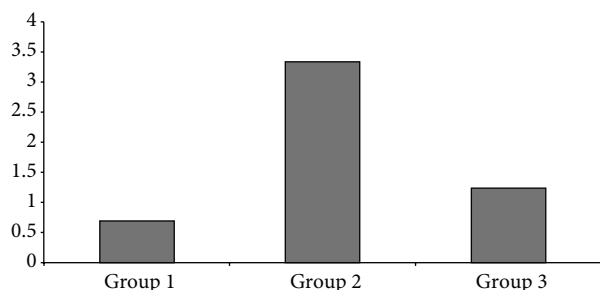
Parameters	Group 1 (Controls)	Group 2 (CCl <sub>4</sub> -treated rats)	Group 3 (CCl <sub>4</sub> + green tea-treated rats)
ALT (IU/L)	52.55 ± 3.39	896.49 ± 39.08*	487.73 ± 18.64*
ALP (IU/L)	484.16 ± 19.06	947.16 ± 27.08**	528.83 ± 69.52

**Table 3.** Effects of green tea on total and direct bilirubin in liver damage in rats.

Parameters	Group 1 (Controls)	Group 2 (CCl <sub>4</sub> -treated rats)	Group 3 (CCl <sub>4</sub> + green tea-treated rats)
Total bilirubin ( $\mu\text{mol/L}$ )	13.45 $\pm$ 2.61	24.82 $\pm$ 0.83*	8.87 $\pm$ 0.28*
Direct bilirubin ( $\mu\text{mol/L}$ )	4.32 $\pm$ 1.09	16.77 $\pm$ 3.34*	6.58 $\pm$ 0.67*

**Table 4.** Effects of green tea on tissue lipid peroxidation in liver damage in rats.

Parameter	Group-1 (Controls)	Group-2 (CCl <sub>4</sub> treated rats)	Group-3 (CCl <sub>4</sub> + green tea treated rats)
MDA (nmol/g)	1.09 $\pm$ 0.26	2.17 $\pm$ 0.61*	1.83 $\pm$ 0.48*

**Figure 2.** ACE gene expression in controls (Group 1), CCl<sub>4</sub>-induced cirrhotic rats (Group 2), and green tea-treated cirrhotic rats (Group 3).

#### 4. Discussion

Green tea has been reported to produce beneficial effects for the treatment of cardiovascular disease, diabetes, dermatological manifestations, obesity, and oral problems (25). The antioxidant activities, including scavenging of such reactive oxygen species (ROS) as superoxide, hydroxyl, and peroxy radicals; inhibition of lipid oxidation; and inhibition of low-density lipoprotein oxidation have been reported to be the properties of the catechins found in green tea. Chemoprotection has been considered as another important feature of green tea that prevents carcinogenesis in liver damage (26).

This study describes the low levels of SOD in a group of rats with cirrhosis of the liver (Table 1), indicating an imbalance in free radicals leading to cellular damage. The administration of CCl<sub>4</sub> caused liver injury; as a result new cells were not synthesized and SOD was not properly formed, which caused the low levels of SOD in cirrhotic rats. The negative feedback caused by the multiplication of cancer cells is an activity of SOD in cirrhotic cells. The

loss of lipid peroxidation describes the malignancy of hepatocarcinoma and enhanced lipid peroxidation in liver cells, which may lead to necrosis (27).

The levels of SOD in green tea-treated rats were significantly lower ( $P < 0.01$ ) when compared to the controls, while the combined effect of CCl<sub>4</sub> and green tea resulted in a nonsignificantly higher level of SOD. Antioxidants are usually found to elevate SOD expression while SOD mimetic suppresses tumorigenesis both in vivo and in vitro (28).

The cirrhotic rats that received CCl<sub>4</sub> showed a low level of catalase activity (Table 1) that induces liver injury by lowering the antioxidant status. The undermining of antioxidant substances in patients suffering from hepatocellular carcinoma is reported as an indicator of a distortion of the oxidant-antioxidant balance and the decrease in antioxidant system efficiency (29); thus, the participation of free radicals is established in the pathophysiology of carcinoma. H<sub>2</sub>O<sub>2</sub> is produced by green tea extracts in a weak alkaline medium. Intact membranes and embedded enzymes are responsible for integrating the cell membrane in cirrhosis. The polyphenol found in green tea also has cancer preventing potential, whereas endogenous CAT plays a pivotal role in H<sub>2</sub>O<sub>2</sub>-produced cytotoxicity. The accumulation of H<sub>2</sub>O<sub>2</sub> prevents the cleavage of bonds between oxygen that may lead to excessive hydroxylation and production of highly reactive and unstable oxidizing species that can readily react with any biomolecule. Almost all biological membranes are penetrated by H<sub>2</sub>O<sub>2</sub> and can be damaged in various cellular locations far from its original point. That may be the reason why the combined effects of green tea and CCl<sub>4</sub> caused higher levels of CAT activity in hepatotoxic rats, as compared to the controls.

Altered liver enzyme activity (ALT and ALP) (Table 2) and total and direct bilirubin (Table 3) were also observed in this study, which strongly suggested liver injury. The damaged structural integrity of the liver is mainly caused by the increased serum ALT and ALP located in cytosol that, after cellular damage, are released into main circulation. The ALT was significantly increased in CCl<sub>4</sub>-treated rats where the cells of liver were inflamed and ALT leaked into the blood stream, while ALP was significantly decreased in CCl<sub>4</sub>-treated rats. ALP was synthesized in the bile canalicular cells and appeared in the blood stream only whenever the biliary duct was inflamed or blocked. It might be possible that CCl<sub>4</sub> produced only hepatic damage and not biliary damage. The magnesium deficiency is transcribed into a low ALP level, which is inhibited due to chelation of zinc and magnesium, enzyme cofactors (30). Direct bilirubin was elevated in CCl<sub>4</sub>-treated rats, which indicated elevated production and lower liver uptake, and lower conjugation, as well as secretion from the liver or biliary tract obstructions (17), decreasing amounts of reducing equivalents such as NADPH reductase, and reduced glutathione (GSH). GSH maintains the integrity of red blood cell membranes; its reduced level increases hemolysis and increases bilirubin. Green tea increases the biliary flow, and bile helps to eliminate the bile salts, fats, and toxins from the body. The antioxidant and prooxidant

activities are performed by scavenging ROS via enzymatic and nonenzymatic reactions by polyphenolic compounds in cells (31,32).

The cirrhotic and green tea-treated group showed low MDA (Table 4) because of the inhibition of lipid peroxidation by polyphenol-rich green tea extracts (30). This study also indicates the severe impairment in the antioxidant system of cirrhotic rats, which results in high levels of MDA in cancerous tissues. Similarly, a low catalase activity was observed due to oxidative tissue damage. A significant decrease in the antioxidant enzyme system was also observed, due to the inflammatory oxidative stress (33).

The findings of this study suggest that the consumption of green tea has protective and beneficial effects in cirrhotic conditions caused by the administration of CCl<sub>4</sub> in experimental animals. This may be because of the potential antioxidant properties and oxidative stress-reducing ability of green tea extracts. It is therefore recommended that it should be supplemented with diet for a sufficient time period to minimize the chances of hepatic injury due to the presence of free radicals. It can be concluded that these kinds of herbal extracts may be used for protection upon possible exposure to hepatotoxicities. In the future, more focused studies will be of help to improve the knowledge regarding green tea extracts and their beneficial effects on liver damages and injuries.

## References

1. Wolf AT, Maurer R, Glickman J, Grace ND. Hepatic venous pressure gradient supplements liver biopsy in the diagnosis of cirrhosis. *J Clin Gastroenterol* 2008; 42: 199–203.
2. Bruno RS, Dugan CE, Smyth JA, DiNatale DA, Koo SI. Green tea extract protects leptin-deficient spontaneously obese mice from hepatic injury. *J Nutr* 2008; 138: 323–1.
3. Sayed-Tabatabaei FA, Oostra BA, Isaacs A, van Duijn CM, Witteman JCM. ACE polymorphism. *Circ Res* 2006; 98: 1123–33.
4. Shahid SM, Tabassum M. Integration of ionic hypothesis to non-communicable disorders. *J Basic Appl Sci* 2005; 1: 55–60.
5. Ruiz-Ortega M, Ruperez M, Esteban V. Molecular mechanisms of angiotensin II induced vascular injury. *Curr Hypertens Rep* 2003; 5: 73–9.
6. Stroth U, Unger T. The renin-angiotensin system and its receptors. *J Cardiovasc Pharmacol* 1999; 33: 21–8.
7. Jayapalan JJ, Muniandy S, Chan SP. Null association between ACE gene I/D polymorphism and diabetic nephropathy among multiethnic Malaysian subjects. *Indian J Hum Genet* 2010; 16: 78–86.
8. Baudin B. New aspects on angiotensin-converting enzyme: from gene to disease. *Clin Chem Lab Med* 2002; 40: 256–65.
9. Nakade Y, Yoneda M, Nakamura K, Makina I, Terano A. Involvement of endogenous CRF in carbon tetrachloride-induced acute liver injury in rats. *Am J Physiol Regul Integr Comp Physiol* 2002; 282: 1782–8.
10. Ozardali I, Bitiren M, Ali ZK, Zerim M, Aksoy N, Musa D. Effects of selenium on histopathological and enzymatic changes in experimental liver injury of rats. *Exp Toxicol Pathol* 2004; 56: 59–64.
11. Amutha M, Arunachalam R, Umamaheswari M, Usharamalakshmi A, Ramakrishnan S, Annadurai G. Medicinal use of *Camellia sinensis* on lactose intolerance. *J Biol Sci* 2010; 10: 1–5.
12. Kyung JL, Jea HC, Hye GJ. Hepatoprotective and anti-oxidant effects of the coffee diterpenes kahweol and cafestol on carbon tetrachloride-induced liver damage in mice. *Food Chem Toxicol* 2007; 45: 2118–25.
13. Aydın M, Çelik S. Effect of lycopene on plasma glucose, insulin level, oxidative stress, and body weights of streptozotocin induced diabetic rats. *Turk J Med Sci* 2012; 42 (Sup. 2): 1406–13.
14. Dona M, Dell'Aica I, Calabrese F, Benelli R, Morini M, Albini A et al. Neutrophil restraint by green tea: inhibition of inflammation, associated angiogenesis and pulmonary fibrosis. *J Immunol* 2003; 170: 4335–41.

15. Das M, Sur P, Gomes A, Vedasiromoni JR, Ganguly DK. Inhibition of tumor growth and inflammation by consumption of tea. *Phytother Res* 2002; 16: S40–4.
16. Jin X, Zheng RH, Li YM. Green tea consumption and liver disease: a systematic review. *Liver Int* 2008; 28: 990–6.
17. Bun SS, Bun H, Guedon D, Rosier C, Ollivier E. Effect of green tea extracts on liver functions in Wistar rats. *Food Chem Toxicol* 2006; 44: 1108–13.
18. Sinha KA. Colorimetric assay of catalase. *Anal Biochem* 1972; 47: 389–94.
19. Kono Y. Generation of superoxide radical during auto-oxidation of hydroxylamine and an assay for superoxide dismutase. *Arch Biochem Biophys* 1978; 186: 189–95.
20. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 1979; 95: 351–8.
21. Jendrassik L, Gróf P. Vereinfachte photometrische Methoden zur Bestimmung des Blutbilirubins. *Biochem Zeitschrift* 1938; 297: 82–9.
22. Korstanje R, Li R, Howard T, Kelmenson P, Marshall J. Influence of sex and diet on quantitative trait loci for HDL cholesterol levels in an SM/J by NZB/BLN intercross population. *J Lipid Res* 2004; 45: 881–8.
23. Hilbert P, Lindpaintner K, Beckmann JS, Serikawa T, Soubrier F. Chromosomal mapping of two genetic loci associated with blood-pressure regulation in hereditary hypertensive rats. *Nature* 1991; 353: 521–9.
24. Sulikowski T, Domanski L, Zietek Z, Adler G, Pawlik A, Kaczmarczyk M et al. The effect of preservation solutions UW and EC on the expression of renin I, angiotensinogen and angiotensin I-converting enzyme genes in rat kidney. *Ann Transplant* 2011; 16: 108–13.
25. Hara Y. *Green Tea: Health Benefits and Applications*. New York: Marcel Dekker; 2001.
26. Luper S. A review of plants used in the treatment of liver disease: part two. *Alt Med Rev* 1999; 4: 178–88.
27. Beckman JS, Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and the ugly. *Am J Physiol* 1996; 271: C1424–37.
28. Elchuri S, Oberley TD, Wenbo Q, Eisenstein RS, Roberts LJ, Remmen HV et al. CuZnSOD deficiency leads to persistent and widespread oxidative damage and hepatocarcinogenesis later in life. *Oncogene* 2005; 24: 367–80.
29. Reddy EP, Suchitra MM, Bitla AR, Sivakumar V, Rao PS. Anti-oxidant enzymes status in South Indian hemodialysis patients. *Int J Biol Med Res* 2011; 2: 682–7.
30. Zaidi SM, Kashif R, Al-Qirim, Tariq M, Banu N. Effects of anti-oxidant vitamins on glutathione depletion and lipid peroxidation induced by restraint stress in the rat liver. *Drugs RD* 2005; 6: 157–65.
31. Pyo YH, Lee TC, Logendra L, Rosen RT. Hepatoprotective activity of *Azadirachta indica* leaf extract: part II. *J Ethnopharmacol* 2004; 89: 217–9.
32. Kandemir O, Polat G, Saraçoğlu G, Taşdelen B. The predictive role of AST level, prothrombin time, and platelet count in the detection of liver fibrosis in patients with chronic hepatitis C. *Turk J Med Sci* 2009; 39: 857–62.
33. Kuper H, Tzonou A, Kaklamani E, Hsieh CC, Lagiou P, Adami HO. Tobacco smoking, alcohol consumption and their interaction in the causation of hepatocellular carcinoma. *Int J Cancer* 2000; 85: 498–502.