

**Turkish Journal of Medical Sciences** 

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# An immunohistochemical examination of cinnamon extract administration on distribution of NGF (nerve growth factor) and Trk-A (tyrosine kinase-A) receptor for diabetic rats with pancreatic tissue

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Received: 23.12.2020 • Accepted/Published Online: 24.06.2021	•	Final Version: 21.10.2021
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Background/aim: The aim of this study was to investigate the administration of cinnamon extract that is known to be effective in decreasing the high blood glucose and the distribution of NGF and Trk-A receptor in pancreas with immunohistochemistry way.

Materials and methods: The experimental groups were defined as control, sham, cinnamon, diabetes, and diabetes-cinnamon. At the end of the experiment, the pancreatic tissue samples were obtained for the rats. The hematoxylin-eosin and triple staining were used to examine histology. The immunohistochemical methods were performed on the sections of pancreatic tissue. In all groups, the body weight and fasting blood glucose obtained from the male and female rats and the values were statistically evaluated.

Results: The NGF immunoreactivity was observed in acinus, excretory pars, excretorius ducts, and islets of Langerhans for the pancreatic tissues of female and male rats in all groups. The Trk-A immunoreactivity was observed in acinus and islets of Langerhans for the pancreatic tissues of female and male rats in the control, sham, and cinnamon groups.

Conclusion: As a result, it was determined that the cinnamon, which is effective on blood glucose levels, has a positive effect on the NGF production in pancreas.

Key words: Cinnamon, diabetes, female and male rats, NGF, pancreas, Trk-A

# 1. Introduction

In diabetes which is a chronic metabolic disease, lack of insulin or problems in insulin utilization occur [1]. It was stated that the significant reduction in the beta ( $\beta$ ) cell found in the pancreas has an important role in the appearance of diabetes. The diabetes disease has two types. While the immune destruction of all  $\beta$  cells occurs in type-1 diabetes, the  $\beta$  cells are partially destructed in type-2 diabetes [2]. The cinnamon is a native herb belonging to Sri Lanka and is also a tropical Asian spice derived from the inner bark of several trees of the Cinnamomum genus. Some types of the cinnamon are used not only as a spice in foods, but also in traditional and modern medicine. Both Ceylon cinnamon (cinnamon oil) and Chineese cinnamon oils are used in the pharmaceutical production [3–5]. The cinnamon was reported that it reduces the high blood glucose level, repairs the damaged  $\beta$  cells, and has positive effects on the diabetes [6,7]. The neurotrophins (NTs) are the growth factors with polypeptide structure. The neurotrophins are necessitated for controlling of the

synaptic functions and plasticity and maintaining of the neural life morphology and differentiation. However, the neurotrophins were remarked to have different functions in the systems apart from the nerve system [8,9]. The nerve growth factor (NGF) is one of the first discovered members of the neurotrophin family. The NGF was defined as a trophic protein that has a role in different processes such as neuroblast proliferation, dorsal root ganglion maturation and axonal growth and acts as a message-receiver between the tissues that react against the peripheral stimulation and nerves stimulating these tissues [9-12]. The NGF and Trk-A receptors have important roles in the development, differentiation, survival and regulation of neurotransmitters of the peripheral and central nerves in the embryonic and postnatal life [13]. The NGF was reported to attach to two types of receptors located on the cell surface. One of these receptors is known as tyrosine kinase A (Trk-A) and the other one is known as p75, which does not have a tyrosine kinase activity [14,15]. The NGF synthesis does not only arise from the

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target tissues of central and peripheral nerve system, but also from the mast cells, lymphocytes, lipid tissue cells, pancreatic  $\beta$  cells, hair follicles, thymus, spleen [16–20], smooth muscle cells, fibroblasts [10,11], and trigeminal mesencephalic neurons [21]. The aim of this study is to determine the NGF and Trk-A levels in pancreatic tissue on cinnamon extract administration to diabetic rats by immunohistochemical methods.

# 2. Materials and methods

## 2.1. Animals

The approval for the research was received from Cumhuriyet University Animal Experimentation Local Ethics Committee with 65202830/118 board number on 19.06.2014. The animals used in the research were supplied from Ataturk University Laboratory Animals Unit.

## 2.2. Method

We designed a prospective experimental study. In this research, 60 (30 males + 30 females) one-month old Sprague dawley rats were used. The rats were housed at  $25 \pm 2$  °C room temperature and 60%–65% humidity conditions for 12 h in dark cycle and for 12 h in light cycle in the cages that are cleaned daily. This study complies with International Helsinki Declaration principles. All rats planned to be used in the experiment were weighed. Rats were randomly divided into five groups. The groups were formed as described below:

3. Control Group (n = 12) (6 females+6 males): The rats were fed ad-libitum with normal rat food.

4. Sham Group (n = 12) (6 females+6 males): 50 mg/kg of sodium citrate solution were intraperitoneally applied (i.p.).

5. Cinnamon Group (n = 12) (6 females+6 males): The cinnamon extract was given to the cinnamon groups as oral gavage method for 14 days (200 mg/kg) by dissolving in 1 mL of distilled water [22].

6. Diabetes Group (n = 12) (6 females + 6 males): The streptozotocin (STZ) (the streptozotocin was dissolved in the citric acid and disodium hydrogen phosphate buffer and pH was 4.5) was intraperitoneally applied for only one dose as 50 mg/kg. The animals were considered as diabetic, if their blood glucose values were above 250 mg/dL on the third day after the STZ injection [23].

7. Diabetes+Cinnamon Group (n = 12) (6 females+6 males): The streptozotocin (STZ) was dissolved in the citric acid and disodium hydrogen phosphate buffer while the pH was 4.5) was intraperitoneally applied for only one dose as 50 mg/kg. The animals were considered as diabetic, if their blood glucose values were above 250 mg/dL on the third day after the STZ injection. The cinnamon extract was applied by oral gavage method for 14 days (200 mg/ kg) [22,23].

At the end of the study, the body weights and blood glucose of the rats were measured and then, they were taken to the deep sedation with disinfection by using an anesthetic substance named Sevoflurane, and the pancreatic tissue samples were subsequently obtained. At the end of the study, the blood glucose levels were measured by taking from the tail vein of the rats fasted for 8 h.

# 2. 2. 1. Preparation of cinnamon extract

Ceylon cinnamon was used in the study in powder form. Ten grams of powdered cinnamon were taken and dissolved in 100 mL of ethanol. The mixture obtained was left to shake on a shaker for 24 h at the room temperature. After the time expired, the mixture was filtered by the help of a filter paper. The solvent content was removed in the rotary evaporator at the low pressure and low temperature. The extracts were stored at -20 °C until the experiment [24]. The cinnamon extract was applied by dissolving in 1 mL of distilled water.

## 2. 2. 2. Histological examinations

The pancreatic tissue samples were fixed within 10% formalin solution. Following the routine procedures, they were embedded into the paraffin and 5  $\mu$ m sections were obtained from the paraffin blocks. In order to demonstrate the histological structure of pancreatic tissue, the sections were performed by Crossman's triple staining and hematoxylin-eosin (HE) staining [25] methods and examined under a light microscope (Olympus BX51; Olympus Optical Co. Osaka, Japan).

### 2. 2. 3. Immunohistochemical examinations

The sections obtained from the paraffin blocks were incubated in 3% H<sub>2</sub>O<sub>2</sub> (hydrogen peroxide) and prepared in 0.1 M of phosphate buffered saline (PBS) for 15 min to inhibit the endogenous peroxidase activity after the deparaffinization and rehydration procedures. Then, the sections were washed in the PBS solution. The NGF was applied to the tissues in order to release the antigenic receptors that were boiled by a microwave for 10 min at 600 watts in a citrate buffer solution (pH: 6.0), the Trk-A was applied to the tissues in order to release the antigenic receptors that were boiled by a microwave for 10 min at 600 watts in a tris-EDTA Buffer solution (pH: 6.0). The avidine-biotin-peroxidase protocol was used to investigate the NGF and Trk-A immunoreactivity [26]. The tissues were incubated by Blocking Solution A (Invitrogen-Histostain plus Bulk Kit) for 10 min and then, the NGF (Abcam-AB6198) (1/600 dilution) primary antibody and Trk-A (Abcam-AB76291) (1/400 dilution) primary antibody were diluted by the PBS applied. The tissues were treated by the NGF incubated overnight at +4 °C. The tissues were treated by the Trk-A incubated by the primary antibody at the room temperature for

1 h. Then, the biotinylated secondary antibody and streptavidin-peroxidase solutions (Invitrogen-Histostain plus Bulk Kit) were applied for 30 min. The DAB-H2O2 (diaminobenzidinehidrogenperoxide) [27] was applied to the tissues washed by the PBS as an encolouring substrate. The hematoxylin was applied on the slides for counterstaining after washing with the distilled water. The slides were then dehydrated and covered by the immunmount. The density of immunoreactivity was evaluated semi-quantitatively depending on the severity of reaction and density in the tissue. The evaluation was made by two independent observers. Depending on the staining properties, the slides were scored within the range of 0-3 during their evaluation: weak staining (1), intermediate staining (2), and intense staining (3) [28,29]. All procedures were performed in the same way without adding the primary antibody to the slides in the negative control group. The slides prepared for the histological and immunohistochemical investigations were evaluated and photographed by a light microscope (Olympus BX51, Japan). The number of NGF and Trk-A immunoreactivity positive cells was counted by using the image-j (vI.50i) software. The numerical distribution of NGF and Trk-A positive cells was observed in six different sections chosen from eight unit areas of the islets of Langerhans and acinar cells for each animal [30].

### 2. 2. 4. Statistical analysis

The SPSS (v.20.0) package software was used to evaluate the data obtained in the study. Kruskal–Wallis H test was performed to determine the differences between the groups (control, sham, cinnamon, diabetes, diabetes + cinnamon) for the evaluation of live weight and blood glucose data. Mann–Whitney U test was used to compare the differences between the significant groups. Wilcoxon test was used to determine the differences between the live weights of the rats in the groups on the 1st day and the 17th day. In order to determine the differences between the blood glucose levels of the rats on the 1st, 3rd, and 17th days, the Friedman test was performed first and then the Wilcoxon test between the measurements. The Chi-square test was used to determine the difference between the groups on account of the NGF and Trk-A immunoreactivity positive cells. Results are expressed as median (min–max). P values < 0.05 were considered statistically significant.

#### 3. Results

#### 3. 1. Live weight results

The live weight of male and female rats was statistically evaluated within and between the groups and the results were given as tables and figures. The female rats in the diabetes and diabetes+cinnamon groups did not show any statistically significantly difference on the 17th day of the research in compared to the live weight averages (p > 0.001). However, at the end of the research, the weight averages of diabetes and diabetes+cinnamon groups showed a statistically significant decrease compared to the first-day results (p < 0.001) (Table 1, Figure 1). The male rats in the diabetes and diabetes+cinnamon groups showed a statistically significant difference on the 17th day of the research in compared to the live weight averages (p < 0.001). The weight averages of both groups showed a statistically significant decrease on the 17th day in compared to the 1st day. It was observed that the live weight of rat decreased significantly in the diabetes+cinnamon group (Table 2, Figure 2).

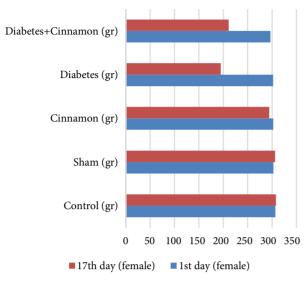
#### 3. 2. Blood glucose levels

The blood glucose levels of male and female rats were statistically evaluated within and between the groups and the results were given as tables and figures. The female rats in the diabetes and diabetes+cinnamon groups did not show any statistically significant difference between the 3rd and 17th days in blood glucose levels. But, it was observed that the 17th day average blood glucose levels in the diabetes+cinnamon group decreased though they were not directly reflected by the statistics (Table 3, Figure 3). The male rats in the diabetes group did not show any statistically significant difference between the 3rd and 17th days in blood glucose levels. However, it was observed that the average blood glucose levels. However, it was observed that the average blood glucose levels in the diabetes+cinnamon group decreased significantly on the 17th day (p < 0.001) (Table 4, Figure 4).

Table 1. The statistical evaluation of live weights of the female rats according to the groups.

Days	Control (g)	Sham (g)	Cinnamon (g)	Diabetes (g)	Diabetes+ Cinnamon (g)
1st day	305.5 (304-315) <sup>aA</sup>	297.5 (289–324) <sup>aA</sup>	296.5 (285-338) <sup>aA</sup>	312.0 (276-317) <sup>aA</sup>	297.5 (289-302) <sup>aA</sup>
17th day	305.0 (296-339) <sup>aA</sup>	303.5 (296-322) <sup>aA</sup>	291.0 (277-324) <sup>aB</sup>	198.5 (151-236)ыв	210 (200-226)ыв

A, B: As a result of Wilcoxon test; the differences between the averages in the same column expressed with the different letters are statistically significant (p < 0.001), a, b: As a result of the Kruskal–Wallis H test and the Mann–Whitney U test performed afterwards; the differences between the averages in the same line expressed with the different letters are statistically significant (p < 0.001).



Body Weight Changes of Female Rats

Figure 1. The body weight changes of female rats.

#### 3. 3. Histological results

The islets of Langerhans, acinar, pars initialis, pars excretoria, ductus excretorius, and blood vessels were seen at the pancreatic tissues of female and male rats. The pancreatic tissue of male and female rats was seen in normal histomorphology belonging to the control, sham and cinnamon groups (Figure 5A-C and Figures 6A-C). A significant decrease in the cell density and a particular decrease in the peripheral density were observed in the islets of Langerhans of the pancreatic tissue of female and male rats in the diabetes groups (Figure 5D, Figure 6D). The acinar of the pancreatic tissue of male and female rats in the diabetes+cinnamon groups was observed as normal and the cell density of islets of Langerhans showed a view close to normal (Figures 5E, Figure 6E).

## 3. 4. Immunohistochemical results

#### 3. 4. 1. NGF Immunoreactivity

The NGF immunoreactivity was determined in the endocrine and exocrine parts of the pancreatic tissue of

female and male rats in all groups. A moderate diffuse cytoplasmic NGF immunoreactivity was observed in the epithelium cells of acinar, pars initialis, pars excretoria, and ductus excretorius but a strong diffuse cytoplasmic NGF immunoreactivity was detected in the islets of Langerhans on the control, sham and cinnamon groups in the female and male rats. A weak diffuse cytoplasmic NGF immunoreactivity was detected in the islets of Langerhans of diabetes group whereas a moderate diffuse cytoplasmic NGF immunoreactivity was detected in the diabetes+cinnamon group in the female (Table 5, Figure 7) and male rats (Table 6, Figure 8). The count of NGF immunoreactivity positive cells, acinar and islets of Langerhans of groups was presented in Tables 7 and 8.

### 3. 4. 2. Trk-A immunoreactivity

A strong granular Trk-A immunoreactivity was determined in the cytoplasm of acinar cells and a weak diffuse cytoplasmic Trk-A immunoreactivity was detected in the islets of Langerhans of the pancreatic tissue of female and male rats in the control, sham and cinnamon groups. The Trk-A immunoreactivity in the acinar cells of pancreatic tissues and pancreatic duct in the diabetes and diabetes+cinnamon groups and the female and male rats showed similar properties with the control group. Apart from the control group, the Trk-A immunoreactivity was not detected in the islets of Langerhans in the diabetes and diabetes+cinnamon groups (Tables 5 and 6, Figures 9 and 10). The count of Trk-A immunoreactivity positive in the acinar cells and islets of Langerhans of groups was presented in Tables 9 and 10.

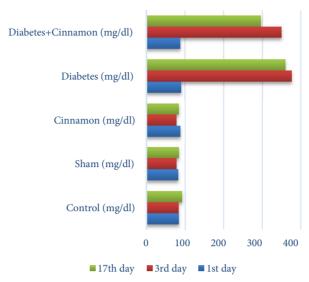
#### 4. Discussion

The cinnamon was determined to regulate the adipocyt gene expression in the mouse, decrease the body weight and body lipid mass in the obese female rats, regulate the liver enzymes and lipid levels to the normal levels, increase the serum insulin levels, and decrease the glucose levels [31,32]. It was stated that the use of food additives containing the cinnamon decreased the fasting blood glucose, body mass index, body weight, and lipid mass significantly in the obese, overweight prediabetic, and type-

Table 2. The statistical evaluation of live weights of the male rats according to the groups.

Days	Control (g)	Sham (g)	Cinnamon (g)	Diabetes (g)	Diabetes+ Cinnamon (g)
1st day	421 (414-449) <sup>Aa</sup>	425 (413-436) <sup>Aa</sup>	429 (424-436) <sup>Aa</sup>	436 (428–443) <sup>Aa</sup>	420 (400–437) <sup>Aa</sup>
17th day	441 (422-450) <sup>Aa</sup>	439 (425–450) <sup>Ba</sup>	420 (406–437) <sup>Ab</sup>	404 (324–325) <sup>Bb</sup>	307.5 (261-354) <sup>Bc</sup>

A, B: As a result of Wilcoxon test; the differences between the averages in the same column expressed with the different letters are statistically significant (p < 0.05), a, b, c: As a result of the Kruskal–Wallis H test and the Mann–Whitney U test performed afterwards; the differences between the averages in the same line expressed with the different letters are statistically significant (p < 0.05).



Changes in Fasting the Blood Sugar of Female Rats

Figure 2. The body weight changes of male rats.

Table 3. The statistical evaluation of fasting blood glucose level of the female rats according to the groups.

Days	Control (mg/dL)	Sham (mg/dL)	Cinnamon (mg/dL)	Diabetes (mg/dL)	Diabetes+ Cinnamon (mg/dL)
1st day	83 (80–91) <sup>aB</sup>	84 (75-90) <sup>aAB</sup>	85.5 (78–102) <sup>aA</sup>	92 (84–94) <sup>aB</sup>	87.5 (82–98) <sup>aB</sup>
3rd day	84.5 (79-87) <sup>aB</sup>	77 (73–87) <sup>bA</sup>	77 (73-82)ыв	380.5 (288–427) <sup>сА</sup>	328.5 (320-397) <sup>cA</sup>
17th day	92 (88–95) <sup>aA</sup>	84 (78–91) <sup>bB</sup>	83 (76–90) <sup>bA</sup>	362.5 (280-400) <sup>cA</sup>	308.5 (158–383) <sup>cA</sup>

A, B: As a result of the Friedman test and the Wilcoxon test performed afterwards; the differences between the levels in the same column expressed with the different letters are statistically significant (p < 0.05), a, b, c: As a result of the Kruskal–Wallis H test and the Mann–Whitney U test performed afterwards; the differences between the levels in the same line expressed with the different letters are statistically significant (p < 0.05).

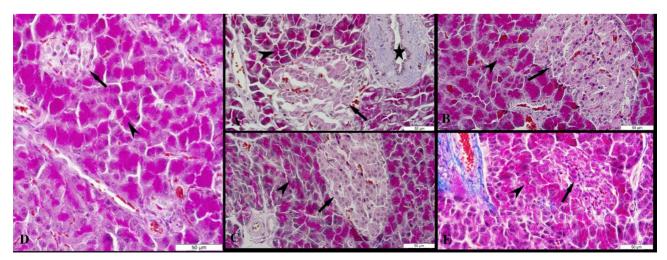


Figure 3. The changes in fasting the blood sugar of female rats.

Days	Control (mg/dL)	Sham (mg/dL)	Cinnamon (mg/dL)	Diabetes (mg/dL)	Diabetes+ Cinnamon (mg/dL)
1st day	76.5 (67–85) <sup>bA</sup>	77.5 (72-80) <sup>bA</sup>	90.5 (75–97)ªA	83.5 (82–91) <sup>aB</sup>	88.5 (77–97) <sup>aC</sup>
3rd day	75.5 (66–86) <sup>cA</sup>	77 (74–84) <sup>cA</sup>	76.5 (72–84) <sup>cA</sup>	373.5 (358-400) <sup>aA</sup>	338 (295-359) <sup>bA</sup>
17th day	82 (70-86) <sup>cA</sup>	80.5 (70-82) <sup>cA</sup>	79.5 (73–81) <sup>cA</sup>	359 (346-388) <sup>aA</sup>	264.5 (95-326) <sup>bB</sup>

Table 4. The statistical evaluation of fasting blood glucose level of the male rats according to the groups.

A, B, C: As a result of the Friedman test and the Wilcoxon test performed afterwards; the differences between the levels in the same column expressed with the different letters are statistically significant (p < 0.05), a, b, c: As a result of the Kruskal–Wallis H test and the Mann–Whitney U test performed afterwards; the differences between the levels in the same line expressed with the different letters are statistically significant (p < 0.05).

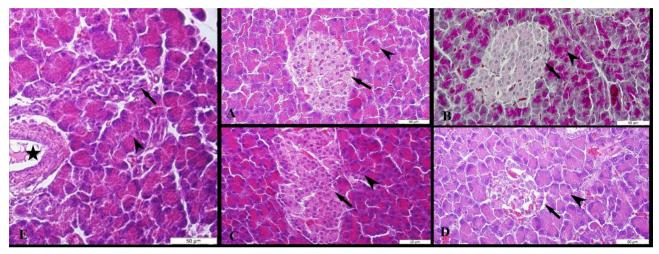


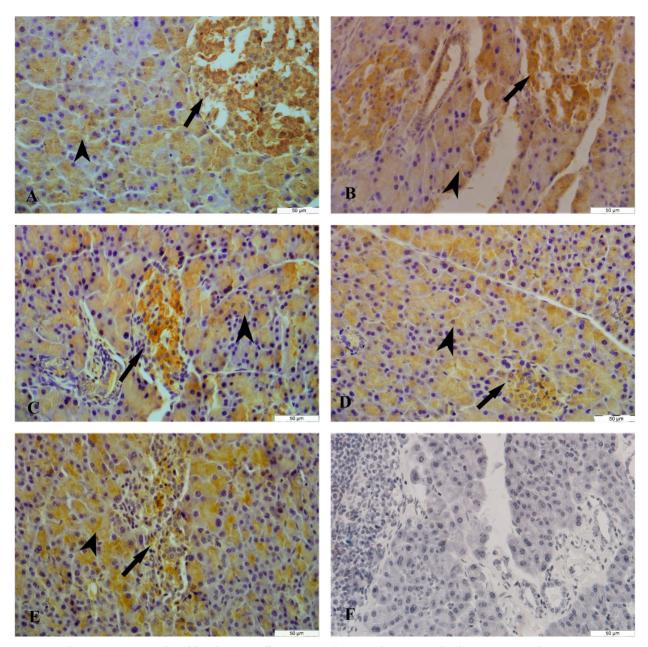
Figure 4. The changes in fasting the blood sugar of male rats.

**Table 5.** The semiquantitative analysis results of NGF and Trk-A immunoreactivity in the pancreatic tissue of female rats.

NGF immunoreactivity					
Area	Control n:6	Sham n:6	Cinnamon n:6	Diabetes <i>n</i> :6	Diabetes+ cinnamon <i>n</i> :6
Acini	2	2	2	2	2
Pars initialis	2	2	2	2	2
Pars excretoria	2	2	2	2	2
Ductus excretorius	2	2	2	2	2
Langerhans islets	3	3	3	1	2
Trk-A immunoreactivity					
Area	Control n:16	Sham n:16	Cinnamon n:6	Diabetes n:6	Diabetes+ cinnamon n:6
Acini	3	3	3	3	3
Langerhans islets	1	1	1	0	0

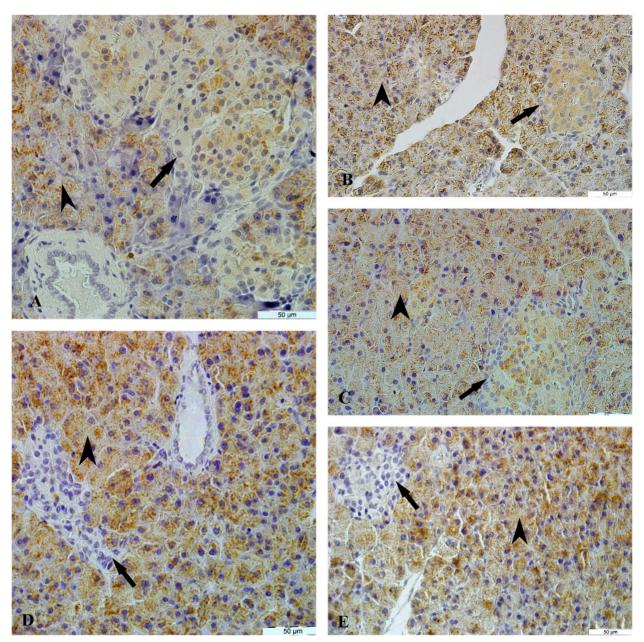
2 diabetic male and female patients [33,34]. In our research, the female rats in the diabetes and diabetes+cinnamon groups did not show any statistically significantly

difference on the 17th day of the research with regards to the live weight averages (p > 0.001). At the end of the research, both groups showed a statistically significantly



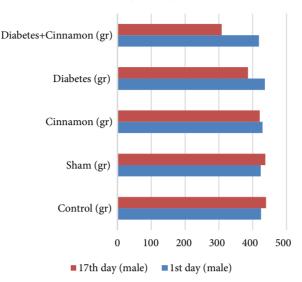
**Figure 5.** The microscopic results of female rats in all groups. A: The control group, B: The sham group, C: The cinnamon group, D: The diabetes group, E: The diabetes+cinnamon group. Arrow: The islets of Langerhans, arrowhead: The acinar, star: The pars excretoria. A, B, C, E: The triple staining, D: The Hematoxylin & Eosin staining.

decrease with regard to the first-day results. The male rats in the diabetes and diabetes+cinnamon groups showed a statistically significantly difference on the 17th day of the research with regards to the live weight averages (p <0.001). We observed a decrease in average in the male rats, especially in the diabetes+cinnamon group. A significant level of differences in the results obtained from the male rats made us think that the application of cinnamon might have an effect on decreasing the live weight in the male rats. In addition, the cinnamon application was thought to contribute positively to the treatment and recovery process, especially by providing the weight control in the diabetic male patients. The cinnamon was pointed out to decrease the high levels of blood glucose in the female rats, regulate the lipid metabolism, repress the blood sugar by decelerating the absorption of carbohydrates from the intestines, and have a healing role by showing the insulinlike effect in the type-2 diabetes disease [35,36]. Ranasinghe et al. [37] deemed that the cinnamon had positive effects as a result of the decrease in the high blood glucose and



**Figure 6.** The microscopic results of male rats in all groups. A: The control group, B: The sham group, C: The cinnamon group, D: The diabetes group, E: The diabetes+cinnamon group. Arrow: The islets of Langerhans, arrowhead: The acinar, star: The pars excretoria. A, B, C, E: The triple staining, D: The Hematoxylin & Eosin staining.

the absence of any histochemical toxic effect in the kidney and pancreas tissues after a single dose cinnamon extract treatment in the male and female rats. They came up with the suggestion that the application dose of cinnamon was unimportant. The decrease in the high blood sugar level in the male rats in the diabetes and diabetes+cinnamon group showed similarities with some results in the literature [37]. Our results made us think that the antidiabetic effect of cinnamon might differ depending on the sex. In the histopathological evaluation of pancreatic tissues of the rats in the streptozotocin mediated diabetes, the results such as degeneration in the  $\beta$  cells, damage and necrosis were detected [38]. In our study no pathological results were detected in female and male rats of the cinnamon group. We thought that the cinnamon application might have roles in the cellular regeneration and tissue repair because of the decrease in the islets of Langerhans density in the male and female rats in the diabetes group and



Body Weight Changes of Male Rats

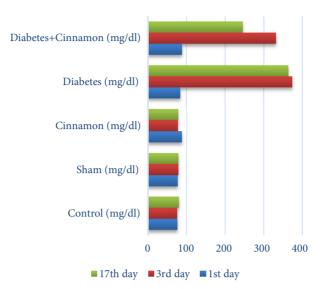
**Figure 7.** The NGF immunoreactivity in the pancreas tissue of female rats. A: The control group, B: The sham group, C: The cinnamon group, D: The diabetes group, E: The diabetes+ cinnamon group, F: The negative control. Arrow: The islets of Langerhans, arrowhead: The acinar.

NGF immunoreactivi	ty				
Area	Control n:6	Sham n:6	Cinnamon n:6	Diabetes n:6	Diabetes+ cinnamon n:6
Acini	2	2	2	2	2
Pars initialis	2	2	2	2	2
Pars excretoria	2	2	2	2	2
Ductus excretorius	2	2	2	2	2
Langerhans islets	3	3	3	1	2
Trk-A immunoreactiv	ity				
Area	Control n:16	Sham <i>n:16</i>	Cinnamon <i>n</i> :6	Diabetes n:6	Diabetes+ cinnamon <i>n:6</i>
Acini	3	3	3	3	3
Langerhans islets	1	1	1	0	0

Table 6. The semiquantitative analysis results of NGF immunoreactivity in the pancreatic tissue of male rats.

the detection in a similar level for the cell density in the control group and diabetes+cinnamon group in our study.

The NGF was detected in the primary cell cultures to be synthesized from the  $\beta$  cells of the pancreatic tissue of adult and fetus. The NGF was known to arrange the sensitivity of  $\beta$  cells and stimulate the nerve cell development [39]. The expression of Trk-A and its ligand NGF in the pancreas during the embriyonic and fetal life suggests that the NGF and its receptor could play an important role in the development of pancreatic tissue [40]. In our research, the Trk-A immunoreactivity was determined in the cytoplasm of acinar cells and islets of Langerhans of the pancreatic tissue of female and male rats in the control, sham, and cinnamon groups. The NGF immunoreactivity was determined in the endocrine and exocrine parts of the pancreatic tissue of female and male rats in all groups. When the immunohistochemical distribution of NGF and Trk-A receptor in the pancreatic tissue was examined, it was determined that there was no difference between the sexes in terms of the severity of immunoreactivity,



Changes in Fasting the Blood Sugar of Male Rats

**Figure 8.** The NGF immunoreactivity in the pancreas tissue of male rats. A: The control group, B: The sham group, C: The cinnamon group, D: The diabetes group, E: The diabetes+cinnamon group. Arrow: The islets of Langerhans, arrowhead: The acinar.

**Table 7.** The comparison of count of the NGF immunoreactivity positive cells acini and Langerhans island among the groups in the female rats.

Section	Control	Sham	Cinnamon	Diabetes	Diabetes+Cinnamon
Acinar cell	1474 (-)	1475 (-)	1474 (-)	1474 (-)	1475 (-)
Langerhans islets cell	188ª	188 <sup>a</sup>	188 <sup>a</sup>	69 <sup>b</sup>	135 °
Number (unit area)	48	48	48	48	48

-; There is no statistical difference between the groups.

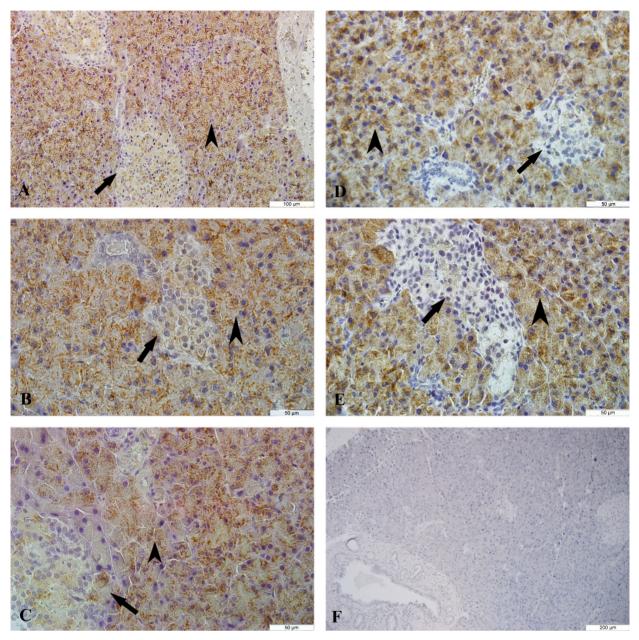
The unit area: The total number of regions on which the slides were counted for the immunoreactivity positive cells. a,b,c; The differences superscripts in the same line indicate the significant differences between the groups of islets of Langerhans (p = 0.000, p < 0.001).

**Table 8.** The comparison of count of the NGF immunoreactivity positive cells acini and Langerhans island among the groups in the male rats.

Section	Control	Sham	Cinnamon	Diabetes	Diabetes+Cinnamon
Acinar cell	1177 (-)	1176 (-)	1177 (-)	1177 (-)	1176 (-)
Langerhans islets cell	248 ª	248 ª	249 <sup>a</sup>	49 <sup>b</sup>	128 °
Number (unit area)	48	48	48	48	48

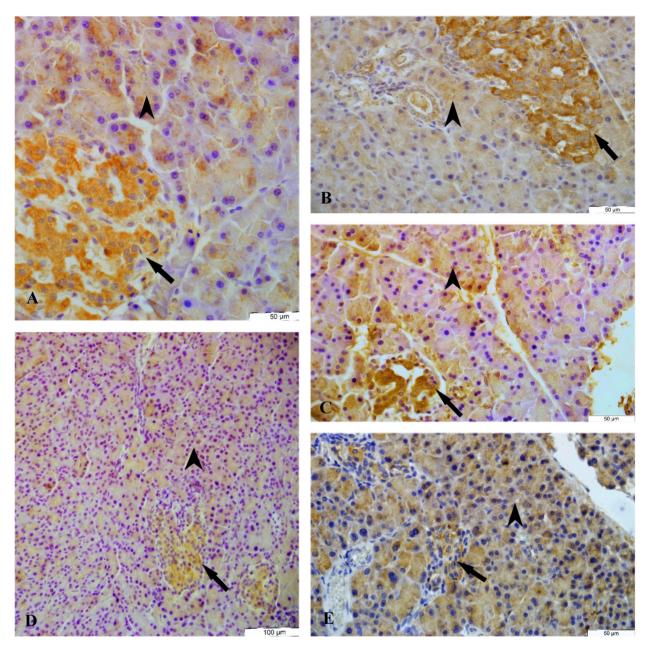
-; There is no statistical difference between the groups.

The unit area: The total number of regions on which the slides were counted for the immunoreactivity positive cells. a,b,c; The differences superscripts in the same line indicate significant differences between the groups islets of Langerhans (p = 0.000, p < 0.001).



**Figure 9.** The Trk-A immunoreactivity in the pancreas tissue of female rats. A: The control group, B: The sham group, C: The cinnamon group, D: The diabetes group, E: The diabetes+cinnamon group. Arrow: The islets of Langerhans, arrowhead: The acinar.

the regions in which it was seen. These results made us conclude that the NGF and Trk-A might have roles in providing the persistence of cell signals maintaining the survival of exocrine and endocrine pancreatic cells and they might also have roles in the synthesis and secretion of hormones in these regions. Schneider et al. [41] investigating the distribution of NGF and Trk-A in the pancreatic tissue revealed that there was a moderate diffuse cytoplasmic reaction in the exocrine pancreas, a weak diffuse cytoplasmic reaction in the nerve cords and no NGF reaction in the endocrine pancrease [41]. The Trk-A immunoreactivity in the pancreatic duct and tunica muscularis layer of arteries was stated to be weak, but the immunoreactivity in the islet cells was moderate [41]. In another research, the NGF immunoreactivity in the pancreatic tissue was notified as weak in the acinar cells and nerve cords and as moderate in the arteries and vein walls. The chronic pancreatitis patients showed increasing NGF and Trk-A immunoreactivity. The Trk-A immunoreactivity was reported to be moderate only in the nerve fibre, arterial, and venous walls. It was stated that the NGF/Trk-A pathway interactions might lead to the morphological



**Figure 10.** The Trk-A immunoreactivity in the pancreas tissue of male rats. A: The control group, B: The sham group, C: The cinnamon Group, D: The diabetes group, E: The diabetes+cinnamon group, F: The negative control. Arrow: The islets of Langerhans, arrowhead: The acinar.

changes and pain syndromes [42]. In our research where the diffuse cytoplasmic NGF immunoreactivity occurred at the acinus, pancreatic duct and islets of Langerhans of pancreatic tissue of the rats. We detected the Trk-A immunoreactivity in the acinus and islets of Langerhans. The granular property of Trk-A immunoreactivity in the acinar cells was similar to the literature data, but the diffuse cytoplasmic Trk-A immunoreactivity of islets of Langerhans was dissimilar to the recent literature [41]. Although the region where the Trk-A immunoreactivity was observed was the same as the literature, the type of reaction was different from the literature. This data made us think that there might be differences between the density of immunoreactivity and the regions were as they were seen.

On the evaluation of histologic investigation of the pancreatic tissue, after the diabetes was formed, the research suggested that the decrease of cell number in the islets of Langerhans, NGF and Trk-A protein levels was as it was in the western blot analysis [43]. The live cell numbers after the NGF blockage and insulin signals in the rat pancreatic

Section	Control	Sham	Cinnamon	Diabetes	Diabetes+Cinnamon
Acinar cell	1400 (-)	1399 (-)	1400 (-)	1399 (-)	1400 (-)
Langerhans islets cell	183ª	184 <sup>a</sup>	183 <sup>a</sup>	0 <sup>b</sup>	0 <sup>b</sup>
Number (unit area)	48	48	48	48	48

**Table 9.** The comparison of count of the Trk-A immunoreactivity positive cells acini and Langerhans island among the groups in the female rats.

-; There is no statistical difference between the groups.

The unit area: The total number of regions on which the slides were counted for the immunoreactivity positive cells. a,b,c; The differences superscripts in the same line indicate the significant differences between the groups of islets of Langerhans (p = 0.000, p < 0.001).

**Table 10.** The comparison of count of the Trk-A immunoreactivity positive cells acini and Langerhans island among the groups in the male rats.

Section	Control	Sham	Cinnamon	Diabetes	Diabetes+Cinnamon
Acinar cell	1055 (-)	1054 (-)	1054 (-)	1054 (-)	1055 (-)
Langerhans islets cell	233 ª	232 ª	232 ª	0 в	0 <sup>b</sup>
Number (unit area)	48	48	48	48	48

-; There is no statistical difference between the groups.

The unit area: The total number of regions on which the slides were counted for the immunoreactivity positive cells. a,b,c; The differences superscripts in the same line indicate the significant differences between the groups of islets of Langerhans (p = 0.000, p < 0.001).

tissue were decreased compared with the control groups. Apoptosis occurred in 17% of the cells cultured in the glucose for 16 h and apoptosis was partially inhibited with the NGF and insulin [44]. Similarly, we detected a decrease in the cellularity of islets of Langerhans for the female and male rats in the diabetes group, a significant decrease in the NGF immunoreactivity and total disappearance of Trk-A immunoreactivity. These findings made us think that the streptozocin application might cause changes in the NGF and Trk-A levels by the islet cell damage and cellular damage occurring in the islets of Langerhans in the diabetes disease that negatively affected the Trk-A receptor level. In healthy individuals, the NGF was secreted and synthesized from the pancreatic  $\beta$  cells and regulated the Na duct density by the paracrine and autocrine pathways [45]. In the cell tissue culture study, the NGF levels were stated to increase three-fold when the  $\beta$  cells of female rats were treated in the glucose culture [46] and the Trk-A mRNA numbers were stated to increase 6-fold when they were applied to the NGF culture [47]. In addition, an increase in the insulin secretion depending on the increase of NGF secretion was notified [47]. In our research, we detected that the NGF immunoreactivity decreased in the islets of Langerhans of rats in the diabetes groups and increased in diabetes+cinnamon groups. The Trk-A immunoreactivity was unchanged. It might be due to the disruption of NGF

and Trk-A signal pathways in the diabetes disease. It was concluded that the Trk-receptor immunoreactivity did not change in the diabetes and diabetes+cinnamon groups. Our results suggested that the cinnamon led to an increase in the NGF immunoreactivity in the islet cells and the NGF might have a role in the regulation of blood glucose in the diabetes. In addition, it was also concluded that the increase in the level of NGF could contribute positively to the prevention of diabetes-related diseases.

Based upon these data, we concluded that the cinnamon is effective in reducing the high blood glucose, significantly decrease the live weight in the diabetic male rats, change the Trk-A levels, and it also causes an increase in the NGF levels in the islets of Langerhans and it may have positive effects by increasing the NGF levels on the treatment of diabetes disease and by hindering the complications depending on the diabetes.

#### Acknowledgement and/or disclaimers, if any

This study was supported by the Scientific and Technical Research Council of the Kafkas University (Project 2015-VF-04), Kars-TURKEY. The present study was summarized from a Ph.D Thesis and was presented at the 15th International Congress of Histochemistry and Cytochemistry (ICHC 2017), 18-21 May 2017 in Kervansaray Lara Hotel Antalya, Turkey.

### References

- İrak K, Mert N, Mert H, Ayşin N. The effects of grape seed extract on the some enzymes and metabolites in diabetic rats. Van Veterinary Journal 2018; 29 (3): 147-152.
- Pipeleers D, Hoorens A, Marichal-Pipeleers M, Van de Casteele M, Bouwens L et al. Role of pancreatic cells in the process of cell death. Diabetes 2001; 50 (1): 52-57. doi: 10.2337/ diabetes.50.2007.s52
- Jayaprakasha GK, Rao LJM. Chemistry, biogenesis, and biological activities of Cinnamomum zeylanicum. Critical Reviews in Food Science and Nutrition 2011; 51: 547-562. doi: 10.1080/10408391003699550
- Shreaz S, Wani WA, Behbehani JM, Raja V, Irshad M et al. Cinnamaldehyde and its constituents, a novel class of antifungal agents. Fitoterapia 2016; 12: 116-131. doi: 10.1016/j. fitote.2016.05.016
- Rao PV, Gan SH. Cinnamon: a multifaceted medicinal plant. Evidence Based Complement Alternative Medicine 2014. doi: 10.1155/2014/642942
- Jia Q, Liu X, Wu X, Wang R, Hu X et al. Hypoglycemic activity of a polyphenolic oligomer-rich extract of cinnamomum parthenoxylon bark in normal and Streptozotocin induced diabetic rats. Pythomedicine 2009; 16: 744-750. doi: 10.1016/j. phymed.2008.12.012
- Kumar S, Vasudeva N, Sharma S. GC-MS analysis and screening of antidiabetic, antioxidant and hypolipidemic potential of cinnamomum tamala oil in streptozotocin induced diabetes mellitus in rats. Cardiovascular Diabetology 2012; 10; 11: 95. doi: 10.1186 / 1475-2840-11-95
- Reichardt FL. Neurotrophin-regulated signalling pathways. Philosophical transactions of the Royal Society B 2006; 361: 1545-1564. doi: 10.1098/rstb.2006.1894
- Bayar M, Ozer B, Bestas A, Ceribasi S, Ozercan I. Effects of anti-NGF on apoptosis in rats with experimentally induced sepsis model. Turkiye Klinikleri Journal of Medical Sciences 2010; 30 (4): 1127-1133. doi: 10.5336 / medsci.2009-11654
- Schäper C, Gläser S, Groneberg DA, Kunkel G, Ewert R et al. Nerve growth factor synthesis in human vascular smooth muscle cells and its regulation by dexamethasone. Regulatory Peptides 2009; 157 (1-3): 3-7. doi: 10.1016/j.regpep.2009.07.004
- Ieda M, Fukuda K, Hisaka Y, Kimura K, Kawaguchi H et al. Endothelin-1 regulates cardiac sympathetic innervation in the rodent heart by controlling nerve growth factor expression. The Journal of Clinical Investigation 2004; 113 (6): 876-884. doi: 10.1172/JCI19480
- Murase K, Hattori A, Kohno M, Hayashi K. Stimulation of nerve growth factor synthesis/secretion in mouse astroglial cells by coenzymes. Biochemistry and Molecular Biology International 1993; 30 (4): 615-621.
- Johnson Jr EM, Gorin PD, Brandeis LD, Pearson J. Dorsal root ganglion neurons are destroyed by exposure in utero to maternal antibody to nerve growth factor. Science 1980; 210: 916-918. doi: 10.1126/science.7192014

- Trim N, Morgan S, Evans M, Issa R, Fine D et al. Hepatic stellate cells express the low affinity nevre growth factor receptor p75 and undergo apoptosis in response to nevre growth factor stimulation. The American Journal of Pathology 2000; 156 (4): 1235-1243. doi: 10.1016/S0002-9440(10)64994-2
- 15. Dang C, Zhang Y, Ma Q, Shimahara Y. Expression of nerve growth factor receptors is correlated with progression and prognosis of human pancreatic cancer. Journal of Gastroenterology and Hepatology 2006; 21: 850-858. doi: 10.1111/j.1440-1746.2006.04074.x
- 16. Gelincik A, Aydın F, Ozerman B, Ergüven M, Aydın S et al. Enhanced nerve growth factor expression by mast cells does not differ significantly between idiopathic and allergic rhinitis. Annals of Allergy, Asthma & Immunology 2012; 108 (6): 396-401. doi:10.1016/j.anai.2012.04.006
- Artico M, Bronzetti E, Felici LM, Alicino V, Ionta B et al. Neurotrophins and their receptors in human lingual tonsil: an immunohistochemical analysis. Oncology Reports 2008; 20 (5): 1201-1206.
- Levanti MB, Germanà A, Catania S, Germanà GP, Gauna-Añasco L et al. Neurotrophin receptor-like proteins in the bovine (Bos taurus) lymphoid organs, with special reference to thymus and spleen. Anatomia, Histologia, Embryologia 2001; 30 (4): 193-198. doi: 10.1046/j.1439-0264.2001.00329.x
- Chaldakov GN, Tonchev AB, Aloe L. NGF and BDNF: from nerves to adipose tissue, from neurokines to metabokines. Relevance to neuropsychiatric and cardiometabolic diseases. Rivista di Psichiatria 2009; 44 (2): 79-87.
- Sornelli F, Fiore M, Chaldakov GN, Aloe L. Adipose tissuederived nerve growth factor and brain-derived neurotrophic factor: results from experimental stress and diabetes. General Physiology and Biophysics 2009; 28: 179-183.
- 21. Davies AM, Lumsden AGS, Rohrer H. Neural crest-derived proprioceptive neurons express NGF receptors but is not supported by NGF in culture. Neuroscience 1987; 20: 37-46. doi: 10.1016/0306-4522(87)90004-2
- 22. Jia Q, Liu X, Wu X, Wang R, Hu X et al. Hypoglycemic activity of a polyphenolic oligomer-rich extract of cinnamomum parthenoxylon bark in normal and Streptozotocin induced diabetic rats. Pythomedicine 2009; 16: 744-750. doi: 10.1016/j. phymed.2008.12.012
- 23. Ganda OP, Rossi AA, Like AA. Studies on streptozotocin diabetes. Diabetes 1976; 25: 595-603.
- Sharafeldin K, Rizvi RM. Effect of traditional plant medicines (cinnamomum zeylanicum and syzygium cumini) on oxidative stress and insulin resistance in streptozotocin-induced diabetic rats. The Journal of Basic & Applied Zoology 2015; 72: 126-134. doi:10.1016/j.jobaz.2015.09.002
- 25. Luna LG. Manual of histologic staining methods of the armed forces institute of pathology. 3rd ed. New York: Blakiston Division, McGraw-Hill; 1968.

- Hsu SM, Raine L, Fanger H. Use of avidin-biotin-peroksidase complex (ABC) in immunoperoksidase techniques: a comparison between ABC and unabeled antibody (PAP) procedures. Journal of Histochemistry & Cytochemistry 1981; 29: 577-580. doi: 10.1177/29.4.6166661
- Shu S, Ju G, Fan LZ. The glucose oxidase-DAB-nickel method in peroxidase histochemistry of the nervous system. Neuroscience Letters 1988; 85: 169-171. doi: 10.1016/0304-3940(88)90346-1
- Zhu QY. Analysis of blood vessel invasion by cells of thyroid follicular carcinoma using image processing combined with immunohistochemistry. Zhonghua Yi Xue Za Zhi 1989; 69: 573-575.
- 29. Seidal T, Balaton AJ, Battifora H. Interpretation and quantification of immunostains. The American Journal of Surgical Pathology 2001; 25: 1204-1207. doi: 10.1097/00000478-200109000-00013
- Eliş Yıldız S, Yediel Aras Ş, Dağ S, Karadağ Sarı E. Immunohistochemical distribution of somatostatin in gastric tissue of diabetic rats treated with cinnamon extracts. Kafkas Universitesi Veteriner Fakultesi Dergisi 2019; 25 (3): 427-433. doi: 10.9775/kvfd.2018.21175
- Cao H, Graves DJ, Anderson RA. Cinnamon extract regulates glucose transporter and insülin-signaling gene expression in mouse adipocytes. Phytomedicine 2010; 17: 1027-1032. doi: 10.1016/j.phymed.2010.03.023
- Shalaby MA, Saifan HY. Some pharmacological effects of cinnamon and ginger herbs in obese diabetic rats. Journal of Intercultural Ethnopharmacology 2014; 3 (4): 144-149. doi: 10.5455/jice.20140818050741
- 33. Vafa M, Mohammadi F, Shidfar F, Sormaghi MS, Heidari I et al. Effects of cinnamon consumption on glycemic status, lipid profile and body composition in type 2 diabetic patients. International Journal of Preventive Medicine 2012; 3: 531-536.
- 34. Liu Y, Cotillard A, Vatier C, Bastard JP, Fellahi S et al. A dietary supplement containing cinnamon, chromium and carnosine decreases fasting plasma glucose and increases lean massin overweightor obese pre-diabetic subjects: a randomized, placebo-controlled trial. Plos One 2015; 10 (9): e0138646. doi: 10.1371 / journal.pone.0138646
- Kim H, Hyun HS, Choung YS. Anti-diabetic effect of cinnamon extract on blood glucose in db/db mice. Journal of Ethnopharmacology 2006; 104: 119-123. doi: 10.1016/j. jep.2005.08.059
- Cheng MD, Khun P, Poulev A, Rojo LE, Lila AM et al. In vivo and in vitro anti diabetic effects of aqueous cinnamon extract and cinnamon polyphenol-enhanced food matrix. Food Chemistry 2012; 135 (4): 2994-3002. doi: 10.1016/j.foodchem.2012.06.117

- 37. Ranasinghe P, Perera S, Gunatilake M, Abeywardene E, Gunapala N et al. Effects of cinnamon zeylanicum on blood gucose and lipids in a diabetic and healthy rat model. Pharmacognosy Research 2012; 4 (2): 73-79. doi: 10.4103/0974-8490.94719
- Mohammadi J, Naik PR. The histopathologic effects of Morus Alba leaf extract on the pancreas of diabetic rats. Turkish Journal of Biology 2012; 36: 211-216. doi: 10.3906/biy-1008-51
- Polak M, Scharfmann R, Seilheimer B, Eisenbarth G, Dressler D et al. Nerve growth factor induces neuron-like differentiation of an insulin-secreting pancreatic beta cell line. Proceedings of the National Academy of Sciences 1993; 90: 5781-5785. doi: 10.1073/pnas.90.12.5781
- Miralles F, Philippe P, Czernichow P, Scharfmann R. Expression of nerve growth factor and its high-affinity receptor Trk-A in the rat pancreas during embryonic and fetal life. Journal of Endocrinology 1998; 156: 431-439. doi: 10.1677/joe.0.1560431
- Schneider MB, Standop J, Ulrich A, Wittel U, Friess H et al. Expression of nerve growth factors in pancreatic neural tissue and pancreatic cancer. Journal of Histochemistry & Cytochemistry 2001; 49 (10): 1205-1210. doi: 10.1177/002215540104901002
- Friess H, Zhu ZW, Di Mola FF, Kulli C, Graber HU et al. Nerve growth factor and its high-affinity receptor in chronic pancreatitis. Annals of Surgery 1999; 230 (5): 615-624. doi: 10.1097/0000658-199911000-00002
- Sposato V, Manni L, Chaldakov GN, Aloe L. Streptozotocininduced diabetes is associated with changes in NGF levels in pancreas and brain. Archives Italiennes de Biologie 2007; 145: 87-97. doi: 10.4449/aib.v145i2.867
- Navarro-Tableros V, Sa'nchez-Soto MC, Garcia S, Hiriart M. Autocrine regulation of single pancreatic b cell survival. Diabetes 2004; 53 (8): 2018-2023. doi: 10.2337/ diabetes.53.8.2018
- 45. Vidaltamayo R, Sánchez-Soto MC, Hiriart M. Nerve growth factor increases sodium channel expression in pancreatic b cells: implications for insulin secretion. Federation of American Societies for Experimental Biology 2002; 16 (8): 891-892. doi: 10.1096/fj.01-0934fje
- Rosenbaum T, Vidaltamayo R, Sánchez-Soto MC, Hiriart MA. Pancreatic b cells synthesize and secrete nerve growth factor. Proceedings of the National Academy of Sciences 1998; 95 (13): 7784-7788. doi: 10.1073/pnas.95.13.7784
- Rosenbaum T, Sánchez-Soto MC, Hiriart MA. Nerve growth factor increases insulin secretion and barium current in pancreatic b-cells. Diabetes 2001; 50 (8): 1755-1762. doi: 10.2337/diabetes.50.8.1755