Tr. J. of Medical Sciences 29 (1999) 105-107 © TÜBITAK

Süleyman BAYKAL Savaş CEYLAN Haydar USUL Yüksel ALİYAZICIOĞLU Hasan EFE Kayhan KUZEYLİ Soner DURU Fadıl AKTÜRK

Received: October 23, 1996

KTU Medical Faculty, Departments of Neurosurgery and Biochemistry Trabzon-Turkey.

Effects of Nerve Growth Factor on Acetylcholinesterase Activity of Injured Spinal Cord

Abstract: Fourteen rats were examined and divided into two groups of 7: the control (Group a) and NGF-treated animals (Group B). All animals received a 50 g clip-compression injury to tthe spinal cord at the T9 level. In the NGF-treated animals, NGF (0.1 mg/kg NGF) was injected subcutaneously for seven days. After spinal cord injury, the mean AChE activity was

41.47 \pm 1.7122 activated substrate min/ml/gr. wet tissue and 47.52 \pm 1.471 in NGF-treated animals. The difference between Groups A and B was statistically significant (p<0.01). These results suggest the possible role of NGF in the cholinergic system.

Key Words: Acetylcholinesterase, Nerve Growth Factor, Spinal Cord Injury.

Introduction

The ongoing search for neurotrophic factors for motoneurone survival and function. Nerve growth factor (NGF) exerts neurotrophic activity after binding to the NGF receptor and being internalised and retrogradely transported to the nuronal cell body as NGF/NGF receptor complex (2). In some studies, it has been demonstrated that NGF has an effect on acetylcholinesterase (AChE) enzyme in both culture mediums (6, 9, 13, 15, 16) and animal experiments (6, 11, 14, 16, 19). Human NGF has perevented the degeneration of basal forebrain cholinergic neurones in primates (15, 17).

In the present study the spinal cord was injured in order to observe the effects of a single dose of NGF on the content of the AChE activities of these tissues on day one and seven.

Materials and Methods

Fourteen winstar rats, weihting 230-300 g, were divided into two groups of 7: the control (Group A) and NGF-treated animals (Group B).

The animals were anaesthetised with ketamin hydrochloride (75mg/kg, intraperitoneally). Under aseptic conditions, a T8-10 laminectomy was performed and the each rat received a 50 gr clip (Aesculap, Germany, Yaşargil Clips) compression injury to the cord with microsurgery at approximately the T9 level. A single dose of NGF (SIGMA Chemical Company, USA) (0.1 mg/kg) was injected in each animal in the NGF-treated group subcutaneously for seven days. In the control animals, equal volumes of serum physiologic were injeced subcutaneously.

Spinal cord tissues were obtained from the groups on the 7th day, 12 hours after the last treatment with placebo or NGF. To determine the levels of AChE, the spinal cord were rapidly removed. They were approximately 2 cm long. All samples were stored at - 70°C until they were assayed.

A ChE activity measurements

The tissues were homogenised in a Potter-Elvenjem homogenizer (20mg tissue for 1 ml, pH 8, 0.1 M phosphate buffer). 0.4 ml of this homognate was placed in a spectrophotometer cuvet containing 2.6 ml (pH, 0.1 M) phosphate buffer. 100µl DTNB (5,5-dithiohis (2-nitrobenzoic aacid), Elmon's Reagent) was added. Absorbency was read at 412 nm. Thus absorbency was set to zero.

2 μ l substrate was added and thhe reaction begun. Changes in absorbency were recorded every minute for at least 6 minutes. Enzyme was calculated using following formula. R=5.74*10⁻⁴* (A/Co) (where R is the rate, A is changes of absorbance per minute and Co is the concentration of the tissue (mg/ml) (3).

Statistical Analysis

AChE activity was expressed as the means and standard deviation of the means of the the results of the

measurements. Statistical analysis of the comparisons between groups was performed using the one-way unpaired t test.

Results

AChE Activity in Contusive Spinal Cords

After spinal cord injury, the mean AChE activity was 41.74 ± 1.722 activated substrate min/ml/gr. wet tissue in the controls and 47.52 ± 1.471 in the NGF-treated animals (Table 1, Fig 1). The difference between two groups was statistically significant (p<0.01).

Table 1. Mean AChE Activity in Spinal Cord Injury (Activated substrate min/ml/gr wet tissue)*

	N. of animals	7th Day
Control Animals	7	41.13±2.99
NGF-treated Animals	7	47.7±5.75

*= Statistically significant (p<0.001).

Discussion

It is a well-known fact that cholinergic neurones are widely distributed in the spinal cord (1, 4, 12). It is also well established that both preganglionic autonomic neurone and somatic motoneurones are cholinergic (1, 4, 10) and make up a significant proportion of the total neuronal poll (1). Acetylcholinesterase, the enzyme that catalyzes a reaction of hydrolysis of acetylcholine to choline and acetate (7), is regarded as specifiic marker for cholinergic function (7, 8, 18) and is used as a differentiation marker (19). Bakhit et al. reported that NGF has a role in the spinal motoneurones. They also determined NGF receptors in rat spinal motoneurones during development and observed that contusion injury of the spinal cord resulted in increased expression of the NGF recopter mRNA (1). Brunello et al. also reported that this NGF receptor mRNA increases in the contused rat spinal cord (2). Femandez et al. infused NGF locally on the

References

1. Bakhit C, Armanini M, Wong WLT, Bennett GL, Wlrathall JR: Increase in nerve growth factor-like immunoreactivity and decrease in choline acetyltransferase following contusive spinal cord injury. Brain research, 554: 264-271, 1991.







,Figure 1. Graph showing AChE Acitvity levels in both groups. This difference is statistically significant (P<0.01).

transected spinal cord in adult rats and observed increased induction of axonal sprouting by histological staining in NGF-treated animals (5).

In one study, it was demonstrated that NGF increases AChE activity and this activity increment reflects the extent of the fiber network of the cholinergic neurones (9). Thus, the extent of the spinal cord injury may correlate with the extent of reduced function of the cholinergic neurones and, hence, in AChE activity.

Our findings show that AChE levels in injured spinal cords are more significantly suppressed the 7 th day in the control animals than in the NGF-treated group. This supports the results of the studies cited above and is in accordance with them. In the NGF-treated animals, some decrease in AChE activity was observed on the 7 th day. The difference in the AChE activities of the control and the NGF-enhancing effects on AChE activity and the possibility of a role for NGF in promoting the recovery of function after spinal cord injury.

- Emfors PE, Henschon A, Olson L, Persson H: Expression of nerve growth factor receptor mRNA is developmentally regulated and increased after axotomy in rat spinal cord motoneurons. Neuron 2: 1605-73, 1989.
- Femandez E, Pallini R, Lauretti L, Mercanti D, Seerra a, Calissano P: Spinal cord transection in adult rats: effects of local infusion of nerve growth factor on the corticospinal tract axons. Neurosurgery, 33: 889-93, 1993.
- Gage FH, Tuszynski MH, Chen KS, Armstrong D, Buzsaki G: Survival, growth and function of damaged cholinergic neurons. EXS. 57:259-74, 1989.
- Ganong WF: Synaptic and Junctional Transmission In: Ganong WF (Ed) Review of Medical Physiology. Lange Medical Book, California, 1991, pp 78-106.
- Gyevai AT, Bartha E: Early and late hormonal modulation of cholinergic maturation in culture of embryonic mesecephali. Biotherapy 5(3): 205-14, 1992.
- Hartikka J. Hefti F: Comparison of nerve growth factor's effects on development of septum, striatum, and nucleus basalis cholinergic neurons in vitro. J. Neurosci. Res. 21: 352-64, 1998.

- He Y, Yao Z, Gu Y, Kuang G, Chen Y: Nerve growth factor promotes collateral sprouting of cholinergic fibers in the septohippocampal cholinergic system of aged rats with fimbria transection. Brain Res. 586(1): 27-35, 1992.
- Houser CR, Crawfor GD, Barber RP, Salveterra PM, Vaughn JE: Organization and morfhological characterization of cholinergic nurons: an immunocytohemical study with monoclonal antibody to choline acetyransferase. Brain Research, 266: 97-119, 1983.
- Kasa P: The cholinergic system in brain and spinal cord. Progr. Neurobiol. 26: 211-72, 1986.
- Kushima Y, Nishio C, Nonomura T, Hatanaka H: Effects of nerve growth and basic fibroblast growth factor on survivcal of cultured septal cholinergic neurons from adult rats. Brain Res. 598(1-2): 264-70, 1992.
- Miyamoto O, Itano T, Fujisawa M, Tokuda M, Matsui H, Nagao S, Hatase O: Exogenous basic fibroblast growth factor and nerve growth factor enhance sprouting of acetylcholinesterase positivi fibers in denervated rat hippocampus. Acta. Med., Okayama, 47(3): 139-44, 1993.

- Nonomura T, Hatanaka H: Neurotrophic effect of brain-derived neurotrophic factor on basal forebrain cholinergic neurons in culture from postnatal rats. Neurosci. Res. 14(3): 226-33, 1992.
- Ojika K, Mitake S, Kamiya T, Kosuge N, Taiji M: Two different molecules NGF and free-HCNP, stimulate cholinergic activity in septal nuclei in vitro in a different manner. Brain. Res. Dev. Brain Res. 79(1): 1-9, 1994.
- 17. Peterson GM, Ginn SR, Lanford GW: Fibers immunoreactive for nerve growth factor receptor in adult rat cortex and hippocampus mimic the innervation pattern of AChE-positive fibers. Brain Res Bull. 33(2): 123-36, 1994.
- Raivich G, Kreuzberg GW: Topography of beta NGF receptor-positive and AChE-reactive neurons in the central nervous system. EXS. 57-50, 1989.
- Yung KK, Tang F, Vacca-Galloway LL : Alterations in acetylcholinesterase and choline acetyltransferase activites and neuropeptide levels in the ventral spinal cord of the wobbler mouse during inherited motoneuron disease. Brain Res. 638(1-2): 337-42, 1994.