

Effects of Dietary Aflatoxin and Hydrate Sodium Calcium Aluminosilicate on Triiodothyronine, Thyroxine, Thyrotrophin and Testosterone Levels in Quails

Gökhan ERASLAN¹, Mehmet AKDOĞAN², Bilal Cem LİMAN¹, Murat KANBUR¹, Namık DELİBAŞ²

¹Department of Pharmacology and Toxicology, Faculty of Veterinary Medicine, Erciyes University, Kayseri - TURKEY

²Department of Biochemistry, Faculty of Medicine, Süleyman Demirel University, Isparta - TURKEY

Received: 30.07.2003

Abstract: This study was performed on 80 male 14-day-old Coturnix coturnix japonica breed quails. The quails were divided into 8 groups with 10 animals in each as 1 control and 7 trial groups. While the control group was fed a commercial basal ration, groups 2-8 received, respectively, 2.5 g/kg feed hydrate sodium calcium aluminosilicate (HSCAS), 5.0 g/kg feed HSCAS, 10.0 g/kg feed HSCAS, 2.5 ppm aflatoxin (AF B₁ 78.30%, AF B₂ 14.60%, AF G₁ 4.50%, AF G₂ 2.60%); 2.5 ppm AF with 2.5 g/kg feed HSCAS, 2.5 ppm AF with 5.0 g/kg feed HSCAS, and 2.5 ppm AF with 10.0 g/kg feed HSCAS, respectively, for 21 days. At the end of the trial, blood samples were taken from the animals and triiodothyronine (T₃), thyroxine (T₄), thyrotrophin (TSH) and testosterone levels in the blood were measured. Statistically significant increases were detected in T₃ levels in groups 3 and 4 and significant decreases in groups 5 and 6, while there were significant increases in T₄ levels in groups 2, 4, 5, 7 and 8 and significant decreases in blood testosterone levels in all trial (groups 2 to 8) groups compared to the control (group 1).

Key Words: Aflatoxin, HSCAS, hormone, quail

Bıldırcınlarda Triiodotironin, Tiroksin, Tirotropin ve Testosteron Düzeyleri Üzerine Yemle Verilen Aflatoksin ve Hidrate Sodyum Kalsiyum Alüminosilikatın Etkisi

Özet: Çalışma, 80 adet, erkek Coturnix coturnix japonica ırkı, 14 günlük bıldırcınlar üzerinde gerçekleştirildi. Bıldırcınlar, biri kontrol diğer yedisi deneme olmak üzere sekiz gruba ayrıldı ve her bir grupta 10 hayvan bulunduruldu. Kontrol grubuna ticari rasyon verilirken deneme gruplarından Grup 2, 3, 4, 5, 6, 7 ve 8'in yemlerine sırasıyla 2,5 g/kg yem HSCAS; 5,0 g/kg yem HSCAS; 10,0 g/kg yem HSCAS; 2,50 ppm aflatoksin (Af B₁ % 78,30, Af B₂ % 14,60, Af G₁ % 4,50, Af G₂ % 2,60); 2,50 ppm AF+2,50 g/kg yem HSCAS; 2,50 ppm AF+5,0 g/kg yem HSCAS; 2,50 ppm AF+10,0 g/kg yem HSCAS katıldı ve 21 gün süreyle verildi. Denemenin sonunda, hayvanlardan kan alındı ve kanda triiodotironin (T₃), tiroksin (T₄), tirotropin (TSH) ve testosteron düzeyleri ölçüldü. Sonuçta, kontrol grubuna göre istatistiksel olarak, T₃ düzeyinde Grup 3 ve 4'de önemli bir artış, Grup 5 ve 6'da önemli bir düşüş; T₄ düzeyinde Grup 2, 4, 5, 7 ve 8'de önemli bir artış; testosteron düzeyinde bütün deneme gruplarında (Grup 2-8) önemli bir düşüş tespit edildi.

Anahtar Sözcükler: Aflatoksin, HSCAS, hormon, bıldırcın

Introduction

Aflatoxins (AFs) are natural contaminants of feed and feedstuffs (1). Poultry are most sensitive to these toxins (2). Although ducks are claimed to be the most sensitive poultry animals (3), sensitivity tests carried out on quails revealed that these animals may be easily affected by AFs present in feed (4,5). The severity of poisoning by AFs depends on the age, sex and species of the animal, the amount being exposed to and duration of exposure. The vitamins, minerals and antibiotics present in feed are among the factors that change the severity of poisoning.

In addition, the amount of protein in the feed composition is closely related to poisoning (6,7). The liver is one of the organs most affected by AFs (8). They have various effects on other organs (9). The most suitable method to bind AFs in digestive tract is to add adsorbents to animals' feeds at certain rates, which hinder their absorption and alleviate their adverse effects (10,11). These compounds bind to AFs irreversibly in the digestive tract, and reduce the rate of AF absorbed and released into systemic circulation (12,13).

In this study our aim was to determine whether AF and hydrate sodium calcium aluminosilicate (HSCAS), which were given to quails alone and in combination at certain doses for 21 days, had any effect on thyroid hormones and testosterone. Previously, no study was carried out to find the effects that may occur in hormones when AF and HSCAS are given alone and in combination in quails. For this reason, the effects that occurred in animals due to subacute exposure to both compounds were evaluated and it was determined whether one or more parameters used in in-vitro efficacy tests were determinative criteria concerning the efficacy of the HSCAS in quails. This study will also be a guide for determining the risk of any poisoning based on these parameters long before the appearance of clinical symptoms in AF poisonings in quails.

Materials and Methods

Eighty male 14-day-old *Coturnix coturnix japonica* breed quails were used. The quails were divided into 8 groups, 1 control and 7 trial groups. While the animals in the control group were fed a commercial basal ration, 2.5 g/kg feed HSCAS, 5 g/kg feed HSCAS, 10 g/kg feed HSCAS, 2.5 ppm aflatoxin, 2.5 ppm AF with 2.5 g/kg feed HSCAS, 2.5 ppm AF with 5 g/kg feed HSCAS, 2.5 ppm with 10 g/kg feed HSCAS, respectively, were given to the animals in groups 2-8 for 21 days. The study was performed in separated quails, cages that were equipped with 24 h lighting, free access to water and feed, at 27-29 °C on a daily basis. On day 21, blood samples were taken from the animals and triiodothyronine (T_3), thyroxine (T_4), thyrotrophin (TSH) and testosterone levels in the blood were determined. The detection of plasma T_3 , T_4 , TSH and testosterone levels was performed in a Boehringer Mannheim Elecsys 2010 brand immunoassay analyzer using in vitro electrochemiluminescence (ECL). AF was added to the feed according to Demet et al.'s (14) method based on the method of Shotwell et al. (15). The species of AF in the rice was detected based on the method described by Roberts and Patterson (16), according to the method described reported by Şanlı et al. (17). It was determined that rice flour contained AF B₁, B₂, G₁, G₂ and G₂. Their rates were calculated according to Nabney and Nesbit's method (18). The rates for AF B₁, B₂, G₁ and G₂ were, respectively, 78.30%, 14.60%, 4.50% and 2.60%. Total AF level in rice flour was detected in an ELISA

apparatus using a Ridascreed® total AF kit and according to the method suggested in the kit's instructions. Accordingly, 84.68 ppm total AF was detected in rice flour with AF. The data were evaluated as arithmetic means and standard deviations; the significance of the groups was detected by one-way variance analysis. Duncan's test was used to determine differences between the groups (using SPSS 10.0 for Windows).

Results

Significant differences were detected in the parameters except for TSH levels in the groups that received adsorbent and AF alone and in combination for 21 days. Compared to the control group, significant increases were detected in T_3 levels in groups 3 and 4 and a significant decrease in groups 5 and 6, while there were significant increases in T_4 levels in groups 2, 4, 5, 7 and 8, and significant decreases in testosterone levels in all trial groups (groups 2 to 8) (Table).

Discussion

Thyroid hormones are among the major hormones playing important roles in the protection of the physiological balance of the body (19). Malfunctions in these hormones directly affect the general situation of the living being (20). Firstly, it was evaluated whether AF had any effect on thyroid hormones. There are numerous studies about animals that were given AF in feed and thus seriously affected (9-11,21). In this study, when an evaluation was performed in terms of thyroid hormones, a significant decrease was detected in T_3 levels and a significant increase in T_4 levels in the groups that received AF alone compared to the control group. No statistically significant difference was found in TSH levels. The results showed that T_3 and T_4 might have been greatly affected by AF. However, it is clear that this effect did not occur directly through TSH because insignificant changes were observed in the trial groups concerning this hormone level compared to the control. However, changes in T_3 and T_4 levels must cause a change, even though indirectly, in TSH levels since a decrease in blood T_3 level directly stimulates T_3 -sensitive nuclear receptors in the thyroid gland thus causing TSH synthesis and release (22,23). An increase in blood TSH level accelerates the absorption of iodide from the digestive tract and its diffusion into the thyroid gland (23,24). Iodide that is diffused into the

Table. T₃, T₄, TSH and testosterone levels in control and experimental groups.

Groups*	T ₃ (ng/ml)	T ₄ (µg/dL)	TSH (µIU/ml)	Testosterone (ng/dl)
Group 1	1.70 ± 0.23 ^c	0.65 ± 0.18 ^a	0.22 ± 0.06	109.01 ± 15.86 ^a
Group 2	1.69 ± 0.24 ^c	1.20 ± 0.21 ^b	0.27 ± 0.08	84.63 ± 11.54 ^b
Group 3	1.91 ± 0.24 ^d	0.81 ± 0.12 ^a	0.24 ± 0.04	74.50 ± 13.00 ^c
Group 4	1.93 ± 0.23 ^d	1.62 ± 0.33 ^d	0.22 ± 0.05	62.23 ± 7.81 ^c
Group 5	1.22 ± 0.18 ^a	1.41 ± 0.10 ^c	0.23 ± 0.07	29.70 ± 11.69 ^e
Group 6	1.44 ± 0.17 ^b	0.74 ± 0.10 ^a	0.20 ± 0.09	50.60 ± 15.33 ^d
Group 7	1.59 ± 0.15 ^{bc}	1.15 ± 0.13 ^b	0.21 ± 0.05	47.72 ± 6.87 ^d
Group 8	1.71 ± 0.19 ^c	1.04 ± 0.13 ^b	0.23 ± 0.10	37.55 ± 9.10 ^e

^{a, b, c, d, e} Means within the same column with different letters are statistically significant ($P < 0.05$).

* Group 1, control; Group 2, 2.5 g/kg feed HSCAS; Group 3, 5.0 g/kg feed HSCAS; Group 4, 10.0 g/kg feed HSCAS; Group 5, AF; Group 6, AF + 2.5 g/kg feed HSCAS; Group 7, AF + 5.0 g/kg feed HSCAS; Group 8, AF + 10.0 g/kg feed HSCAS.

thyroid gland forms a complex with a thyroglobulin molecule. Following that, every one or two thyroglobulin molecules unite and so T₃ and T₄ synthesis occurs. In this way, T₃ and T₄ levels in the blood are kept at a certain level (23-25). However, in our study, the lack of a significant increase in TSH levels might have been caused by the reduction in the sensitivity of the receptors in the thyroid gland due to AF. It is known that AFs cause lipid peroxidation in cells (26). Reactive oxygen species, which cause lipid peroxidation and whose formation is accelerated by AFs, may lead to conformational changes in receptors. Such changes may also hinder T₃'s binding to these receptors and the activation of the intracellular messenger system. Hence, a physiological response may not develop in the body concerning changes in blood T₃ levels. On the other hand, the reason for an increase in T₄ and a decrease in T₃, which were observed in the group that received AF alone compared to the control group, might have been a slowdown in the conversion of T₄ to T₃ in peripheral tissues. While some of the T₃ diffused into the bloodstream is synthesized in the thyroid gland, a major proportion of it occurs as a result of conversion of T₄, synthesized in the thyroid gland, into T₃ in peripheral tissues. 5'-deiodinase is the enzyme primarily responsible for this conversion. On the other hand, malic enzyme and 6-phosphogluconate dehydrogenase also take part in this process. These enzymes convert NAD into NADP, which is responsible for this conversion (23). It is possible that AF limits the conversion of T₄ into T₃ by causing the changes,

mainly in 5'-deiodinase as well as in other enzymes activities, that were mentioned above. As a result, while the T₃ level decreases, the T₄ level may increase. It is very likely that this mechanism has effects concerning the changes in the blood levels of these hormones. On the other hand, in the groups that received feed containing both AF and HSCAS significant changes were observed in T₃ and T₄ levels, compared to the groups that received AF alone. These changes were an increase in T₃ and a decrease in T₄. The decrease in T₃ and the increase in T₄ revealed that HSCAS was bound to AF in the digestive tract. The fact that neither of these parameters (T₃ and T₄) were close to the values of the control group and that there was a statistically significant difference between most of the groups indicated that the binding was not complete and a certain proportion of AF diffused into the blood and exerted its effect. Interesting results were obtained in the groups that received adsorbent alone. Some of them exhibited significant increases in both T₃ and T₄ levels. However, some researchers reported that sodium bentonite altered the levels of some minerals in the body. It was stressed that it caused an increase in the absorption of some and a decrease in the absorption of others (27). Similarly, the adsorbent causes an increase in the absorption and this may lead to an increase in the synthesis of both hormones.

Secondly, blood testosterone levels change as a result of malfunctions and extreme damage to Leydig cells, where testosterone is synthesized (28). It is known that

AF causes damage to the testes (29). When blood testosterone level was evaluated, a significant decrease was detected in the groups that received AF alone compared to the control group. This decrease also indicates an important malfunction in the testes. Of the groups that received AF with HSCAS, an increase was detected in testosterone levels, compared to the group that received AF alone. This increase also indicates that HSCAS was bound to AF. In fact, although an increase was observed in testosterone levels in the groups that received AF with HSCAS alone, compared to the group that received AF alone, this increase never reached the value of the control group. A significant decrease was detected in testosterone levels in the groups that received certain doses of HSCAS alone, compared to the control group. The decrease in testosterone level in the group received HSCAS only, although not certain, may be related directly or indirectly to the fact that the adsorbent bound to some compounds or caused an increase in the absorption of the others mentioned above.

In conclusion, AF given at a dose of 2.5 ppm for a subacute period affects blood T_3 , T_4 and testosterone levels negatively. It is quite likely that the main mechanism for AF to change T_3 and T_4 levels was not directly through the hypothalamus, hypophysis or thyroid

gland but decreases in the activities of enzymes such as mainly 5'-deiodinase, malic enzyme and 6-phosphogluconate dehydrogenase, which are responsible for the conversion of T_4 into T_3 , might have played an important role in the changes of T_3 and T_4 levels in peripheral tissue. As a result, the T_4 level increased and the T_3 level decreased. The decrease in testosterone level in the group that received AF alone, compared to the control group, showed that AF caused damage to Leydig cells of the testes. On the other hand, the statistically significant differences in hormone levels of the groups that received AF with adsorbent, compared to the group that received AF alone, revealed that HSCAS bound to AF in the digestive tract but this binding was not completed. It was concluded that the determinations of T_3 , T_4 and testosterone levels with other parameters (yield from quail feed performance, histopathological findings, biochemical parameters) will be important in in-vitro efficacy trials with HSCAS in quails against AF and an evaluation can be made based on the levels of these parameters (T_3 , T_4 and testosterone) in the blood, long before the appearance of poisoning symptoms that may result from this toxin. Nevertheless, the data obtained show that more detailed research is needed to determine the exact effects of these 2 compounds on these hormones in quails.

References

1. Gugnani, H.C.: Ecology and taxonomy of pathogenic aspergilli. *Front. Biosci.*, 2000; 8: 346-357.
2. Robens, J.F., Richard, J.L.: Aflatoxins in animal and human health. *Rev. Environ. Contam. Toxicol.*, 1992; 127: 69-94.
3. Ostrowski-Meissner, H.T.: Effect of contamination of diets with aflatoxins on growing ducks and chickens. *Trop. Anim. Health Prod.*, 1983; 15: 161-168.
4. Parlat, S.S., Ozcan, M., Oguz, H.: Biological suppression of aflatoxicosis in Japanese quail (*Coturnix coturnix japonica*) by dietary addition of yeast (*Saccharomyces cerevisiae*). *Res. Vet. Sci.*, 2001; 71: 207-211.
5. Parlat, S.S., Yildiz, A.O., Oguz, H.: Effect of clinoptilolite on performance of Japanese quail (*Coturnix coturnix japonica*) during experimental aflatoxicosis. *Br. Poult. Sci.*, 1999; 40: 495-500.
6. Dalvi, R.R.: An overview of aflatoxicosis of poultry: its characteristics, prevention and reduction. *Vet. Res. Commun.*, 1986; 10: 429-443.
7. Kaya, S.: Mikotoksinler. İçinden: Kaya, S., Pirincci, I., Bilgili, A. ed: Veteriner Hekimliğinde Toksikoloji. Medisan Yayın Serisi 53, Ankara, 544-573, 2002.
8. Pier, A.C., Richard, J.L., Cysewski, S.J.: Implications of mycotoxins in animal disease. *J. Am. Vet. Med. Assoc.*, 1980; 176: 719-724.
9. Ortatlatli, M., Oguz, H.: Ameliorative effects of dietary clinoptilolite on pathological changes in broiler chickens during aflatoxicosis. *Res. Vet. Sci.*, 2001; 71: 59-66.
10. Kececi, T., Oguz, H., Kurtoglu, V., Demet, O.: Effects of polyvinylpyrrolidone, synthetic zeolite and bentonite on serum biochemical and haematological characters of broiler chickens during aflatoxicosis. *Br. Poult. Sci.*, 1998; 39: 452-458.
11. Harvey, R.B., Kubena, L.F., Phillips, T.D.: Evaluation of aluminosilicate compounds to reduce aflatoxin residues and toxicity to poultry and livestock: a review report. *Sci. Total Environ.*, 1993; Suppl. Pt. 2: 1453-1457.

12. Marquez Marquez, R.N., Tejada de Hernandez, I.: Aflatoxin adsorbent capacity of two Mexican aluminosilicates in experimentally contaminated chick diets. *Food Addit. Contam.*, 1995; 12: 431-433.
13. Diaz, D.E., Hagler, W.M. Jr, Hopkins, B.A., Whitlow, LW.: Aflatoxin binders I: in vitro binding assay for aflatoxin B1 by several potential sequestering agents. *Mycopathologia*, 2002; 156: 223-226.
14. Demet, Ö., Oğuz, H., Çelik, İ., Nizamlioğlu, F.: Pirinçte aflatoksin üretilmesi. *Vet. Bil. Derg.*, 1995; 11: 19-23.
15. Shotwell, O.L., Hesseltine, C.W., Stubblefield, R.D., Sorenson, W.G.: Production of aflatoxin on rice. *Appl. Microbiol.*, 1966; 14: 425-428.
16. Roberts, B.A., Patterson, D.S.P.: Detection of twelve mycotoxins in mixed animal feedstuffs, using a novel membrane cleanup procedure. *J. Assoc. Off. Anal. Chem.*, 1975; 58: 1178-1181.
17. Şanlı, Y., Ceylan, S., Kaya, S.: Karma yemlerde aflatoksin analizi. *Ankara Üniv. Vet. Fak. Derg.*, 1982; 29: 50-70.
18. Nabney, J., Nesbit, B.F.: A spectrophotometric method for determination of the aflatoxins. *Analyst*, 1965; 3: 155-159.
19. Nakamura, H., Nakao, K.: Mechanism of regulation of TSH-biosynthesis and secretion. *Nippon Rinsho.*, 1993; 51: 2611-2617.
20. Rose, S.R.: Disorders of thyrotropin synthesis, secretion, and function. *Curr. Opin. Pediatr.*, 2000; 12: 375-381.
21. Oguz, H., Kurtoglu, V.: Effect of clinoptilolite on performance of broiler chickens during experimental aflatoxicosis. *Br. Poult. Sci.*, 2000; 41: 512-517.
22. Wiersinga, W.M.: Nuclear thyroid hormone receptors. *Neth. J. Med.*, 1985; 28: 74-82.
23. Noyan, A.: Yaşamda ve Hekimlikte Fizyoloji. *Hormonlar*, 977-1033, 1993, Ankara.
24. Markou, K., Georgopoulos, N., Kyriazopoulou, V., Vagenakis, A.G.: Iodine-induced hypothyroidism. *Thyroid*, 2001; 11: 501-510.
25. Dunn, J.T., Dunn, A.D.: The importance of thyroglobulin structure for thyroid hormone biosynthesis. *Biochimie*, 1999; 81: 505-509.
26. Rastogi, R., Srivastava, A.K., Rastogi, A.K.: Long term effect of aflatoxin B1 on lipid peroxidation in rat liver and kidney: effect of picroliv and silymarin. *Phytother. Res.*, 2001; 15: 307-310.
27. Grosicki, A., Kowalski, B.: Influence of bentonite on trace element kinetics in rats. *Bull. Vet. Inst. Pulawy*, 2003; 47: 555-558.
28. Aydinler, A., Aytakin, Y., Topuz, E.: Effects of cisplatin on testicular tissue and the Leydig cell-pituitary axis. *Oncology*, 1997; 54: 74-78.
29. Ortatatlı, M., Ciftci, M.K., Tuzcu, M., Kaya, A.: The effects of aflatoxin on the reproductive system of roosters. *Res. Vet. Sci.*, 2002; 72: 29-36.