

Turkish Journal of Veterinary and Animal Sciences

http://journals.tubitak.gov.tr/veterinary/

Research Article

Turk J Vet Anim Sci (2015) 39: 23-28 © TÜBİTAK doi:10.3906/vet-1409-31

Melatonin receptor 1A gene RsaI and inhibin alpha subunit gene HaeII polymorphisms in Honamli and Hair goat breeds reared in Western Mediterranean region of Turkey

Özgecan KORKMAZ AĞAOĞLU^{1,*}, Mustafa SAATCI¹, Bilal AKYÜZ², Özkan ELMAZ¹, Mehmet ÇOLAK¹, Burcu Menekşe BALKAN³, Emel ZEYTÜNLÜ⁴

¹Department of Animal Science, Faculty of Veterinary Medicine, Mehmet Akif Ersoy University, Burdur, Turkey ²Department of Genetics, Faculty of Veterinary Medicine, Erciyes University, Kayseri, Turkey ³Department of Biochemistry, Faculty of Veterinary Medicine, Mehmet Akif Ersoy University, Burdur, Turkey ⁴Department of Animal Science, Institute of Health Sciences, Mehmet Akif Ersoy University, Burdur, Turkey

Received: 11.09.2014	٠	Accepted: 16.12.2014	٠	Published Online: 12.01.2015	٠	Printed: 09.02.2015
----------------------	---	----------------------	---	------------------------------	---	---------------------

Abstract: The melatonin receptor 1A (MTNR1A) and inhibin alpha subunit (INHA) genes play a significant role in the reproductive characteristics of animals. Blood samples were collected from 371 goats (Honamli and Hair) reared in Antalya and Burdur. The polymerase chain reaction (PCR) products were digested by RsaI for the MTNR1A gene and HaeII for the INHA gene. Two alleles (A and G) and three genotypes (AA, AG, and GG) were observed for the INHA gene, while two alleles (R and r) and two genotypes (RR and Rr) were observed for the MTNR1A gene. The highest allelic frequency value for G (91.8%) was found in Honamli goat breeds for the INHA gene while the highest value for R (98.1%) was found in Hair goat breeds for the MTNR1A gene. The GG genotype for the INHA gene and the RR genotype for the MTNR1A gene were identified as the most common genotypes of the Honamli and Hair goat breeds. The rr genotype for the MTNR1A gene could not be determined in the breeds. Both Honamli and Hair goat breeds were in Hardy-Weinberg equilibrium for the genes that were studied. In conclusion, this study confirms the existence of genetic polymorphism in the MTNR1A and INHA genes as detected by PCR-RFLP analysis in Honamli and Hair goat breeds.

Key words: Goat, Honamli, Hair, INHA, MTNR1A

1. Introduction

There are many potential genes known to be related to the economic traits in farm animals and that can be used for selection criteria. Economic traits are quantitative characters controlled by several genes and also are strongly affected by environmental conditions. Since molecular genetic technologies have become more powerfully applicable in industry in recent years, it has become possible to identify which genes have an effect on variations that can be observed in these quantitative traits. This helps to speed up and improve the effectiveness of desired selections. In this regard, there has been a significant increase in the number of studies on the polymorphism of genes that affect the economic traits of livestock species (1-4). Reproductive traits are the most important economic characters in farm animal breeding. There are a number of genes that affect reproduction and that can be employed in selection programs. Two of these genes are inhibin alpha subunit (INHA) and melatonin receptor 1A (MTNR1A), which play a significant role in the reproductive process in animals (1,2). Inhibins are dimeric glycoproteins that

* Correspondence: ozgecanagaoglu@mehmetakif.edu.tr

are made up of a common inhibin alpha subunit (INHA) that is covalently linked to one of two related subunits, inhibin beta A or inhibin beta B (INHbA and INHbB) (5). Inhibin subunits are encoded by INHA, INHbA, and INHbB. Inhibin A inhibits FSH secretion by suppressing its receptor expression in granulosa cells, thus affecting the recruitment and development of ovarian follicles during folliculogenesis (6,7). The Ala257Thr missense mutation of the INHA gene has been shown to play an important role in receptor binding. Furthermore, the INHA gene has been suggested as a very likely cause of premature ovarian failure (8). Hou et al. (3) identified the polymorphisms in the 5' promoter region of the INHA gene and concluded that these polymorphisms could be potential genetic markers for determining the litter size of goats. In Boer goats, INHA 651A/G polymorphism can have a significant effect on the mean litter size of parity-two animals (2). Goat breeds with both seasonal and year-round estrus also have different genotype distributions of the INHA gene, which points to a relationship between the INHA gene and fecundity (8). Tang et al. (9) found MspI polymorphism in the bovine INHA gene as well as a correlation with the features of superovulation. Considerable influence on reproductive traits makes the INHA gene a prominent candidate for consideration (10). Melatonin is secreted from the pineal gland and has two receptors, which are classified as subtypes MTNR1A and MTNR1B. MTNR1A influences the regulation of seasonal reproductive activity (11). Chu et al. (12) reported that a polymorphic site in position 53 (GenBank AF419334) in Asian goat breeds has a correlation with seasonal reproduction. It has been found that Sarda, Saanen, Chamois Coloured, Maltese, and Nubian goat breeds have MTNR1A gene polymorphism (13). Furthermore, Mateescu et al. (1) identified a link between the MTNR1A gene and lambing frequency. These findings indicate that the MTNR1A gene is potentially an important DNA marker for breeding.

Hair goats are most frequently raised in the Mediterranean and Aegean regions and at higher altitudes in villages and small towns in and near the forested regions of Central Anatolia (14). On the other hand, Honamli goats are usually bred in the provinces of Antalya, Burdur, and Konya, which are located near the foothills of the Taurus Mountains in the western part of the Mediterranean region (14,15). The Honamli goat is a combined productive goat raised for meat, milk, and hair (14). Although there are limited reproductive studies about Honamli goats, the findings of some studies (15,16) revealed that the Honamli breed was superior to other local goat breeds in terms of various reproductive characteristics. Similarly, the number of reproductive studies on Hair goats is considerably limited (17-19). Determination of desirable genotypes of genes that have effects on reproduction traits is critically important in animal breeding programs. Therefore, determination of the MTNR1A and INHA gene polymorphisms, previously reported to have effects on reproductive characteristics, in these breeds will contribute to the literature.

The goal of this study was to investigate polymorphisms of *INHA* and *MTNR1A* genes, which have been reported to affect reproduction, using the polymerase chain reactionrestriction fragment length polymorphism (PCR-RFLP) method.

2. Materials and methods

2.1. Samples

Blood samples were randomly collected from 371 goats belonging to the Honamli (n = 183) and Hair (n = 188) goat breeds reared in Antalya and Burdur. Blood samples were collected in tubes with K_{q} -EDTA.

2.2. DNA isolation and genotyping

DNA was isolated using a DNA isolation kit (GeneJET Genomic DNA Purification Kit). Quantity and quality of DNA samples were examined using a NanoDrop 2000 (Thermo Scientific). DNA amplification of the *MTNR1A* and *INHA* genes was carried out by PCR and all PCR reactions were performed on an Amplitronyx Series 6 thermal cycler. All procedures were carried out at the Molecular Genetics Research Laboratories of the Mehmet Akif Ersoy University Faculty of Veterinary Medicine, Department of Animal Science.

The PCR amplification reaction was carried out in a total volume of 25 µL consisting of MgCl²⁺ (2.5 mM for INHA and 1.5 mM for MTNR1A), dNTP (200 µM), primers (5 pmol) (Table 1), 1X buffer, Taq DNA polymerase (1 U/ µL), and DNA (~100 ng). The PCR conditions including an initial denaturing step of 94 °C for 5 min, followed by 35 cycles of 94 °C for 45 s for INHA and 60 s for MTNR1A, 58.4 °C for 30 s for INHA and 60 s for MTNR1A, and 72 °C for 45 s, with a final cycle at 72 °C for 10 min. The PCR products of the INHA and MTNR1A genes were digested with HaeII (Thermo Scientific, #FD2184) and RsaI (Thermo Scientific, #FD1124) restriction endonuclease enzymes according to instructions from the manufacturer (Fermentas), respectively. Digested PCR products were electrophoresed on 3% (MTNR1A) or 4% (INHA) agarose gels and then visualized under a UV-transilluminator.

2.3. Data analysis

Allele and genotype frequencies, observed and expected heterozygosity values, and Hardy–Weinberg equilibrium were calculated using the PopGene32 (www.ualberta. ca/~fyeh/Pop32.exe) program.

3. Results

The amplified PCR product of the *INHA* and *MTNR1A* genes produced 217-bp and 824-bp fragments, respectively.

Gene		Primer	PCR product size	RE	Reference
INHA	Forward Reverse	5'-CCACACAGGACTGGACAGACA-3' 5'-GCAGGAACAGAGAGGACAACG-3'	217 bp	HaeII	(20)
MTNR1A	Forward Reverse	5'-TGTGTTTGTGGTGAGCCTGG-3' 5'-ATGGAGAGGGTTTGCGTTTA-3'	824 bp	RsaI	(21)

Table 1. Primer sequence and restriction endonucleases.

*Hae*II digestion of the *INHA* gene resulted in fragments of 27 (not appear on gel photo) and 190 bp for the GG, 27, 190 and 217 bp for the AG, and 217 bp for the AA genotype (Figure 1).

Restricted PCR products for the *MTNR1A* gene are given in Figure 2. Digestion with *RsaI* enzyme produced five fragments (23, 53, 70, 267, and 411 bp); however, a site in position 53 was polymorphic. The presence of this cleavage site produces two fragments of 53 and 267 bp (R allele), while the absence of this site produces only one fragment of 320 bp (r allele). Restriction digestion of PCR products with *RsaI* enzymes revealed two genotypes (Figure 2) of RR (267 bp) and Rr (320 and 267 bp), but no rr (320 bp/320 bp) genotype was detected.

Two alleles (A and G) and three genotypes (AA, AG and GG) were observed for the INHA gene, while two alleles (R and r) and two genotypes (RR and Rr) were observed for the MTNR1A gene. The results of statistical analysis are presented in Tables 2 and 3. The highest allelic frequency value for G (91.8%) was found in Honamli goats for the INHA gene while the highest value for R (98.1%) was found in Hair goats for the MTNR1A gene. The GG genotype for the INHA gene and the RR genotype for the MTNR1A gene were identified as the most common genotypes of the Honamli and Hair goat breeds. Based on genotype frequency values, the MTNR1A rr genotype was not determined in these breeds. The observed heterozygosity value was 0.04 and 0.10 for the MTNR1A gene in Hair and Honamli goat breeds, while the observed heterozygosity value was 0.21 and 0.13 for the INHA gene in Hair and Honamli goat breeds, respectively. The expected heterozygosity for the MTNR1A and INHA genes of Hair and Honamli goats is shown in Tables 2 and 3. According to the Hardy-Weinberg equilibrium test, both Honamli and Hair goat breeds were in equilibrium for these genes.

4. Discussion

Genetic improvement in reproductive traits associated with seasonal reproduction in livestock such as goats,

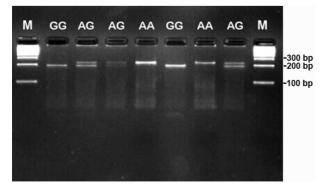


Figure 1. Gel image for the *INHA* genotypes by PCR-RFLP analysis. Lane M, molecular marker (100-bp DNA ladder).

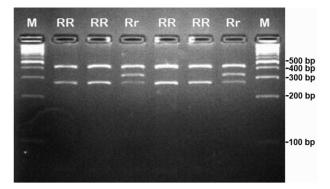


Figure 2. Gel image for the *MTNR1A* genotypes by PCR-RFLP analysis. Lane M, molecular marker (100-bp DNA ladder).

sheep, and buffalo is difficult for the following reasons. First, these traits have low heritabilities (22); second, they are generally not expressed until puberty; third, they are usually important only in females; and fourth, they are only monitored in different birth seasons and in different locations depending on changes in the length of the day (23). Therefore, especially for goat and sheep breeders, seasonal reproduction is the primary factor that limits economic production. For this reason, work has been focused on improvements out of breeding season in goats and sheep. However, due to the aforementioned reasons, success has been limited with conventional improvement programs in these species. The effect of seasonality on reproduction in small ruminants can be limited by implementing marker-assisted selection programs using genetic markers. Genes are selected to either increase the ovulation rate or eliminate the limiting effect of seasonality. The information that is currently available indicates that the MTNR1A gene can be used to carry out more efficient selection for animal reproduction in the nonbreeding season. In addition, the INHA gene may be used in superovulation studies in small ruminants. In the present study, the genetic polymorphisms of the MTNR1A and INHA genes in two Turkish local goat breeds (Honamli and Hair) were examined using the PCR-RFLP method.

In mammals, the secretion of melatonin is triggered by the elongation and shortening of the day. In turn, seasonal reproductive activity for livestock such as goats, sheep, and buffalo is significantly influenced by melatonin (24). On the other hand, MTNR1A is thought to be the main receptor involved in the regulation of seasonal reproductive activities in mammals (11). Moreover, *MTNR1A* gene polymorphism has been found to be significantly related to seasonal reproduction in sheep (25), goats (13,26), and buffalo (27). Therefore, *MTNR1A* gene polymorphism can be used to regulate seasonal and nonseasonal reproductive activities in goats.

In this study, the 824-bp PCR products of exon II of the *MTNR1A* gene were digested with the restriction

Breed		Allele f. (%)		Genotype f. (%)			Heterozygosity		-2(df - 1)
	n	A	G	AA	AG	GG	Но	He	$-\chi^2 (df = 1)$
Hair	188	12.77	87.23	2.12	21.28	76.60	0.21	0.22	0.43 ^{ns}
Honamli	183	8.20	91.80	1.64	13.11	85.25	0.13	0.15	3.24 ^{ns}

Table 2. Allele and genotype frequencies of INHA gene for HaeII site in Hair and Honamli goat breeds.

f.: frequency, ns: nonsignificant.

Table 3. Allele and genotype frequencies of MTNR1A gene for RsaI site in Hair and Honamli goat breeds.

Breed r		Allele f. (%)		Genotyj	Genotype f. (%)			zygosity	.2 (16 1)
	n	R	r	RR	Rr	rr	Но	He	$- \chi^2 (\mathrm{df} = 1)$
Hair	188	98.1	1.9	96.28	3.72	0.00	0.04	0.04	0.06 ^{ns}
Honamli	183	94.8	5.2	89.62	10.38	0.00	0.10	0.09	0.52 ^{ns}

f.: frequency, ns: nonsignificant.

endonuclease RsaI in Honamli and Hair goat breeds. The results indicated that the 267-bp and 320-bp fragments were polymorphic. However, the RR genotype was found to have a higher frequency than the Rr genotype in Hair (3.72%) and Honamli (10.38%) goat breeds. No rr homozygotes were detected in the two Turkish local goat breeds. Similar to our results, the homozygote rr genotype was not found in Chinese local goat breeds (Jining Grey, Liaoning Cashmere, Inner Mongolia Cashmere, Wendeng milk, and Beijing native goats) and Boer goats in China (26). Similarly, this genotype was not found in original European goat breeds such as Sarda, Saanen, Chamois Coloured, and Maltese goats, as well as Nubian goats that originated from Africa. In addition, the Rr genotype was only found in the Sarda breed, while the other five goat breeds (Saanen, Chamois Coloured, Maltese, Nubian) were found to be monomorphic (only the RR genotype) in terms of the MTNR1A gene (13).

Polymorphism at the *Rsa*I site of the *MTNR1A* gene was associated with year-round estrus and seasonal anovulatory activity in Small Tailed Han sheep (26). Similarly, the *Rsa*I site of the *MTNR1A* gene was found to be polymorphic and RR genotype frequency (0.13) was found to be lower than Rr (0.43) and rr (0.44) genotypes in Dorset sheep, which represents year-round estrus (1).

However, an association was found between the RR genotype and year-round estrus in Jining Grey and Boer goats, and an association between the Rr genotype and seasonal estrus was reported in goats (Liaoning Cashmere, Inner Mongolia Cashmere, Wendeng milk. and Beijing native goats) in China (12).

The RsaI-Rr genotype was found in some breeds that have seasonal reproduction while only the RR genotype was found in Jining Grey goats, which is not a seasonal breed (26). It was reported that the Rr genotype, even if it was found in only a few breeds, showed a strong link with reproductive activity in goats. On the other hand, it has been suggested that the absence of RsaI polymorphism in some breeds may be associated with different origins of breed groups, because in European breeds (Saanen and Chamois Coloured) there is no polymorphism, while in Asian and African groups, some breeds exhibit polymorphism (13). Similarly, the RsaI site of the MTNR1A gene was found to be polymorphic in the breeds that were examined (Hair and Honamli) in this study, which are raised in Western Mediterranean region of Turkey. Neither Hair nor Honamli goats have planned selection programs conducted to increase production and achieve better control over reproductive activity. In these breeds, sexual activity has been always influenced by the photoperiod, which ensures seasonal lambing based on climatic conditions. Thus, it is reasonable to think that in Hair and Honamli breeds, low selective pressure has led to the existence of the r allele. In these goat breeds, lambing in a favorable climatic period is absolutely necessary to guarantee the survival of the offspring.

Another way to increase efficiency in small ruminants is to increase fecundity. Several fecundity genes have been described in some sheep breeds, including *INHA* (28,29). However, studies on these genes in goats are limited. Nevertheless, the *INHA* gene has been reported to have a positive correlation with litter size in Boer goats (2). In this study, all three genotypes were detected in the two native Turkish breeds. It was determined that there is still sufficient genetic diversity in Hair and Honamli goat breeds. However, more studies are needed to investigate the relationship between the *INHA* gene and litter size in goat breeds.

In conclusion, results obtained for the two Turkish native goat breeds examined in this study show the existence of genetic polymorphism in the *MTNR1A* and *INHA* genes. Future studies are required to evaluate the relationship between different *MTNR1A* and *INHA* genotypes and reproductive seasonality and offspring in goats. Considering the cultural, historical, and environmental importance of goat production in Turkey, the data obtained here could be used as an initial guide for developing rational breeding strategies for increasing goat production as well as for preserving and utilizing local goat breeds in the region. The relationships between the *Rsa*I-RR genotype and polyestrus as well as the *Rsa*I-

References

- Mateescu RG, Lunsford AK, Thonney ML. Association between melatonin receptor 1A gene polymorphism and reproductive performance in Dorset ewes. J Anim Sci 2009; 87: 2485–2488.
- Wu WS, Hua GH, Yang LG, Wen QY, Zhang CY, Zoheir KM, Chen SL. Association analysis of the *INHA* gene with litter size in Boer goats. Small Ruminant Res 2009; 82: 139–143.
- Hou J, An X, Li G, Wang Y, Song Y, Cao B. Exploring polymorphisms and their effects on reproductive traits of the *INHA* and *INHβA* genes in three goat breeds. Anim Sci J 2012; 83: 273–278.
- 4. Akçay A, Akyüz B, Bayram D. Determination of the *Alu*I polymorphism effect of bovine growth hormone gene on carcass traits in Zavot cattle with analysis of covariance. Turk J Vet Anim Sci 205; 39 16-22
- Ling N, Ying SY, Ueno N, Esch F, Denoroy L, Guillemin R. Isolation and partial characterization of a Mr 32,000 protein with inhibin activity from porcine follicular fluid. P Natl Acad Sci USA 1985; 82: 7217–7221.
- Groome NP, Illingworth PJ, O'Brien M, Cooke I, Ganesan TS, Baird DT, McNeilly AS. Detection of dimeric inhibin throughout the human menstrual cycle by two-site enzyme immunoassay. Clin Endocrinol (Oxf) 1994; 40: 717–723.
- Groome NP, Illingworth PJ, O'Brien M, Pai R, Rodger FE, Mather JP, McNeilly AS. Measurement of dimeric inhibin B throughout the human menstrual cycle. J Clin Endocrinol Metab 1996; 81: 1401–1405.
- He Y, Ma X, Liu X, Zhang C, Li J. Candidate genes polymorphism and its association to prolificacy in Chinese goats. J Agric Sci 2010; 2: 88–92.
- Tang KQ, Li SJ, Yang WC, Yu JN, Han L, Li X, Yang LG. An *MspI* polymorphism in the inhibin alpha gene and its associations with superovulation traits in Chinese Holstein cows. Mol Biol Rep 2011; 38: 17–21.

Rr genotype and seasonal estrus were reported (12,24). In this study, higher RR genotypic frequencies were found in Honamli and Hair goats, although these animals are known as seasonal polyestrous breeds. Therefore, studies should be planned for investigation of correlation between genotypes of *MTNR1A-RsaI* and animals showing estrus out of season, and between polymorphism of *INHA-HaeII* and multiple pregnancies. The data obtained from these studies may have potential for studies to increase fertility traits of Honamli and Hair goats.

Acknowledgments

The authors acknowledge the support of the General Directorate of Agricultural Research and Policies (GDAR-TAGEM) of the Turkish Ministry of Food, Agriculture, and Livestock. This research was partly supported by the "Genetic Improvement of Honamli and Hair Goat in Breeders' Condition" projects. The authors would like to thank the staff of these projects.

- 10. Hiendleder S, Lewalski H, Jaeger C, Pracht P, Erhardt G. Genomic cloning and comparative sequence analysis of different alleles of the ovine β A-inhibin/activin (INHBA) genes as a potential QTL for litter size. Anim Genet 1996; 27: 91–92.
- Dubocovich ML, Rivera-Bermudez MA, Gerdin MJ, Masan MI. Molecular pharmacology, regulation and function of mammalian melatonin receptors. Front Biosci 2003; 8: 1093– 1108.
- Chu MX, He YY, Cheng DX, Ye SC, Fang L, Wang YY. Association between expression of reproductive seasonality and alleles of melatonin receptor 1A in goats. Anim Reprod Sci 2007; 101: 276–284.
- Carcangiu V, Vacca GM, Mura MC, Dettori M, Pazzola M, Luridiana S, Bini PP. Relationship between MTNR1A melatonin receptor gene polymorphism and seasonal reproduction in different goat breeds. Anim Reprod Sci 2009; 110: 71–78.
- 14. TAGEM. The Presentation Catalogue of Genetic Resources of Turkey's Domestic Animals. Ankara, Turkey: TAGEM; 2009 (in Turkish).
- Elmaz Ö, Saatci M, Dağ B, Aktaş AH, Ata A, Gülay MŞ, Mamak N, Gök B. Some descriptive characteristics of a new goat breed called Honamli in Turkey. Trop Anim Health Prod 2012; 44: 1913–1920.
- Gök B, Aktaş AH, Dursun Ş. Honamli goat: Rising star of the Taurus Mountains. In: RBI 8th Global Conference on the Conservation of Animal Genetic Resources. Tekirdağ, Turkey; 2011. pp. 65–72.
- Toplu HDO, Altinel A. Some production traits of indigenous Hair goats bred under extensive conditions in Turkey. 1st communication: Reproduction, milk yield and hair production traits of does. Arch Tierz Dummerstorf 2008; 51: 498–506.

- Atay O, Gökdal Ö, Eren V. Reproductive characteristics and kid marketing weights of hair goat flocks in rural conditions in Turkey. Cuban J Agri Sci 2010; 44: 353–358.
- Erten Ö, Yılmaz O. Investigation of reproductive and milk yield traits of Hair Goats raised under extensive conditions. YYÜ Vet Fak Derg 2013; 24: 105–107 (in Turkish with an English abstract).
- Hua GH, Chen SL, Yao HW, Wu WS, Shen Z, Chen QK, Chen L, Wen QY, Yang LG. *Hae*II RFLP of INHA and its relationship to goat litter size. Hereditas 2007; 29: 972–976.
- Reppert SM, Weaver DR, Ebisawa T. Cloning and characterization of mammalian melatonin receptor that mediates reproductive and circadian responses. Neuron 1994; 13: 1177–1185.
- 22. Yılmaz O, Küçük M, Denk H, Bolacalı M. The effect of adding ram at out of season on fertility of Norduz ewes and survival rate of lambs. YYÜ Vet Fak Derg 2006; 17: 99-102 (in Turkish with an English abstract).
- 23. Akçapınar H. Koyun yetiştiriciliği. 2nd ed. Ankara, Turkey: İsmat Matbaacılık; 2000.
- Lai P, Zhang BY, Wang PQ, Chu MX, Song WJ, Ca BJ. Polymorphism of the melatonin receptor genes and its relationship with seasonal reproduction in the Gulin Ma goat breed. Reprod Dom Anim 2013; 48: 732–737.

- 25. Carcangiu V, Mura MC, Vacca GM, Pazzola M, Dettori ML, Luridiana S, Bini PP. Polymorphism of the melatonin receptor MT1 gene and its relationship with seasonal reproductive activity in the Sarda sheep breed. Anim Reprod Sci 2009; 116: 65–72.
- Chu MX, Cheng DX, Liu WZ, Fang L, Ye SC. Association between melatonin receptor 1A gene and expression of reproductive seasonality in sheep. Asian-Aust J Anim Sci 2006; 19: 1079–1084.
- 27. Carcangiu V, Mura MC, Pazzola M, Vacca GM, Paludo M, Marchi B, Daga C, Bua S, Luridiana S. Characterization of the Mediterranean Italian buffaloes melatonin receptor 1A (MTNR1A) gene and its association with reproductive seasonality. Theriogenology 2011; 76: 419–426.
- Montgomery GW, Galloway SM, Davis GH, McNatty KP. Genes controlling ovulation rate in sheep. Reproduction 2001; 121: 843–852.
- George HD. Major genes affecting ovulation rate in sheep. Genet Sel Evol 2005; 37: 11–23.