

## Assessment of the molluscicidal activity of wormwood (*Artemisia dubia*, Wallich) leaves ethanolic extract on *Oncomelania hupensis quadrasi*, Möllendorff

Claudine C. TERCİÑO , Carl Leonard M. PRADERA , Melvin A. BAGOT\* 

College of Veterinary Medicine, Visayas State University, Visca, Baybay City, Leyte, Philippines

Received: 03.07.2019 • Accepted/Published Online: 20.01.2020 • Final Version: 06.04.2020

**Abstract:** This study assessed the in vitro molluscicidal activity of *Artemisia dubia* (wormwood) leaf ethanolic extract against adult and juvenile *Oncomelania hupensis quadrasi* and profiled its qualitative phytochemical content. The plants and snails were collected locally. There were 8 concentrations used for adult and juvenile snails: distilled water and 1% ethanol as negative controls; niclosamide (2 mg/L) as a positive control; and different concentrations of wormwood ethanolic extracts for adult snails including 3.98% (T1), 4.46% (T2), 5% (T3), 5.61% (T4), and 6.30% (T5) and for juvenile snails including 7.94% (T1), 8.91% (T2), 10% (T3), 11.22% (T4), and 12.59% (T5). Each treatment was replicated 5 times with 20 snails per replicate. Results showed that the concentrations that were statistically comparable with niclosamide (2 mg/L) were 6.30% (T5) with 92% mortality for adults and 10% (T3), 11.22% (T4), and 12.59% (T5) for juveniles with 94%, 95%, and 98% mortality, respectively ( $P > 0.05$ ). The qualitative phytochemical tests for secondary metabolites revealed the presence of tannins, saponins, and terpenoids. Independently or in combination, these secondary metabolites may be responsible for the mortality of the snails. This study indicates the possibility of using wormwood ethanolic extract as a potent and a possible alternative for synthetic molluscicides.

**Key words:** In vitro, mortality, schistosomiasis, snails

### 1. Introduction

Eliminating *Oncomelania hupensis quadrasi* (family Pomatiopsidae; Möllendorff, 1895) is important in schistosomiasis control. The control of these mollusks through the use of molluscicides will eliminate them and interrupt the life cycle of *Schistosoma japonicum* (family Schistosomatidae; Katsurada, 1904), and it keeps the prevalence of schistosomiasis in both animals and humans at a low level [1]. In developing countries where schistosomiasis is endemic, snail control is difficult owing to the high cost of synthetic compounds. This has resulted in frequent and increased studies on plants as alternative source of molluscicides [2,3].

*Artemisia dubia* var. *orientalis*, known as wormwood (family Compositae; Pamp., 1930), is considered one of the herbal plants that have potential molluscicidal activity. Species of plants of the genus *Artemisia* have been used for centuries as moth repellants, general pesticides, and as tea or spray to repel slugs and snails [4,5]. Thus, this study investigated the use of wormwood leaf ethanolic extract as molluscicides, specifically on *O. hupensis quadrasi*, and determined its potential as a molluscicide. It may be able to control the *O. hupensis quadrasi* populations where schistosomiasis is prevalent.

\* Correspondence: melvin.bagot@vsu.edu.ph

### 2. Material and methods

#### 2.1 Collection of wormwood leaves

The plants were collected from Barangay Gabas, Baybay City, Leyte, Philippines during the growth stage between in April and May and were identified by a botanist, Dr. Beatriz S. Belonias of the Department of Biological Sciences, College of Arts and Sciences, Visayas State University. The identification of plants was based on morphological keys as formerly described by previous studies [6,7]. The dirt and debris from the leaves were washed with running tap water. After washing, the leaves were air-dried at room temperature for 3 days [8]. Dried leaves were chopped finely into manageable pieces, ground finely, and air-dried again at room temperature for 7 days to remove the remaining moisture content of the leaves [9].

#### 2.2. Extraction of bioactive compounds

Ground dried leaves were completely submerged with ethanol for infusion for 48 h. After infusion, the preparation was filtered using a funnel topped with filter paper. The filtrate was placed in a beaker and the plant leaf residue was discarded. The filtrate was concentrated in a rotary evaporator at 40 °C and recovered 1/3 of its original volume. The recovered concentrated extract of wormwood

leaves ethanolic extract (wormwood EE) was stored in a properly labeled and sealed amber bottle. It was kept in the refrigerator until needed, but not more than 7 days to preclude possible contamination.

### 2.3. Phytochemical analysis

Qualitative phytochemical analysis of the wormwood EE was conducted by determining the following secondary metabolites: for alkaloids (Mayer's test and Wagner's test), a 2% HCl was mixed in 5 mL of extract and placed in a steam bath for warming; 2 different preparations were made and were added with 2 drops of Mayer's and Wagner's solutions, respectively. For tannins (tannins test), 2 drops of 5% FeCl<sub>3</sub> were added to 2 mL of extract; for saponins (Froth test), the extract in a test tube was simply shaken; for flavonoid tests, both alkaline reagent and lead acetate tests were performed, wherein 2–3 drops of 20% sodium hydroxide solution and 10% acetate solution were added to 2 mL of the prepared extracts; and for terpenoids (chloroform and sulfuric acid test), 2 mL of the extract was added and shaken into 2 mL of chloroform and 5 mL of concentrated sulfuric acid was added carefully. A qualitative grading system of the phytochemicals was applied as follows: strongly positive (+++), moderately positive (++), weakly positive (+), traces (-), and undetected (0) [10].

### 2.4. Collection of *Oncomelania hupensis quadrasi*

Adult and juvenile *O. hupensis quadrasi* snails were collected from the municipality of Palo, Leyte, Philippines. The collected snails were identified by a zoologist, Emelda R. Legaspi of the Center for Health Development, Department of Health - Eastern Visayas, using a previously described identification key [11]. Protective gears were used during the collection to avoid contamination and possible infection with *Schistosoma japonicum*. The snails were placed in a plastic container layered with moistened filter paper during transport and were transferred to an aquarium with an aerator in the laboratory until used for assays. Prior to assays, snails were sorted into juvenile and adult groups. Juvenile snails measure up to a maximum of 3 mm and snails that reached more than 3 mm were considered adult [12].

### 2.5. Molluscicidal assay

The different treatments and corresponding concentrations of wormwood EE used were as follows: T0<sub>(-1)</sub> - distilled water and T0<sub>(-2)</sub> - 1% ethanol as negative controls; T0<sub>(+)</sub> - 0.0002% of niclosamide as a positive control; for adult snails T1 (3.98%), T2 (4.46%), T3 (5%), T4 (5.61%), and T5 (6.30%); and for juvenile snails T1 (7.94%), T2 (8.91%), T3 (10%), T4 (11.22%), and T5 (12.59%). All treatments were replicated 5 times with 20 snails per replicate. The different concentrations were computed based on preliminary trials conducted, wherein the LC<sub>50</sub> values of wormwood EE for both adult and juvenile *O. hupensis quadrasi* were determined. The LC<sub>50</sub> values for adult and

juvenile snails were 4.66 and 4.69, respectively. The LC<sub>50</sub> was based on a probit analysis at 95% confidence level. After the establishment of the LC<sub>50</sub>, 2 higher and lower concentrations were computed through logarithmic methods.

The adult and juvenile snails were sorted in different petri dishes and tested separately. For the assay, a flooding technique [9] was carried out with viable snails to determine the molluscicidal efficacy of wormwood EE. The snails were observed and viewed under a 10× scanning objective; they were considered viable when their head-foot part protruded from the shell and their tentacles were extended as if attempting to crawl. Each snail was placed in a petri dish with individual circles (5 mm diameter) drawn on the plate and 10 mL of different concentrations of the extract was added. The assay was conducted for 24 h, after which the snails were transferred to another petri dish with distilled water and left for another 24 h to observe further whether they were dead or alive.

For the motility test, snails that moved out from the circles were considered alive, while those that remained immobile and retained their positions after 3 h were further tested for sensitivity.

For the sensitivity test, snails were observed under a microscope (10×) and it was checked whether the snails were sensitive to mechanical stimulation through needle prodding. Live snails usually retract their operculum when prodded. Immobile snails that were sensitive to prodding were considered alive.

### 2.6. Data evaluation

The experiment was laid out in completely randomized design. Analysis of variance (ANOVA) and Tukey's HSD (honestly significant difference) test were used to analyze the variation among groups and to determine the comparisons among groups, respectively. The differences among means were considered statistically significant at  $P < 0.05$ . The statistical analysis was carried out with SPSS (IBM Corp., Armonk, NY, USA).

## 3. Results

For the adult snails, variation in the number of dead snails was observed and was concentration-dependent. The % efficacy of wormwood EE was based on the percent mortality of the dead snails [13]. For 6.30% (T5) and 5.61% (T4), both treatments were highly effective with a mortality rate of 92% and 84%, respectively. This result was statistically comparable with niclosamide ( $P > 0.05$ ). On the other hand, 5% (T3) had a mortality rate of 63%, 4.46% (T2) a rate of 38%, and 3.98% (T1) a rate of 23%; these were considered ineffective and not comparable among treatments including niclosamide ( $P < 0.05$ ). Based on the percent efficacy of the European Agency for the Evaluation of Medicinal Products, treatments T1, T2, T3, and T4 were

considered not effective since the mortality rate was <90%, while T5 was considered effective as its mortality rate was >91%.

For juveniles, variation in the number of dead snails was also concentration-dependent. The efficacy of wormwood EE based on the percent mortality for 7.94% (T1) was 86%, for 8.91% (T2) was 90%, for 10% (T3) was 94%, for 11.22% (T4) was 95%, and for 12.59% (T5) was 98%, and these were all considered to be highly effective [13]. Based on the percent efficacy of the European Agency for the evaluation of Medicinal Products, T1 and T2 were considered not effective and not comparable with niclosamide ( $P < 0.05$ ). On the other hand, T3, T4, and T5 were considered effective as the mortality rates were >91%. Overall, T3, T4, and T5 are statistically comparable with commercially available molluscicide niclosamide ( $P > 0.05$ ) (Table 1).

Qualitative phytochemical tests were conducted in order to determine the possible bioactive components that might have contributed to the molluscicidal activity of the wormwood EE. Based on the phytochemical tests conducted, the secondary metabolites that were found to be positive were tannin, saponin, and terpenoids (Table 2). A condensed tannin was indicated with blue-black and brownish-green discoloration of the test solutions; for saponin, a copious lather formation indicates a positive result; a reddish-brown coloration of the test solution was positive for terpenoids. Mayer's test and Wagner's test for alkaloids both showed negative results, as did the alkaline test and ferric chloride test for flavonoids.

#### 4. Discussion

The wormwood EE exhibited potential molluscicidal activity against adult and juvenile *O. hupensis quadrasi* at

24 h of exposure. One of the bioactive components that is found in *Artemisia* spp. is vulgarone B, and this bioactive component, although not isolated and quantitatively determined in this study, is known to have molluscicidal activity [4,14]. Phytochemical tests of wormwood EE revealed the presence of tannins, saponins, and terpenoids. Previous studies indicated that these compounds were also present in concentrated extracts of other *Artemisia* spp. [15] and in different solvent extracts of *Artemisia dubia* [5,16]. The presence of these secondary metabolites as bioactive components that were identified during the qualitative phytochemical tests may be responsible for the molluscicidal activity of wormwood EE. These classes of compounds independently or in combination may be responsible for the mortality of snails [9,13].

It is well known that tannin-bearing plants have molluscicidal activity and these plants are generally avoided by mollusks [17,18]. Previous studies indicated that plants containing hydrolysable and condensed tannins exhibited strong molluscicidal activity against *Biomphalaria glabrata*, an intermediate host of *S. japonicum* [19]. In a different study, a bark extract of *Stryphnodendron polyphyllum* contained rich tannins and demonstrated promising molluscicide action against *Biomphalaria glabrata* [20]. Saponins have also been reported as potent molluscicides [20,21,22] with hemolytic properties and have toxic effects against cold-blooded animals including snails [23]. In addition, some effects of saponins on animal cells are the formation of a complex reaction with plasma and membrane cholesterol causing cell membrane damage [24]; the molluscicidal activity of terpenoids could be due to the reaction of the compound with thiol-containing enzymes [25,26]. Terpenoids can diffuse easily across cell membranes and induce biological reactions [27]. Most

**Table 1.** Mortality rates of wormwood leaf ethanolic extract against adult and juvenile *Oncomelania hupensis quadrasi* (mean  $\pm$  SEM).

Treatment	Mean no. of snails	Mean no. of dead adult snails	% Mortality	Mean no. of snails	Mean no. of dead juvenile snails	% Mortality
T0 <sub>(-1)</sub>	20	0 $\pm$ 0	0 <sup>a</sup> $\pm$ 0	20	0 $\pm$ 0	0 <sup>a</sup> $\pm$ 0
T0 <sub>(-2)</sub>	20	0 $\pm$ 0	0 <sup>a</sup> $\pm$ 0	20	0 $\pm$ 0	0 <sup>a</sup> $\pm$ 0
T0 <sub>(+)</sub>	20	20 $\pm$ 0	100 <sup>f</sup> $\pm$ 0	20	20 $\pm$ 0	100 <sup>d</sup> $\pm$ 0
T1	20	4.6 $\pm$ 0.55	23 <sup>b</sup> $\pm$ 2.74	20	17.2 $\pm$ 1.30	86 <sup>b</sup> $\pm$ 6.52
T2	20	7.6 $\pm$ 0.89	38 <sup>c</sup> $\pm$ 4.47	20	18 $\pm$ 1.22	90 <sup>bc</sup> $\pm$ 6.12
T3	20	12.6 $\pm$ 1.82	63 <sup>d</sup> $\pm$ 9.08	20	18.8 $\pm$ 1.0	94 <sup>bcd</sup> $\pm$ 6.52
T4	20	16.8 $\pm$ 0.84	84 <sup>e</sup> $\pm$ 4.14	20	19 $\pm$ 1.0	95 <sup>bcd</sup> $\pm$ 5.0
T5	20	18.4 $\pm$ 0.55	92 <sup>ef</sup> $\pm$ 2.74	20	19.6 $\pm$ 0.55	98 <sup>cd</sup> $\pm$ 2.74

Mean % mortalities within a column with different superscripts are significantly different ( $P < 0.05$ ). No. of replicates per treatment = 5; no. of snails per replicate = 20; N = 100.

**Table 2.** Secondary metabolites of wormwood leaves ethanolic extract.

Phytochemical	Wormwood ethanolic extract
Alkaloids	0
Tannins	++
Terpenoids	+++
Flavonoids	0
Saponins	++

+++ Strongly positive, ++ moderately positive, + weakly positive, - traces, 0 undetected [10].

likely their action involves the alkylation(s) of the essential enzymes of the snail's metabolism, resulting in death [28,29].

Two separate assays were performed for juvenile and adult snails. Juvenile snails are more susceptible and sensitive to molluscicides than adults [30]. Morphological, physiological, and biochemical characteristics may

## References

- Han B, Chen J, Yang X, Wang S, Li C et al. Molluscicidal activities of medicinal plants from eastern China against *Oncomelania hupensis*, the intermediate host of *Schistosoma japonicum*. *Revista Brasileira de Farmacognosia* 2010; 20 (5): 712-718. doi.org/10.1590/S0102-695X2010005000018
- Kloos H, McCullough FS. Plant molluscicides. *Planta Medica* 1982; 46 (4): 195-209. doi.org/10.1055/s-2007-971215
- Sun F, Zhang JF, Wen LY. Research progress on the molluscicidal effect of niclosamide compounded with other molluscicides against *Oncomelania hupensis*. *Chinese Journal of Parasitology & Parasitic Diseases* 2014; 32 (1): 72-75.
- Joshi RC, Meepagala KM, Sturtz G, Cagauan AG, Mendoza CO et al. Molluscicidal activity of vulgarone B from *Artemisia douglasiana* (Besser) against the invasive, alien, mollusc pest, *Pomacea canaliculata* (Lamarck). *International Journal of Pest Management* 2005; 51 (3): 175-180.
- Prabhakaran G, Bhore S, Ravichandran M. Development and evaluation of poly herbal molluscicidal extracts for control of apple snail (*Pomacea maculata*). *Agriculture* 2017; 7 (3): 22.
- Zeb S, Ali A, Zaman W, Zeb S, Ali S et al. Pharmacology, taxonomy and phytochemistry of the genus *Artemisia* specifically from Pakistan: a comprehensive review. *Pharmaceutical and Biomedical Research* 2018; 4 (4): 1-12. doi: 10.18502/pbr.v4i4.543
- Abid R, Qaiser M. Cypselas morphology and its taxonomic significance of the genus *Artemisia* L. (Anthemideae-Asteraceae) from Pakistan. *Pakistan Journal of Botany* 2008; 40 (5): 1827-1837.
- Fernandez TJ Jr, Landerito EO, Acabal AM. Development of the herbal drugs for the management of common stronglye worm infection in goats. In: Los Baños Laguna; 2009.
- Alinsub CJ, Bagot M. In vitro molluscicidal activity of wormwood (*Artemisia dubia*) leaves against *Oncomelania hupensis quadrasi*. *Annals of Tropical Research* 2019; 41: 16-23. doi: 10.32945/atr4112.2019
- Senguttuvan J, Paulsamy S, Karthika K. Phytochemical analysis and evaluation of leaf and root parts of the medicinal herb, *Hypochoeris radicata* L. for in vitro antioxidant activities. *Asian Pacific Journal of Tropical Biomedicine* 2014; 4: S359-S367. doi: 10.12980/APJTB.4.2014C1030
- Ponder WF, Hallan A, Shea M, Clark SA. *Australian Freshwater Mollusks*. Canberra, Australia: The Australian Museum; 2016.
- Chua JC, Tabios IB, Tamayo PP, Leonardo LR, Fontanilla IC et al. Genetic comparison of *Oncomelania hupensis quadrasi* (Möllendorf, 1895) (Gastropoda: Pomatiopsidae), the intermediate host of *Schistosoma japonicum* in the Philippines, based on 16S ribosomal RNA sequence. *Science Diliman* 2017; 29: 2.
- Kusin AA, Bagot MA, Lumin, JPLM. In vitro acaricidal efficacy of neem (*Azadirachta indica*) oil against ear mites (*Otodectes cynotis*). *Journal of Agriculture and Technology Management* 2016; 19: 15-19.
- Meepagala KM, Sturtz G, Mischke, CC, Wise D, Duke SO. Molluscicidal activity of vulgarone B against ram's horn snail (*Planorbella trivolvis*). *Pest Management Science* 2004; 60 (5): 479-482.

15. Sharma M, Devkota A. Allelopathic influences of *Artemisia Dubia* Wall. ex. Besser on seed germination and seedling vigor of *Parthenium hysterophorus* L. Journal of Institute of Science and Technology 2018; 22 (2): 117-128.
16. Sivagnanam SK, Rao RK, Mudiganti DU, Jeelani PG. Preliminary phytochemical analysis of *Artemisia amygdalina*, *Nerium odorum* and *Strychnos potatorum*. Journal of Pharmacy Research 2012; 5 (7): 3734-3739.
17. Hussein Ayoub SM, Yankov LK. The molluscicidal factor of tannin-bearing plants. International Journal of Crude Drug Research 1986; 24 (1): 16-18. doi: 10.3109/13880208609060881
18. Mølgaard P. Food plant preferences by slugs and snails: a simple method to evaluate the relative palatability of the food plants. Biochemical Systematics and Ecology 1986; 14 (1): 113-121. doi: 10.1016/0305-1978(86)90095-5
19. Schaufelberger D, Hostettmann K. On the molluscicidal activity of tannin containing plants. Planta Medica 1983; 48 (6): 105-107. doi: 10.1055/s-2007-969899
20. Bezerra JCB, Silva IA, Ferreira HD, Ferri PH, Santos SC. Molluscicidal activity against *Biomphalaria glabrata* of Brazilian Cerrado medicinal plants. Fitoterapia 2002; 73 (5): 428-430. doi: 10.1016/S0367-326X(02)00121-1
21. Musman M, Karina S, Rizki F. Saponins extract from *Barringtonia racemosa* as molluscicide to brackishwater pond snails (*Cerithidea cingulata*). International Journal of Applied Research and Technology 2014; 3 (6): 92-97.
22. Suter R, Tanner M, Borel C, Hostettmann K, Freyvogel TA. Laboratory and field trials at Ifakara (Kilombero District, Tanzania) on the plant molluscicide *Swartzia madagascariensis*. Acta Tropica 1986; 43 (1): 69-83.
23. Hostettmann K, Kizu H, Tomimori T. Molluscicidal properties of various saponins. Planta Medica 1982; 44 (1): 34-35.
24. Moses T, Papadopoulou KK, Osbourn A. Metabolic and functional diversity of saponins, biosynthetic intermediates and semi-synthetic derivatives. Critical Reviews in Biochemistry and Molecular Biology 2014; 49 (6): 439-462. doi: 10.3109/10409238.2014.953628
25. Hammami H, Mezghani-Jarraya R, Damak M, Ayadi A. Molluscicidal activity of various solvent extracts from *Solanum nigrum* var. *villosum* L. aerial parts against *Galba truncatula*. Parasite 2011; 18 (1): 63-70. doi: 10.1051/parasite/2011181063
26. Hmamouchi M, Lahlou M, Agoumi A. Molluscicidal activity of some Moroccan medicinal plants. Fitoterapia 2000; 71 (3): 308-314. doi: 10.1016/S0367-326X(99)00152-5
27. Srivastava AK, Singh VK. Biological action of essential oils (terpenes). International Journal of Biological and Medical Research 2019; 10 (3): 6854-6859.
28. Knobloch K, Pauli A, Iberl B, Weigand H, Weis N. Antibacterial and antifungal properties of essential oil components. Journal of Essential Oil Research 1989; 1 (3): 119-128. doi: 10.1080/10412905.1989.9697767
29. McGarvey DJ, Croteau R. Terpenoid metabolism. Plant Cell 1995; 7 (7): 1015.
30. Lemma A. Laboratory and field evaluation of the molluscicidal properties of *Phytolacca dodecandra*. Bulletin of the World Health Organization 1970; 42 (4): 597.
31. Mohammed A. Why are early life stages of aquatic organisms more sensitive to toxicants than adults? In: Gowder S (editor). New Insights into Toxicity and Drug Testing. Rijeka, Croatia: InTech; 2013. pp. 49-62. doi: 10.5772/55187
32. Rand GM. Fundamentals of Aquatic Toxicology: Effects, Environmental Fate and Risk Assessment. Boca Raton, FL, USA: CRC Press; 1995.
33. Pavarini DP, Pavarini SP, Niehues M, Lopes NP. Exogenous influences on plant secondary metabolite levels. Animal Feed Science and Technology 2012; 176 (1-4): 5-16.
34. Sampaio BL, Edrada-Ebel R, Da Costa FB. Effect of the environment on the secondary metabolic profile of *Tithonia diversifolia*: a model for environmental metabolomics of plants. Scientific Reports 2016; 2016: 6. doi: 10.1038/srep29265