

**Turkish Journal of Veterinary and Animal Sciences** 

http://journals.tubitak.gov.tr/veterinary/

Turk J Vet Anim Sci (2020) 44: 740-746 © TÜBİTAK doi:10.3906/vet-1907-50

## Comparative morphological analysis of diploid and triploid oysters, Crassostrea gigas, farmed in the Black Sea

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Received: 15.07.2019 • Accepted/Published Online: 28.01.2020	٠	Final Version: 02.06.2020
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Abstract: Triploid organisms, in particular oysters, are widely used in marine aquaculture. However, the advantage of triploid oysters compared to diploids has not been proven in all cases. Comparative morphological analysis of Pacific oysters Crassostrea gigas of different ploidy levels allows evaluating the benefits of raising triploids in the Black Sea. First, the morphological characteristics of diploid and triploid Pacific oysters C. gigas farmed in the Black Sea were studied. Allometric ratios of weight (W, g) and shell height (H, mm) of oysters were obtained, which are described by the equations  $W_{2n} = 4 \times 10^{-4} \times H_{2n}^{2.56}$ ,  $R^2 = 0.86$  and  $W_{3n} = 9 \times 10^{-5} \times H_{3n}^{2.90}$ ,  $R^2 = 0.86$ 0.91. Triploids showed positive allometry and had more biomass at smaller shell height than diploid oysters. A clear linear relationship of the height and length of oyster shells was established. Significant differences in linear parameters between 2n and 3n C. gigas farmed in Donuzlav Liman are not seen. Based on these results, it is assumed that the advantage of the growth of 3n mollusks is less evident in farms located at high latitudes. The environmental factors and farm technology have probably affected the growth of oysters to a greater degree than their ploidy.

Key words: Crassostrea gigas, triploids, allometric relationship, mariculture, Black Sea

### 1. Introduction

Commercial production of triploids (3n) began more than 30 years ago and became a prospect for industrial aquaculture. Triploid oysters can be produced by two general methods: chemical induction and crossing diploid (2n) mollusks with tetraploid (4n) ones. However, chemical treatment is not always reliable for producing 100% 3n populations, and crossing with tetraploid oysters is the most effective in producing triploids [1]. Nowadays, breeding lines of 4n oysters are produced in the USA, France, and Australia [2-5].

It is commonly recognized that triploids demonstrate faster growth rates, better meat conditions, partial sterility, and a higher survival rate due to resistance to disease. In general, triploids obtained by crossing  $2n \times 4n$  showed 20%–40% superior growth of shell height and 70%–80% increased whole weight compared to diploids [6,7]. Chemically induced 3n also possessed growth advantages compared to normal 2n oysters; however, this advantage was only 8% and 12%–31%, respectively [2,5].

In 3n mollusks, the growth rate, survival, and resistance to diseases may vary depending on species of oysters, cultivation conditions, environmental factors, and duration of the observation period. A number of studies demonstrated that larval stages of 3n and 2n did not differ by survival rate [6]. However, triploids of several oyster species became more vulnerable through the maturation period. For example, 3n Crassostrea virginica revealed higher mortality rates compared to 2n oysters [4]. Resistance to disease of polyploid oysters is also discussed. Some authors reported no influence of triploidy on survival to viral infections, such as resistance to OsHV-1, for C. gigas. Similarly, vibriosis caused the death of both diploids and triploids of C. gigas [8]. Indeed, the mortality rate of diploid oysters was greater during intensive gametogenesis, and for triploids, the peak of mortality was observed during winter months. On the other hand, the results of experiments on C. virginica and S. glomerata demonstrated a greater resistance to disease in triploids compared to diploids [6,9,10].

Abiotic factors (food supply, water temperature, and salinity) also influence the growth and survival rate of 3n oysters. Growth advantages of triploids are observed in waters of rich trophic growing areas, and in "poor" regions there are no differences in growth parameters between 2n and 3n oysters [6,11]. Geographical locations of farms also influence the cultivation of 3n oysters. The growth advantage of 3n mollusks was negligible (less than 7%)

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in farms situated at higher latitudes  $(42-43^{\circ}S)$ , such as in Tasmania, Australia [12]. In warmer regions  $(33-34^{\circ}N)$ , i.e. Japan [7] and Taiwan [13], the whole weight increase was up to 81% higher for 3n Pacific oysters compared to 2n during an 8-month observation period. The difference in growth rate between 2n and 3n *C. gigas* was maximal at 30 °C compared to similar oysters kept at 8–15 °C [14–16].

The main advantage of farming triploid oysters is the absence of the spawning period. The oysters maintain their product quality through the whole year. They are also called "four-season" oysters. 2n oysters undergoing a spawning period have shell growth retardation and decrease of whole weight and soft tissue weight, whereas triploids continue their growth [7], which causes differences not only in size but also in biochemical composition. Glycogen, carbohydrates, and total protein in triploids were 45%–65% higher compared to control diploids [17,18]. Low reproductive activity and additional reserve of carbohydrates and glycogen are supposed to undermine the high survival rate of 3n mollusks under unfavorable conditions.

Recent data confirm that farms currently prefer to cultivate two main species of triploid oysters, the Pacific oyster (*C. gigas*) and the eastern oyster (*C. virginica*) [5]. Higher growth rates of weight and shell length lead to a shorter vegetation period and to reaching the market size of triploids faster than diploids. In the late 1980s *C. gigas* was successfully introduced into the Black Sea. Natural conditions of the Black Sea allowed the development there of marine aquaculture of bivalves and oysters in particular [19,20]. For the first time, triploid oysters obtained in French nurseries appeared in Black Sea farms in 2005 [21].

In the present study, we investigated the main morphometrical parameters, i.e. shell height and length and whole weight, in 2n and 3n oysters, *Crassostrea gigas*, farmed in the estuary of Donuzlav, Black Sea (western coast of the Crimean peninsula). Specific attention has been paid to the ratio of these parameters and the scaling factor b in allometric equations.

### 2. Materials and methods

The objects of the research were diploid and triploid pacific oysters *C. gigas*, which were cultivated on a farm on the western coast of the Crimean peninsula at Donuzlav Liman (45°24'36"N, 33°9'3.2"E). Donuzlav Liman is a half-closed firth of the Black Sea, situated on the western coast of the Crimean peninsula. The modern ecosystem of the estuary was formed after its connection with the Black Sea through a canal built in 1961. The temperature regime is characterized by large annual amplitude with rapid summer warming and winter cooling to the bottom in the shallow areas. Maximal average month water temperature of 26.2 °C is observed during July and August, while the

minimal value of 5 °C is reached during February [22] (Figure 1). Water salinity is 17.2–18.1 ppt. Chlorophyll concentration ranged between 1.77 and 5.6 mg/m<sup>3</sup> [23].

Mollusks were held in plastic oyster cages with density of 350-450 animals/m<sup>2</sup>. The size of the oyster cage is 0.5 m  $\times$  1.00 m  $\times$  0.2 m, with mesh holes of 1.5 cm. Cages with oysters were landed at the shellfish farm and submerged 1.5-3 m from the surface of the sea. The observation period lasted from December 2015 to September 2016. Before measurements, shells were cleaned of fouling with a brush, washed with seawater, and dried with filter paper. Weighing of mollusks (defining W, g) was carried out on electronic scales with accuracy of 0.01 g. Linear parameters were measured by digital caliper with accuracy of 0.01 mm. The study was focused on the parameter of shell height, which is a criterion of marketability and cost for growing oysters. Shell height (H, mm) was estimated as a maximal distance between shell lock and growing edge. Shell length (L, mm) was measured at its widest part and was perpendicular to H [24] (Figure 2). Some authors define this parameter as shell length, or maximal linear size of the shell. This will be taken into account when discussing the results.

Based on the results obtained, the coefficients of variation, isometry equations of the relationship of the studied parameters  $L = m + b \times H$ , and allometry equations of mollusk weight ratio and linear dimensions have been computed:  $W = a \times H^b$  and  $W = a \times L^b$ , where W is oyster weight (g), H is shell height (mm), and L is shell length (mm), with coefficient of determination  $R^2$ . All data processing was done in MS Excel.

### 3. Results

# 3.1. Morphological parameters of diploid and triploid oysters

Variation series of shell height showed that the majority of 2n oysters possessed H 25–35 mm (16%) or 60–80 mm



Figure 1. Dynamic of temperature in the Donuzlav Liman.





Figure 2. Linear parameters of *C. gigas*.

(46.5%) (Figure 3). More than half of 3n oysters were characterized with shell height of 55–75 mm (55.4%).

Biometric parameters (shell height and length) and whole body weight of diploid and triploid oysters are shown in Table 1.

The obtained data demonstrate that variability of linear parameters was in the range of 0.32-0.35. For other species of oysters, for example C. corteziensis in the Gulf of California,  $CV_{\mu}$  may reach 0.38 and  $CV_{\mu}$  0.41 [25]. In our study, body weight of C. gigas varied widely (CV<sub>w</sub> 2n: 0.77;  $CV_w$  3n: 0.63), which was two times higher than the variability of linear parameters (Table 1). Some authors suggest that high coefficients of variation of morphological features may be caused by high heterozygosity of the organism [26]. In triploid mollusks obtained by crossing,  $\mathrm{CV}_{\mathrm{w}}$  and  $\mathrm{CV}_{\mathrm{H}}$  reached 0.85 and 0.55, respectively, which was significantly higher than in diploids [5]. CV is also influenced by various factors, i.e. genetic diversity of growing mollusks, environmental conditions, and methods for the selection of the objects studied and their further examination.

# 3.2. Allometric relationship of biometric parameters and whole weight of diploid and triploid oysters

Shell growth in bivalves is usually determined by the substrate quantity, population density (overpopulation),

Figure 3. Variational series of diploid and triploid oysters (C. gigas).

tidal and wave intensity, current speed, depth, feeding spectrum, etc. [24,27,28]. For cultivating oysters, additional factors are the method of growing (construction of oyster cages, their location in the sea, etc.) [29] and density of mollusks in cages [21,30]. In *C. gigas* shell shape possesses high individual plasticity due to disproportional and uneven increase of shell height and length. Shell length grows much slower than height [24,27]. In juvenile oysters up to 25 mm in size the flap height is equal to the length, and the shell has a rounded shape. As it grows, the shell of the Pacific oyster becomes first oval (in size group 35–55 mm), and then for larger mollusks the shell length is approximately 3/4 the height and shell shape becomes elongated (Figures 2 and 4).

The H-L relationship of the studied oysters is well described by linear equations  $L_{2n} = 0.58 \times H_{2n} + 0.95$ ,  $R^2 = 0.83$  and  $L_{3n} = 0.50 \times H_{3n} + 4.82$ ,  $R^2 = 0.72$ . Similar linear relations between shell height and length were obtained in other works: for *C. gigas*,  $L_{2n} = 0.96 \times H_{2n} - 1.01$ ; *C. commercialis*,  $L_{2n} = 0.65 \times H_{2n} - 0.11$  [24]; *C. virginica*,  $L_{2n} = 0.50 \times H_{2n} + 18.82$ ,  $L_{3n} = 0.48 \times H_{3n} + 22.65$ ; and *C. ariakensis*,  $L_{3n} = 0.54 \times H_{3n} + 33.35$  [31].

The ratio of linear dimensions (H/L) allows assessing shell shape. Disk-shaped and round specimens usually possess H/L close to 1, whereas an elongated shape of mollusks is associated with 1.5-2; H/L > 2 indicates long

		Diploids	Triploids
Number of specimens		485	484
Shell height, mm	M ± SD	61.17 ± 19.88	60.23 ± 17.38
	Min-max	23.0-99.1	23.0-98.9
	CV	0.32	0.29
Shell length, mm	M ± SD	36.53 ± 12.76	35.11 ± 10.27
	Min-max	14.0-67.4	14.0-64.6
	CV	0.35	0.29
Body weight, g	M ± SD	20.45 ± 15.73	19.50 ± 12.33
	Min-max	0.98-65.48	1.33-53.57
	CV	0.77	0.63

**Table 1.** Statistical analysis of the biometric and weight characteristics of cultivated *C. gigas*.

and narrow shells. In our study, the average ratio was 1.71–1.73, varying in the range of 1.21–2.48 for diploids and 1.14–2.6 for triploids. Thus, 3n oysters possessed higher variations of the ratio. More than 64% of mollusks were characterized by ellipsoidal shape; 12%–15% of specimens were elongated.

The relationship between shell height and whole body weight of living oysters was nonlinear but represented a power function (Table 2). The power coefficient b is a universal parameter for allometric equations, because its value does not depend on the type of mollusk weight used (wet, dry, or whole body weight) [28]. According to our data, for 2n C. gigas in the Donuzlav estuary b was 2.56, which was lower than for 3n oysters (2.90) (Table 2). The estimated value of b for bivalves varies in the range of 2.5-3 [28]. It has been determined that species of the family Ostreidae possess sagittally compressed shell shape, presuming a low power coefficient b in allometry equations of 1.5-2.5, or  $b \sim 2$ . For example, for C. columbiensis the estimated b is 2.35 [32]; for C. madrasensis, 1.7-2.0 [33] and 2.49-2.92 [34]; for C. virginica, 1.86 [35], 2.15 [36], 2.17 [1], and 2.26-2.39 [27]; for C. iridescens and C. angulata, 1.48 [35]; and for C. gigas, 1.12-2.79 [37], 1.87 [35], 2.39 [21], 2.43-2.52 [38,39], and 2.81 [40]. In a previous study, we also obtained low values of coefficient b for 2n and 3n C. gigas in the Blue Gulf (southern coast of Crimea) [21] (Table 2).

Triploid oysters from Donuzlav possessed positive allometry and had a greater weight with a smaller shell height compared to diploids. Similarly, in *C. virginica* triploids have greater *b*:  $W_{2n} = 4 \times 10^{-4} \times H^{1.82}$  and  $W_{3n} = 9 \times 10^{-5} \times H^{2.39}$  [41]. The value of the power coefficient



**Figure 4.** The ratio of the height and length of diploid and triploid oysters *C. gigas*.

**Table 2.** Parameters of allometry equation  $W = a \times H^b$  and  $W = a \times L^b$  of diploid and triploid oysters *C. gigas.* 

Area	Oysters' ploidy	а	Ь	R <sup>2</sup>					
$W = a \times H^b$									
Donuzlav Liman	Diploids	$4 \times 10^{-4}$	2.56	0.86	Present data				
	Triploids	$9 \times 10^{-5}$	2.90	0.91	Present data				
Blue Gulf	Diploids	$8 \times 10^{-4}$	2.34	0.94	[21]				
	Triploids	$9 \times 10^{-4}$	2.18	0.95	[21]				
$W = a \times L^b$									
Donuzlav Liman	Diploids	$9 \times 10^{-4}$	2.7	0.81	Present data				
	Triploids	$1.8 \times 10^{-3}$	2.56	0.82	Present data				

may vary depending on environmental conditions [28]. It was demonstrated that water temperature, chlorophyll concentration, tidal currents speed, and cultivation method used influence the ratio of shell height to weight in *C. virginica* [42]. Previously, we obtained the opposite results for oysters in the open coastal area of the Black Sea (the Blue Gulf) (Table 2), which may be caused by the peculiarities of the farm's location.

### 4. Discussion

The morphology of the C. gigas shell is quite plastic; therefore, investigating the shell's morphological parameters allows assessing the conditions of bivalves at various stages of growth. The main criteria for commercial oysters are their size and weight. Controlling the formation of shell and biomass allows obtaining quality seafood. Shell shape depends on environmental factors (temperature, salinity) and habitat conditions (population density, depth, substrate type, etc.) [29,31]. The height to length ratio, in turn, influences oysters' shell surface area, internal volume, and soft tissue weight. Several authors demonstrated that the H-L ratio in C. gigas may be linear or nonlinear, which for the most part depends on habitat conditions or cultivation methods [24]. It is known that the H-L ratio was greater in mollusks inhabiting soft substrates compared to hard substrates. A high density of oyster populations also substantially influenced shell shape and increased H-L values [31].

Notably, not all the studies published demonstrated growth advantages of triploid oysters. In areas with unfavorable environments or intensive marine culturing the advantage of triploids was negligible or was not observed. Previous studies have shown that triploids respond to environmental conditions (low salinity, high water temperature) differently than diploids. Callam et al. [43] showed that at low salinity (6-13 ppt) linear and weight growth of diploid and triploid C. virginica did not substantially differ. It was reported that 4% slower shell growth was seen in triploid C. gigas but there was a more rapid increase of body weight at water temperatures of 17.2-26.5 °C compared to diploids [18]. Besides environmental factors, the cultivation method also influences the conditions of diploids and triploids. Indeed, intensive technological manipulation during the growing period, removing fouling organisms from oyster cages, and mollusk sorting may reduce the differences in size and weight in diploid and triploid mollusks [44]. Construction of oyster carriers also influences the growth of bivalves. Rapid increase of soft tissue weight was observed in 3n oysters growing in plastic cages or cages submerged at defined depths on rope lines compared to oysters raised in mesh bags [29].

Thus, peculiarities of growth of 3n and 2n mollusks are probably determined by a complex influence and/or

#### References

Guo X, Allen SK. Viable tetraploid Pacific oyster (*Crassostrea gigas* Thunburg) produced by inhibiting polar body I in eggs of triploids. Molecular Marine Biology and Biotechnology 1994; 3 (1): 42-50.

interaction of abiotic factors (salinity, water temperature, currents, tides, diversity and amount of phytoplankton food, etc.) and biotic components (species, age, maturation stage, the origin of oysters, etc.). The advantage of triploids becomes most noticeable when oysters reach a certain size. Hand et al. [45] reported significant differences between 2n and 3n Sydney rock oysters *S. commercialis* after they reached weights of 5–10 g or shell heights of 20–40 mm, which corresponds to initiation of gametogenesis and spawning in diploid mollusks. In eastern oyster, *C. virginica*, farmed in the Chesapeake Bay, a similar phenomenon was observed several months after transplanting the mollusks in the sea, starting with size <25 mm or weight <10 g [45].

In the present work, we did not observe significant differences in linear parameters between polyploid and diploid mollusks farmed at Donuzlav (Table 1, Figure 4). Obtained results confirmed the assumption that the advantage of the growth of 3n mollusks is less evident in farms located at high latitudes, the coordinates of the Donuzlav estuary being 45°24′36″N, 33°9′3.2″E. Environmental conditions probably influenced the growth of oysters to a greater extent than their ploidy. The absence of expected advantages in the growth of 3n oysters compared to 2n ones may be caused by the method of culturing in the suspended cages and seasonal wave events in the area, which can damage the growing edge of the shell and reduce the linear dimensions of mollusks.

Comparative analysis of the height-to-length ratio in diploid and triploid *C. gigas* revealed substantial differences depending on the area of cultivation. According to the value of power coefficient *b*, the weight of oysters from the Donuzlav estuary was greater than that for similarly sized mollusks in the Blue Gulf. It is quite difficult to predict the mass of an oyster by its height, since the mass is more variable than shell size. Mollusk weight depends on various factors, including the quality and availability of phytoplankton, filtration rate, maturation stage, age, and season. It is also known that in bivalves the proportion of the shell itself in the total raw weight gradually increases due to age-related shell thickening [38,39].

#### Acknowledgment

These studies were supported by the Ministry of Science and Higher Education of the Russian Federation, Grant № 0828-2018-0003.

 Guo X, DeBrosse GA, Allen SK. All-triploid Pacific oysters (*Crassostrea gigas*, Thunberg) produced by mating tetraploids and diploids. Aquaculture 1996; 142 (3-4): 149-161.

- Nell JA. Farming triploid oysters. Aquaculture 2002; 210: 69-88. doi: 10.1016/S0044-8486(01)00861-4
- 4. Wadsworth PC. Comparing triploid and diploid growth and mortality in farmed oysters, *Crassostrea virginica*, in the northern Gulf of Mexico. MSc, Auburn University, Auburn, AL, USA, 2018.
- Wadsworth P, Wilson AE, Walton WC. A meta-analysis of growth rate in diploid and triploid oysters. Aquaculture 2019; 499: 9-16. doi: 10.1016/j.aquaculture.2018.09.018
- 6. Guo X, Wang Y, Debrosse GA, Bushek D, Ford SE. Building a superior oyster for aquaculture. Jersey Shoreline 2008; 25: 7-9.
- Akashige S, Fushimi T. Growth, survival, and glycogen content of triploid Pacific oyster *Crassostrea gigas* in the waters of Hiroshima, Japan. Nippon Suisan Gakkaishi 1992; 58: 1063-1071.
- De Decker S, Normand J, Saulnier D, Pernet F, Castagnet S et al. Responses of diploid and triploid Pacific oysters *Crassostrea gigas* to *Vibrio* infection in relation to their reproductive status. Journal of Invertebrate Pathology 2011; 106 (2): 179-191. doi: 10.1016/j.jip.2010.09.003
- Matthiessen GC, Davis JP. Observations on growth rate and resistance to MSX *Haplosporidium nelsoni* among diploid and triploid Eastern oysters *Crassostrea virginica* (Gmelin, 1797) in New England. Journal of Shellfish Research 1992; 11: 449-454.
- Nell JA, Hand RE. Evaluation of the progeny of secondgeneration Sydney rock oyster *Saccostrea glomerata* (Gould, 1850) breeding lines for resistance to QX disease *Marteilia sydneyi*. Aquaculture 2003; 228: 27-35. doi: 10.1016/S0044-8486(03)00133-9
- Garnier-Gèrè PH, Naciri-Graven Y, Bougrier S, Magoulas A, Hèral M et al. Influences of triploidy, parentage and genetic diversity on growth of the Pacific oyster *Crassostrea gigas* reared in contrasting natural environments. Molecular Ecology 2002; 11 (8): 1499-1514. doi: 10.1046/j.1365-294X.2002.01531.x
- 12. Maguire GB, Boocock B, Kent GN, Gardner NC. Studies on triploids in Australia: IV. Sensory evaluation of triploid and diploid Pacific oysters, *Crassostrea gigas* (Thunberg), in Tasmania. In: Nell JA, Maguire GB (editors). Evaluation of Triploid Sydney Rock Oysters (*Saccostrea commercialis*) and Pacific Oysters (*Crassostreagigas*) on Commercial Leases in New South Wales and Tasmania. Hobart, Australia: University of Tasmania; 1994. pp. 178-193.
- Chao NH, Tsai HP, Liang CI, Cheng HY, Cheng JH et al. Induction and performance of triploid oysters, *Crassostrea gigas*, in Taiwan COA (Council of Agriculture, Taiwan). Fisheries Series 1999; 65: 99-115.
- Brake J, Davidson J, Davis J. Field observations on growth, gametogenesis, and sex ratio of triploid and diploid *Mytilus edulis*. Aquaculture 2004; 236: 179-191. doi: 10.1016/j. aquaculture.2003.09.016
- Davis JP. Growth rate of sibling diploids and triploid oysters *Crassostrea gigas.* Journal of Shellfish Research 1988; 8: 319-325.

- Shpigel M, Barber BJ, Mann R. Effects of elevated temperature on growth, gametogenesis, physiology, and biochemical composition in diploid and triploid Pacific oysters, *Crassostrea gigas* (Thunberg). Journal of Experimental Marine Biology and Ecology 1992; 161 (1): 15-25.
- Wu X, Zhang Y, Xiao S, Qin Y, Ma H et al. Comparative studies of the growth, survival, and reproduction of diploid and triploid Kumamoto oyster, *Crassostrea sikamea*. Journal of the World Aquaculture Society 2019; 1: 1-12. doi: 10.1111/ jwas.12596
- Ibarra AM, Ascencio-Michel R, Ramírez JL, Manzano-Sarabia M, Rodríguez-Jaramillo C. Performance of diploid and triploid *Crassostrea gigas* (Thunberg, 1793) grown in tropical versus temperate natural environmental conditions. Journal of Shellfish Research 2017; 36: 119-139. doi: 10.2983/035.036.0113
- Zolotnitzkij AP, Orlenko AN, Krjuchkov VG, Sytnik NA. On organization of large scale culture of oysters in the Donuzlav Lake. YugNIRO Proceedings 2008; 46: 48-54 (in Russian).
- Krjuchkov VG. Experience of growing oysters off the east coast of the Black Sea. YugNIRO Proceedings 2010; 48: 29-35 (in Russian).
- 21. Vyalova OYu. The first results of cultivation of triploids pacific oysters *Crassostreagigas* in the Black sea (Southern coast of Crimea). Ekologiya Morya 2009; 79: 37-43 (in Russian).
- 22. Nemirovsky MS, Kovrigina NP. Dynamics of the lake Donuzlav waters. Ecology of Sea 2000; 51: 10-13 (in Russian).
- 23. Grebneva EA, Polonsky AB, Serebrennikov AN. Hydrological characteristics of the waters adjacent to the west coast of Crimea, according to research expeditions to the HS "Donuzlav" in June 2016. Monitoring Systems of Environment 2016; 6 (26): 68-73 (in Russian).
- 24. Nair NU, Nair NB. Height-length relation of shells in the Indian backwater oyster *Crassostrea madrasensis* (Preston) of the Cochi Harbour. Fishery Technology 1986; 23: 27-31.
- 25. Góngora-Gómez AM, Leal-Sepúlveda AL, García-Ulloa M, Aragón-Noriega EA, Valenzuela-Quiñónez W. Morphometric relationships and growth models for the oyster *Crassostrea corteziensis* cultivated at the southeastern coast of the Gulf of California, Mexico. Latin American Journal of Aquatic Research 2018; 46 (4): 735-743. doi: 10.3856/vol46-issue4fulltext-11
- Higgins JPT, Thompson SG. Quantifying heterogeneity in a meta-analysis. Statistics in Medicine 2002; 21 (11): 1539-1558. doi: 10.1002/sim.1186
- 27. Dame RF. Comparison of various allometric relationships in intertidal and subtidal American oysters. Fishery Bulletin 1972; 70 (4): 1121-1126.
- Powell EN, Mann R, Ashton-Alcox KA, Kim Y, Bushek D. The allometry of oysters: spatial and temporal variation in the length-biomass relationships for *Crassostrea virginica*. Journal of the Marine Biological Association of the United Kingdom 2015; 96 (5): 1-18. doi: 10.1017/S0025315415000703

- 29. Walton WC, Rikard FS, Chaplin GI, Davis JE, Arias CR et al. Effects of ploidy and gear on the performance of cultured oysters, *Crassostrea virginica*: survival, growth, shape, condition index and *Vibrio* abundances. Aquaculture 2013; 414-415: 260-266. doi: 10.1016/j.aquaculture.2013.07.032
- Guo X. Use and exchange of genetic resources in molluscan aquaculture. Reviews in Aquaculture 2009; 1 (3-4): 251-259. doi: 10.1111/j.1753-5131.2009.01014.x
- Harding JM. Comparison of growth rates between diploid DEBY eastern oysters (*Crassostrea virginica*, Gmelin 1791), triploid eastern oysters, and triploid Suminoe oysters (*C. ariakensis*, Fugita 1913). Journal of Shellfish Research 2007; 26 (4): 961-972. doi: 10.2983/0730-8000(2007)26[961:COGRBD] 2.0.CO;2
- Cruz AFC, Pena JC, Lopez YS. Growth and sexual maturity of a *Crassostrea columbiensis* (Mollusca: Bivalvia) population. Revista de Biologia Tropical 1997; 45: 335-339.
- Nagi HM, Shenai-Tirodkar PS, Jagtap TG. Dimensional relationships in *Crassostrea madrasensis* (Preston) and *C.* gryphoides (Schlotheim) in Mangrove ecosystem. Indian Journal of Geo-Marine Sciences 2011; 40 (4): 559-566.
- Nurul Amin SM, Zafar M, Halim A. Age, growth, mortality and population structure of the eastern oyster, *Crassostrea madrasensis*, in the Mahaskhali Channel (southeastern coast of Bangladesh). Journal of Applied Ichthyology 2008; 24 (1): 18-25. doi: 10.1111/j.1439-0426.2007.01007.x
- Octavina C, Yulianda F, Krisanti M, Muchlisin ZA. Lengthweight relationship of Ostreidae in the Kuala Gigieng estuary, Aceh Besar District, Indonesia. AACL Bioflux 2015; 8 (5): 817-823.
- Mann R, Southworth M, Harding JM, Wesson JA. Population studies of the native eastern oyster, *Crassostrea virginica* (Gmelin, 1791) in the James River, Virginia, USA. Journal of Shellfish Research 2009; 28 (2): 193-220. doi: 10.2983/035.028.0203

- Yoo SK, Yoo MS. Biological studies on oyster culture (II) morphological characteristics of the oyster, *Crassostrea gigas*. Bulletin of Korean Fishery Society 1973; 6: 65-74.
- 38. Zolotnitzkij AP, Monina OB. Growth and production of Japan oyster (*Crassostrea gigas* Thunberg), acclimated in the Black sea. Ecology of Sea 1992; 41: 77-80 (in Russian).
- Zolotnitzkij AP, Orlenko AN. Ecological patterns of growth of pacific oysters in various areas of the Black Sea. Fishery of Ukraine 1999; 2: 37-39 (in Russian).
- Diederich S, Nehls G, Van Beusekom JEE, Reise K. Introduced Pacific oysters (*Crassostreagigas*) in the northern Wadden Sea: invasion accelerated by warm summers? Helgoland Marine Research 2005; 59 (2): 97-106. doi: 10.1007/s10152-004-0195-1
- Dégremont L, Garcia C, Frank-Lawale A, Allen SK. Triploid oysters in Chesapeake: Comparison of diploid and triploid *Crassostrea virginica*. Journal of Shellfish Research 2012; 31 (1): 21-31. doi: 10.2983/035.031.0103
- 42. Grizzle RE, Ward KM, Peter CR, Cantwell M, Katz D et al. Growth, morphometrics, and nutrient content of farmed eastern oysters, *Crassostrea virginica* (Gmelin), in New Hampshire, USA. Aquaculture Research 2017; 48 (4): 1525-1537. doi: 10.1111/are.12988
- Callam BR, Allen SK, Frank-Lawale A. Genetic and environmental influence on triploid *Crassostrea virginica* grown in Chesapeake Bay: growth. Aquaculture 2016; 452: 97-106. doi: 10.1016/j.aquaculture.2015.10.027
- 44. Stone BW, Hadley NH, Kingsley-Smith PR. Evaluating the potential growth advantage of triploid Eastern oysters (*Crassostrea virginica*) in South Carolina relative to commercially cultured diploid native stocks. Journal of Shellfish Research 2013; 32 (3): 647-655. doi: 10.2983/035.032.0304
- 45. Hand RE, Nell JA, Maguire GB. Studies on triploid oysters in Australia. X. Growth and mortality of diploid and triploid Sydney rock oysters *Saccostrea commercialis* (Iredale and Roughley). Journal of Shellfish Research 1998; 17 (4-5): 1115-1127.